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(54) Title: 37 STAPHYLOCOCCUS AUREUS GENES AND POLYPEPTIDES

(57) Abstract: The present invention relates to novel genes from *S. aureus* and the polypeptides they encode. Also provided are vectors, host cells, antibodies and recombinant methods for producing the same. The invention further relates to screening methods for identifying agonists and antagonists of *S. aureus* polypeptide activity. The invention additionally relates to diagnostic methods for detecting *Staphylococcus* nucleic acids, polypeptides and antibodies in a biological sample. The present invention further relates to novel vaccines for the prevention or attenuation of infection by *Staphylococcus*.

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37 *Staphylococcus aureus* genes and polypeptides

Field of the Invention

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The present invention relates to novel *Staphylococcus aureus* genes (*S. aureus*) nucleic acids and polypeptides. Also provided are vectors, host cells and recombinant or synthetic methods for producing the same. Further provided are diagnostic methods for detecting *S. aureus* using probes, primers, and antibodies to the *S. aureus* nucleic acids and polypeptides of the present invention. The invention further relates to screening methods for identifying agonists and antagonists of *S. aureus* polypeptide activity and to vaccines using *S. aureus* nucleic acids and polypeptides and to therapeutics using agonists and/or antagonists of the invention.

15 Background of the Invention

The genus *Staphylococcus* includes at least 20 distinct species. (For a review see Novick, R. P., *The Staphylococcus as a Molecular Genetic System* in MOLECULAR BIOLOGY OF THE STAPHYLOCOCCI, 1-37 (R. Novick, Ed., VCH Publishers, New York (1990)). Species differ from one another by 80% or more, by hybridization kinetics, whereas strains within a species are at least 90% identical by the same measure.

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The species *S. aureus*, a gram-positive, facultatively aerobic, clump-forming cocci, is among the most important etiological agents of bacterial infection in humans, as discussed briefly below.

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Human Health and S. aureus

Staphylococcus aureus is a ubiquitous pathogen. See, e.g., Mims et al., MEDICAL MICROBIOLOGY (Mosby-Year Book Europe Limited, London, UK 1993). It is an etiological agent of a variety of conditions, ranging in severity from mild to fatal. A few of the more common conditions caused by *S. aureus* infection are burns, cellulitis, eyelid infections, food poisoning, joint infections, neonatal conjunctivitis, osteomyelitis, skin infections, surgical wound infection, scalded skin syndrome and toxic shock syndrome, some

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of which are described further below.

Burns: Burn wounds generally are sterile initially. However, they generally compromise physical and immune barriers to infection, cause loss of fluid and electrolytes and result in local or general physiological dysfunction. After cooling, contact with viable bacteria results in mixed colonization at the injury site. Infection may be restricted to the non-viable debris on the burn surface ("eschar"), it may progress into full skin infection and invade viable tissue below the eschar and it may reach below the skin, enter the lymphatic and blood circulation and develop into septicemia. *S. aureus* is among the most important pathogens typically found in burn wound infections. It can destroy granulation tissue and produce severe septicemia.

Cellulitis: Cellulitis, an acute infection of the skin that expands from a typically superficial origin to spread below the cutaneous layer, most commonly is caused by *S. aureus* in conjunction with *S. pyogenes*. Cellulitis can lead to systemic infection. In fact, cellulitis can be one aspect of synergistic bacterial gangrene. This condition typically is caused by a mixture of *S. aureus* and microaerophilic *Streptococci*. It causes necrosis and treatment is limited to excision of the necrotic tissue. The condition often is fatal.

Eyelid infections: *S. aureus* is the cause of styes and of "sticky eye" in neonates, among other eye infections. Typically such infections are limited to the surface of the eye, and may occasionally penetrate the surface with more severe consequences.

Food poisoning: Some strains of *S. aureus* produce one or more of five serologically distinct, heat and acid stable enterotoxins that are not destroyed by digestive process of the stomach and small intestine (enterotoxins A-E). Ingestion of the toxin, in sufficient quantities, typically results in severe vomiting, but not diarrhea. The effect does not require viable bacteria. Although the toxins are known, their mechanism of action is not understood.

Joint infections: *S. aureus* infects bone joints causing diseases such osteomyelitis. See, e.g., R. Cunningham et al., (1996) J. Med. Microbiol. 44:157-164.

Osteomyelitis: *S. aureus* is the most common causative agent of haematogenous osteomyelitis. The disease tends to occur in children and adolescents more than adults and it is associated with non-penetrating injuries to bones. Infection typically occurs in the long end of growing bone, hence its occurrence in physically immature populations. Most often, infection is localized in the vicinity of sprouting capillary loops adjacent to epiphysis growth plates in the end of long, growing bones.

Skin infections: *S. aureus* is the most common pathogen of such minor skin infections as abscesses and boils. Such infections often are resolved by normal host response mechanisms, but they also can develop into severe internal infections. Recurrent infections of the nasal passages plague nasal carriers of *S. aureus*.

5 *Surgical Wound Infections:* Surgical wounds often penetrate far into the body. Infection of such wound thus poses a grave risk to the patient. *S. aureus* is the most important causative agent of infections in surgical wounds. *S. aureus* is unusually adept at invading surgical wounds; sutured wounds can be infected by far fewer *S. aureus* cells than are necessary to cause infection in normal skin. Invasion of surgical wound can lead to severe *S. aureus* 10 septicemia. Invasion of the blood stream by *S. aureus* can lead to seeding and infection of internal organs, particularly heart valves and bone, causing systemic diseases, such as endocarditis and osteomyelitis.

Scalded Skin Syndrome: *S. aureus* is responsible for "scalded skin syndrome" (also called toxic epidermal necrosis, Ritter's disease and Lyell's disease). This disease occurs in older 15 children, typically in outbreaks caused by flowering of *S. aureus* strains produce exfoliation (also called scalded skin syndrome toxin). Although the bacteria initially may infect only a minor lesion, the toxin destroys intercellular connections, spreads epidermal layers and allows the infection to penetrate the outer layer of the skin, producing the desquamation that typifies the disease. Shedding of the outer layer of skin generally reveals 20 normal skin below, but fluid lost in the process can produce severe injury in young children if it is not treated properly.

Toxic Shock Syndrome: Toxic shock syndrome is caused by strains of *S. aureus* that produce the so-called toxic shock syndrome toxin. The disease can be caused by *S. aureus* infection at any site, but it is too often erroneously viewed exclusively as a disease solely of women 25 who use tampons. The disease involves toxemia and septicemia, and can be fatal.

Nocosomal Infections: In the 1984 National Nosocomial Infection Surveillance Study ("NNIS") *S. aureus* was the most prevalent agent of surgical wound infections in many hospital services, including medicine, surgery, obstetrics, pediatrics and newborns.

Other Infections: Other types of infections, risk factors, etc. involving *S. aureus* are 30 discussed in: A. Trilla (1995) *J. Chemotherapy* 3:37-43; F. Espersen (1995) *J. Chemotherapy* 3:11-17; D.E. Craven (1995) *J. Chemotherapy* 3:19-28; J.D. Breen et al. (1995) *Infect. Dis. Clin. North Am.* 9(1):11-24 (each incorporated herein in their entireties).

Resistance to drugs of S. aureus strains

Prior to the introduction of penicillin the prognosis for patients seriously infected with *S. aureus* was unfavorable. Following the introduction of penicillin in the early 1940s even the worst *S. aureus* infections generally could be treated successfully. The emergence of penicillin-resistant strains of *S. aureus* did not take long, however. Most strains of *S. aureus* encountered in hospital infections today do not respond to penicillin; although, fortunately, this is not the case for *S. aureus* encountered in community infections. It is well known now that penicillin-resistant strains of *S. aureus* produce a lactamase which converts penicillin to penicillinoic acid, and thereby destroys antibiotic activity. Furthermore, the lactamase gene often is propagated episomally, typically on a plasmid, and often is only one of several genes on an episomal element that, together, confer multidrug resistance.

Methicillins, introduced in the 1960s, largely overcame the problem of penicillin resistance in *S. aureus*. These compounds conserve the portions of penicillin responsible for antibiotic activity and modify or alter other portions that make penicillin a good substrate for inactivating lactamases. However, methicillin resistance has emerged in *S. aureus*, along with resistance to many other antibiotics effective against this organism, including aminoglycosides, tetracycline, chloramphenicol, macrolides and lincosamides. In fact, methicillin-resistant strains of *S. aureus* generally are multiply drug resistant.

Methicillin-resistant *S. aureus* (MRSA) has become one of the most important nosocomial pathogens worldwide and poses serious infection control problems. Today, many strains are multiresistant against virtually all antibiotics with the exception of vancomycin-type glycopeptide antibiotics.

Recent reports that transfer of vancomycin resistance genes from enterococci to *S. aureus* has been observed in the laboratory sustain the fear that MRSA might become resistant against vancomycin, too; a situation generally considered to result in a public health disaster. MRSA owe their resistance against virtually all β -lactam antibiotics to the expression of an extra penicillin binding protein (PBP) 2a, encoded by the *mecA* gene. This additional very low affinity PBP, which is found exclusively in resistant strains, appears to be the only pbp still functioning in cell wall peptidoglycan synthesis at β -lactam concentrations high enough to saturate the normal set of *S. aureus* PBP 1-4. In 1983 it was shown by insertion mutagenesis using transposon Tn551 that several additional genes independent of

mecA are needed to sustain the high level of methicillin resistance of MRSA. Interruption of these genes did not influence the resistance level by interfering with PBP2a expression, and were therefore called *fem* (factor essential for expression of methicillin resistance) or *aux* (auxiliary genes).

5 In the meantime six *fem* genes (*femA*- through F) have been described and the minimal number of additional *aux* genes has been estimated to be more than 10. Interference with *femA* and *femB* results in a strong reduction of methicillin resistance, back to sensitivity of strains without PBP2a. The *fem* genes are involved in specific steps of cell wall synthesis. Consequently, inactivation of *fem* encoded factors induce β -lactam hypersensitivity in already
10 sensitive strains. Both *femA* and *femB* have been shown to be involved in peptidoglycan pentaglycine interpeptide bridge formation. FemA is responsible for the formation of glycines 2 and 3, and FemB is responsible for formation of glycines 4 and 5. *S. aureus* may be involved in the formation of a monoglycine mucopeptide precursors. FemC-F influence amidation of the iso-D-glutamic acid residue of the peptidoglycan stem peptide, formation of
15 a minor mucopeptide with L-alanine instead of glycine at position 1 of the interpeptide bridge, perform a yet unknown function, or are involved in an early step of peptidoglycan precursors biosynthesis (addition of L-lysine), respectively.

Summary of the Invention

20 The present invention provides isolated *S. aureus* polynucleotides and polypeptides shown in Table 1 and SEQ ID NO:1 through SEQ ID NO:74. Polynucleotide sequences are shown as the odd numbered SEQ ID NOs (e.g., SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, and so on up to SEQ ID NO:73). The polypeptide sequences are shown as the even numbered SEQ ID NOs (e.g., SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, and so on up to SEQ ID
25 NO:74). One aspect of the invention provides isolated nucleic acid molecules comprising or alternatively, consisting of, polynucleotides having a nucleotide sequence selected from the group consisting of: (a) a nucleotide sequence shown in Table 1; (b) a nucleotide sequence encoding any of the amino acid sequences of the polypeptides shown in Table 1; (c) a nucleotide sequence encoding an antigenic fragment of any of the polypeptides shown in
30 Table 1; (d) a nucleotide sequence encoding a biologically active fragment of any of the polypeptides shown in Table 1; and (e) a nucleotide sequence complementary to any of the nucleotide sequences in (a), (b), (c) and/or (d). The invention further provides for fragments

of the nucleic acid molecules of (a), (b), (c), (d) and/or (e) above.

Further embodiments of the invention include isolated nucleic acid molecules that comprise or alternatively, consist of, a polynucleotide having a nucleotide sequence at least 90% identical, and more preferably at least 95%, 96%, 97%, 98%, 99% or 100% identical, to any of the nucleotide sequences in (a), (b), (c), (d), or (e) above, or a polynucleotide which hybridizes under stringent hybridization conditions to a polynucleotide in (a), (b), (c), (d) or (e) above. Additional nucleic acid embodiments of the invention relate to isolated nucleic acid molecules comprising polynucleotides which encode the amino acid sequences of epitope-bearing portions of a *S. aureus* polypeptide having an amino acid sequence in Table 1, and including but not limited to those epitope-bearing portions shown in Table 4.

The present invention also relates to recombinant vectors, which include the isolated nucleic acid molecules of the present invention, and to host cells containing the recombinant vectors, as well as to methods of making such vectors and host cells. The present invention further relates to the use of these vectors in the production of *S. aureus* polypeptides or peptides by recombinant techniques.

The invention further provides isolated *S. aureus* polypeptides having an amino acid sequence selected from the group consisting of an amino acid sequence described in (a), (b), (c), (d), or (e) above, any of the polypeptides described in Table 1 or the complement thereof, and/or fragments thereof.

The polypeptides of the present invention also include polypeptides having an amino acid sequence with at least 70% similarity, and more preferably at least 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99%, or 100% similarity to those described in Table 1, as well as polypeptides having an amino acid sequence at least 70% identical, more preferably at least 75% identical, and still more preferably 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical to those above; as well as isolated nucleic acid molecules encoding such polypeptides.

The present invention provides antagonists of the polypeptides of the invention (e.g., including but not limited to antibodies to the polypeptides of the invention, small molecule inhibitors of the polypeptides of the invention) as therapeutic treatment in a *S. aureus* mediated disease.

The present invention further provides a vaccine, preferably a multi-component vaccine comprising one or more of the *S. aureus* polynucleotides or polypeptides described in

Table 1, or fragments thereof, together with a pharmaceutically acceptable diluent, carrier, or excipient, wherein the *S. aureus* polypeptide(s) are present in an amount effective to elicit an immune response to members of the *Staphylococcus* genus, or at least *S. aureus*, in an animal. The *S. aureus* polypeptides of the present invention may further be combined with

5 one or more immunogens of one or more other staphylococcal or non-staphylococcal organisms to produce a multi-component vaccine intended to elicit an immunological response against members of the *Staphylococcus* genus and, optionally, one or more non-staphylococcal organisms.

10 The vaccines of the present invention can be administered in a DNA form, e.g., "naked" DNA, wherein the DNA encodes one or more staphylococcal polypeptides and, optionally, one or more polypeptides of a non-staphylococcal organism. The DNA encoding one or more polypeptides may be constructed such that these polypeptides are expressed as fusion proteins.

15 The vaccines of the present invention may also be administered as a component of a genetically engineered organism or host cell. Thus, a genetically engineered organism or host cell which expresses one or more *S. aureus* polypeptides may be administered to an animal. For example, such a genetically engineered organism or host cell may contain one or more *S. aureus* polypeptides of the present invention intracellularly, on its cell surface, or in its periplasmic space. Further, such a genetically engineered organism or host cell may
20 secrete one or more *S. aureus* polypeptides. The vaccines of the present invention may also be co-administered to an animal with an immune system modulator (e.g., CD86 and GM-CSF).

The invention also provides a method of inducing an immunological response in an animal to one or more members of the *Staphylococcus* genus, preferably one or more isolates
25 of the *S. aureus* species, comprising administering to the animal a vaccine as described above.

The invention further provides a method of inducing a protective immune response in an animal, sufficient to prevent, attenuate, or control an infection by members of the *Staphylococcus* genus, preferably at least *S. aureus* species, comprising administering to the
30 animal a composition comprising one or more of the polynucleotides or polypeptides described in Table 1, or fragments thereof (e.g., including, but not limited to, fragments which comprise the epitopes shown in Table 4). Further, these polypeptides, or fragments

thereof, may be conjugated to another immunogen and/or administered in admixture with an adjuvant.

The invention further relates to antibodies elicited in an animal by the administration of one or more *S. aureus* polypeptides of the present invention and to methods for producing
5 such antibodies and fragments thereof. The invention further relates to recombinant antibodies and fragments thereof and to methods for producing such antibodies and fragments thereof.

The invention also provides diagnostic methods for detecting the expression of the polynucleotides and polypeptides of Table 1 by members of the *Staphylococcus* genus in a
10 biological or environmental sample. One such method involves assaying for the expression of a polynucleotide encoding *S. aureus* polypeptides in a sample from an animal. This expression may be assayed either directly (e.g., by assaying polypeptide levels using antibodies elicited in response to amino acid sequences described in Table 1) or indirectly (e.g., by assaying for antibodies having specificity for amino acid sequences described in
15 Table 1). The expression of polynucleotides can also be assayed by detecting the nucleic acids of Table 1. An example of such a method involves the use of the polymerase chain reaction (PCR) to amplify and detect *Staphylococcus* nucleic acid sequences in a biological or environmental sample.

The invention also includes a kit for analyzing samples for the presence of members
20 of the *Staphylococcus* genus in a biological or environmental sample. In a general embodiment, the kit includes at least one polynucleotide probe containing a nucleotide sequence that will specifically hybridize with a *S. aureus* nucleic acid molecule of Table 1 and a suitable container. In a specific embodiment, the kit includes two polynucleotide probes defining an internal region of the *S. aureus* nucleic acid molecule of Table 1, where each
25 probe has one strand containing a 31' mer-end internal to the region. In a further embodiment, the probes may be useful as primers for polymerase chain reaction amplification.

The present invention also relates to nucleic acid probes having all or part of a nucleotide sequence described in Table 1 which are capable of hybridizing under stringent conditions to *Staphylococcus* nucleic acids. The invention further relates to a method of
30 detecting one or more *Staphylococcus* nucleic acids in a biological sample obtained from an animal, said one or more nucleic acids encoding *Staphylococcus* polypeptides, comprising:
(a) contacting the sample with one or more of the above-described nucleic acid probes, under

conditions such that hybridization occurs, and (b) detecting hybridization of said one or more probes to the *Staphylococcus* nucleic acid present in the biological sample.

By "biological sample" is intended any biological sample obtained from an individual, body fluid, cell-line, tissue culture, or other source which contains *S. aureus* polypeptides or polynucleotides of the invention. As indicated, biological samples include body fluids (such as semen, lymph, sera, plasma, urine, synovial fluid and spinal fluid) which contain the *S. aureus* polypeptides or polynucleotides of the invention, and tissue sources found to contain the expressed *S. aureus* polypeptides shown in Table 1. Methods for obtaining tissue biopsies and body fluids from mammals are well known in the art. Where the biological sample is to include mRNA, a tissue biopsy is the preferred source.

The method(s) provided above may preferably be applied in a diagnostic method and/or kits in which *S. aureus* polynucleotides and/or polypeptides of the invention are attached to a solid support. In one exemplary method, the support may be a "gene chip" or a "biological chip" as described in US Patents 5,837,832, 5,874,219, and 5,856,174. Further, such a gene chip with *S. aureus* polynucleotides of Table 1 attached may be used to diagnose *S. aureus* infection in a mammal, preferably a human. The US Patents referenced above are incorporated herein by reference in their entirety.

Detailed Description

The present invention relates to recombinant antigenic *S. aureus* polypeptides and fragments thereof. The invention also relates to methods for using these polypeptides to produce immunological responses and to confer immunological protection to disease caused by members of the genus *Staphylococcus*. The invention further relates to nucleic acid sequences which encode antigenic *S. aureus* polypeptides and to methods for detecting *Staphylococcus* nucleic acids and polypeptides in biological samples. The invention also relates to *Staphylococcus* specific antibodies and methods for detecting such antibodies produced in a host animal. The invention relates to antagonists of polypeptides of the invention, including but not limited to antibodies and small molecule inhibitors.

Definitions

The following definitions are provided to clarify the subject matter which the

inventors consider to be the present invention.

As used herein, the phrase "pathogenic agent" means an agent which causes a disease state or affliction in an animal. Included within this definition, for examples, are bacteria, protozoans, fungi, viruses and metazoan parasites which either produce a disease state or
5 render an animal infected with such an organism susceptible to a disease state (e.g., a secondary infection). Further included are species and strains of the genus *Staphylococcus* which produce disease states in animals.

As used herein, the term "organism" means any living biological system, including viruses, regardless of whether it is a pathogenic agent.

10 As used herein, the term "*Staphylococcus*" means any species or strain of bacteria which is members of the genus *Staphylococcus* regardless of whether they are known pathogenic agents.

As used herein, the phrase "one or more *S. aureus* polypeptides of the present invention" means the amino acid sequence of one or more of the *S. aureus* polypeptides
15 disclosed in Table 1. These polypeptides may be expressed as fusion proteins wherein the *S. aureus* polypeptides of the present invention are linked to additional amino acid sequences which may be of Staphylococcal or non-Staphylococcal origin (e.g. His tagged fusion proteins). This phrase further includes fragments of the *S. aureus* polypeptides of the present invention.

20 As used herein, the phrase "full-length amino acid sequence" and "full-length polypeptide" refer to an amino acid sequence or polypeptide encoded by a full-length open reading frame (ORF). For purposes of the present invention, polynucleotide ORFs in Table 1 are defined by the corresponding polypeptide sequences of Table 1 encoded by said polynucleotide. Therefore, a polynucleotide ORF is defined at the 5' end by the first base
25 coding for the initiation codon of the corresponding polypeptide sequence of Table 1 and is defined at the 3' end by the last base of the last codon of said polypeptide sequence. As is well known in the art, initiation codons for bacterial species may include, but are not limited to, those encoding Methionine, Valine, or Leucine. As discussed below for polynucleotide fragments, the ORFs of the present invention may be claimed by a 5' and 3' position of a
30 polynucleotide sequence of the present invention wherein the first base of said sequence is position 1.

As used herein, the phrase "truncated amino acid sequence" and "truncated

polypeptide" refer to a sub-sequence of a full-length amino acid sequence or polypeptide. Several criteria may also be used to define the truncated amino acid sequence or polypeptide. For example, a truncated polypeptide may be defined as a mature polypeptide (e.g., a polypeptide which lacks a leader sequence). A truncated polypeptide may also be defined as
5 an amino acid sequence which is a portion of a longer sequence that has been selected for ease of expression in a heterologous system but retains regions which render the polypeptide useful for use in vaccines (e.g., antigenic regions which are expected to elicit a protective immune response).

Additional definitions are provided throughout the specification.

10 *Explanation of Table 1*

Table 1 lists the full length *S. aureus* polynucleotide and polypeptide sequences of the present invention and their associated SEQ ID NOs. Each polynucleotide and polypeptide sequence is preceded by a gene identifier. Each polynucleotide sequence is followed by at
15 least one polypeptide sequence encoded by said polynucleotide. For some of the sequences of Table 1, a known biological activity and the name of the homolog with similar activity is listed after the gene sequence identifier.

20 *Explanation of Table 2*

Table 2 lists accession numbers for the closest matching sequences between the polypeptides of the present invention and those available through GenBank and GeneSeq databases. These reference numbers are the database entry numbers commonly used by those of skill in the art, who will be familiar with their denominations. The descriptions of the nomenclature for GenBank are available from the National Center for Biotechnology
25 Information. Column 1 lists the polynucleotide sequence of the present invention. Column 2 lists the accession number of a "match" gene sequence in GenBank or GeneSeq databases. Column 3 lists the description of the "match" gene sequence. Columns 4 and 5 are the high score and smallest sum probability, respectively, calculated by BLAST. Polypeptides of the present invention that do not share significant identity/similarity with any polypeptide
30 sequences of GenBank and GeneSeq are not represented in Table 2. Polypeptides of the present invention that share significant identity/similarity with more than one of the polypeptides of GenBank and GeneSeq may be represented more than once.

Explanation of Table 3.

The *S. aureus* polypeptides of the present invention may include one or more conservative amino acid substitutions from natural mutations or human manipulation as indicated in Table 3. Changes are preferably of a minor nature, such as conservative amino acid substitutions that do not significantly affect the folding or activity of the protein. Residues from the following groups, as indicated in Table 3, may be substituted for one another: Aromatic, Hydrophobic, Polar, Basic, Acidic, and Small,

Explanation of Table 4

Table 4 lists residues comprising antigenic epitopes of antigenic epitope-bearing fragments present in each of the full length *S. aureus* polypeptides described in Table 1 as predicted by the inventors using the algorithm of Jameson and Wolf, (1988) Comp. Appl. Biosci. 4:181-186. The Jameson-Wolf antigenic analysis was performed using the computer program PROTEAN (Version 3.11 for the Power MacIntosh, DNASTAR, Inc., 1228 South Park Street Madison, WI). *S. aureus* polypeptides shown in Table 1 may possess one or more antigenic epitopes comprising residues described in Table 4. It will be appreciated that depending on the analytical criteria used to predict antigenic determinants, the exact address of the determinant may vary slightly. The residues and locations shown described in Table 4 correspond to the amino acid sequences for each full length polypeptide sequence shown in Table 1 and in the Sequence Listing. Polypeptides of the present invention that do not have antigenic epitopes recognized by the Jameson-Wolf algorithm are not represented in Table 2.

Nucleic Acid Molecules

Sequenced *S. aureus* genomic DNA was obtained from the *S. aureus* strain ISP3. *S. aureus* strain ISP3, has been deposited at the American Type Culture Collection, as a convenience to those of skill in the art. The *S. aureus* strain ISP3 was deposited on 7 April 1998 at the ATCC, 10801 University Blvd. Manassas, VA 20110-2209, and given accession number 202108. As discussed elsewhere herein, polynucleotides of the present invention readily may be obtained by routine application of well known and standard procedures for cloning and sequencing DNA. A wide variety of *S. aureus* strains can be used to prepare *S. aureus* genomic DNA for cloning and for obtaining polynucleotides and polypeptides of the

present invention. A wide variety of *S. aureus* strains are available to the public from recognized depository institutions, such as the American Type Culture Collection (ATCC). It is recognized that minor variations in the nucleic acid and amino acid sequence may be expected from *S. aureus* strain to strain. The present invention provides for genes, including both polynucleotides and polypeptides, of the present invention from all the *S. aureus* strains.

Unless otherwise indicated, all nucleotide sequences determined by sequencing a DNA molecule herein were determined using an automated DNA sequencer (such as the Model 373 from Applied Biosystems, Inc., Foster City, CA), and all amino acid sequences of polypeptides encoded by DNA molecules determined herein were predicted by translation of a DNA sequence determined as above. Therefore, as is known in the art for any DNA sequence determined by this automated approach, any nucleotide sequence determined herein may contain some errors. Nucleotide sequences determined by automation are typically at least about 90% identical, more typically at least about 95% to at least about 99.9% identical to the actual nucleotide sequence of the sequenced DNA molecule. The actual sequence can be more precisely determined by other approaches including manual DNA sequencing methods well known in the art. By "nucleotide sequence" of a nucleic acid molecule or polynucleotide is intended to mean either a DNA or RNA sequence. Using the information provided herein, such as the nucleotide sequence in Table 1, a nucleic acid molecule of the present invention encoding a *S. aureus* polypeptide may be obtained using standard cloning and screening procedures, such as those for cloning DNAs using genomic DNA as starting material. See, e.g., Sambrook et al. MOLECULAR CLONING: A LABORATORY MANUAL (Cold Spring Harbor, N.Y. 2nd ed. 1989); Ausubel et al., CURRENT PROTOCOLS IN MOLECULAR BIOLOGY (John Wiley and Sons, N.Y. 1989). Illustrative of the invention, the nucleic acid molecule described in Table 1 was discovered in a DNA library derived from a *S. aureus* ISP3 genomic DNA.

TABLE 1. Nucleotide and Amino Acid Sequences of *S. aureus* Genes.

>HGS010 murC (SEQ ID NO:1)

ATGACACACTATCATTTTGTCTCGGAATTAAAGGTTCTGGCATGAGTTCATTAGCACAAATCATGCATGATTTAGGACATGAAGT
 TCAAGGATCGGATATTGAGAACTACGTATTTACAGAAGTTGCTCTTAGAAATAAGGGGATAAAAAATATTACCATTGATGCTA
 ATAACATAAAAGAAGATATGGTAGTTATACAAAGTAATGCATTCCGAGTAGCCATGAAGAAATAGTACGTGCACATCAATTG
 AAATTAGATGTTGTAAAGTTATAATGATTTTTTAGGACAGATTATGATCAATATACTTCAGTAGCTGTAAGTGGTGACATGG
 TAAACTTCTACAAACAGGTTTTATTATCACATGTTATGAATGGTGATAAAAAAGACTTCATTTTTAATTGGTGATGGCACAGGTA
 TGGGATTGCCTGAAAGTGATTATTTTCGCTTTTGAGGCATGTGAATATAGACGTCACTTTTTAAGTTATAAACCTGATTACGCA
 ATTATGACAAATATGATTTTCGATCATCTGATTATTTTAAAGATATTAATGATGTTTTTGTATGCATTCCAAGAAATGGCACA
 TAATGTTAAAAAGGTATTATTGCTTGGGGTGATGATGAACATCTACGTAAAATTGAAGCAGATGTTCCAATTTATTATTATG
 GATTTAAAGATTCCGATGACATTTATGCTCAAAATATTCAAATTACGGATAAAGGTACTGCTTTTGATGTGTATGGATGGT
 GAGTTTTATGATCACTTCTGTCTCCACAATATGGTGACCATACAGTTTTAAATGCATTAGCTGTAATTGCGATTAGTTATTT
 AGAGAAGCTAGATGTTACAAATATTAAGAAGCATTAGAAACGTTTGGTGGTGTTAAACGTCGTTCAATGAAACTACAATTG
 CAAATCAAGTTATTGTAGATGATTATGACACCATCCAAAGAGAAATAGTGCTACAATTGAAACAGCACGAAAGAAATATCCA
 CATAAAGAAGTTGTTGCAGTATTTCAACCACACACTTTCTCTAGAACACAGGCATTTTTAAATGAATTTGCAGAAAGTTAAG
 TAAAGCAGATCGTGATTCTTATGTGAAATTTTGGATCAATTAGAGAAATACTGGCGCATTAAAGATACAAGATTAAATG
 ATAAATTTGAAGTGCATCGTTAATTAATGAAGATTCTATTAAATGATTAGAACAAATTTGATAATGCTGTATTTTATTATG
 GGTGCAGGTGATATTCAAAAATTACAAAATGCATATTTAGATAAATTAGGCATGAAAAATGCGTTTTAAGCTT

>HGS010 MurC (SEQ ID NO:2)

MTHYHFVGIKSGSMSSLAQIMHDLGHEVQGSIDIENYVFTEVALRNKGIKILPFDANNIKEDMV
 VIQGNFASSHEEIVRAHQLKLDVVSYNDFLQCIIDQYTSVAVTGAHGKTSSTGLLSHVMNGDKKTSFLIGDGTG
 MGLPESDYFAFEACEYRRHFLSYKPDYAIMTNIDFDHPDYFKDINDVFDQFQEMAHNVKKGIIAWGDDEHLRKIE
 ADVPIYYYGFKSDDIYAQNIQITDKGTAFDVYVDGEFYDHFSLPOYGDHTVLNALAVIAISYLEKLDVTNIKEA
 LETFGGVKRRFNETTIANQVIIVDDYAHHPREISATIEARKKYPHKEVVAVFQPHTFSRTOAFLNEFAESLSKAD
 RVFLCEIFGSIRENTGALTIQDLIDKIEGASLINESINVLEQPDNAVILFMGAGDIQKLQNAAYLDKLGKMNAP

>HGS027 Rfl (peptide chain release factor1) (SEQ ID NO:3)

ATGCATTTTGATCAATTAGATATTGTAGAAGAAAGATACGAACAGTTAAATGAAGTGAAGTACCCAGATGTTGTAAATGA
 TTCAGATAAATTACGTAAATATCTAAAGAGCAAGCTGATTTACAAAAAAGTGTAGATGTTATCGTAACTATAAAGCTAAAA
 AAGAAGAATTAGCTGATATTGAAGAAATGTTAAGTGAGACTGATGATAAAGAAGAAGTAGAAATGTTAAAGAGGAGAGTAAT
 GGTATTAAAGCTGAACCTCCAAATCTTGAAGAAGAGCTTAAATATATTGATTCTCTAAAGATCCTAATGATGACAAAGACGT
 TATTGTAGAAATAAGAGCAGCAGCAGGTGGTGATGAGGCTGCGATTTTGTCTGGTGATTTAATGCGTATGTATTCAAAGTATG
 CTGAATCAAGGATTCAAAACTGAAATAGTAGAAGCGCTCTGAAAGTGACCATGGTGGTTACAAAGAAATTAGTTTCTCAGTT
 TCTGGTAATGGCGGTATAGTAAATTTGAAATTTGAAATGGTGCACCGCGTTCAACGTGTGCCTGAAACAGAAATCAGGTGG
 ACGTATTCATACTTCAACAGCTACAGTGGCAGTTTTACCAGAAGTTGAAGATGTAGAAATTGAAATTAGAAATGAAGATTTAA
 AAATCGACACGTATCGTTCAAGTGGTGCAGGTGGTCAGCACGTAAACACAAGTACTCTGCAGTACGTATTACCCATTTACCA
 ACTGGTGTCTATTGCAACATCTTCTGAGAAGTCTCAAATTTCAAACCGTGAAAAGCAATGAAAGTGTAAAAGCACGTTTATA
 CGATATGAAGTTCAAGAAGAACAACAAAGTATGCGTCACAACGTAAATCAGCAGTCCGTACTGGTGATCGTTCAGAACGTA
 TTCGAACTTATAATTATCCAAAGCCGTGTAACAGACCATCGTATAGGTCTAACGCTTCAAAAATTAGGGCAAATTATGGAA
 GGCCATTTAGAAGAAATTATAGATGCACTGACTTTATCAGAGCAGACAGATAAATTGAAAGAACTTAATAATGGTGAA

>HGS027 Rfl (peptide chain release factor1) (SEQ ID NO:4)

MHFDQLDIVEERYEQLNELLSDPDVVNDSDKLKYSKEQADLQKTVDVYRNYKAKKEELADIEEMLSETDDKEEV
 EMLKEESNGIKAEPLNLEELKILLIPKDPNDKDVIVEIRAAAGGDEAAIFAGDLMRMYSKYAESQGFKEIVE
 ASEDHGGYKEISFVSNGAYSKLKFENGHRVQRPETESGGRIHTSTATVAVLPEVEDVEIEIRNEDLKIDT
 YRSSGAGGQHVNTTDSAVRIHLPTGVIATSSSEKSQIQNREKAMKVLKARLYDMKVQEEQKYASQRKSAVGTGD
 RSRIRTYNYPQSRVTDHRIGLTLQKLQIMEGHLEEIIDLTLSEQTDKLELNNGE

>HGS029 Rrf (ribosome recycling factor) (SEQ ID NO:5)

ATGGGGAGTGACATTATTAATGAACTAAATCAAGAAATGCAAAATCAATCGAAAGCTTATCACGTGAATTAGCTAACATCAG
 TGCAGGAAGAGCTAATTCAAATTTATTAAACGGCGTAACAGTTGATTACTATGGTGCACCAACACCTGTACAACAATTAGCAA
 GCATCAATGTTCCAGAAGCACGTTTACTTGTTATTTCTCCATACGACAAAACCTTCTGTAGCTGACATCGAAAAGCGATAATA
 GCAGCTAACTTAGGTGTTAACCACAAGTATGGTGAAGTGATACGTATTGCTGTACCTGCCTTAACAGAAGAAGCTAGAAA
 AGAGCGCGTTAAAGATGTTAAGAAAAATTGGTGAAGAAGCTAAAGTATCTGTTTCAAAATATTTCGTCGTGATGAATGATCAGT
 TGAAAAAGATGAAAAAATGGCGACATTACTGAAGATGAGTTGAGAAGTGGCACTGAAGATGTTTCAAGAAAGCAACAGACAAT
 TCAATAAAGAAATTGATCAAATGATTGCTGATAAAGAAAAAGATATTATGTCAGTA

>HGS029 Rrf (ribosome recycling factor) (SEQ ID NO:6)

MGSDIINETKSRMQKSIESLSRELANISAGRANSNLLNGVTVDYVGAPTPVQQLASINVPEARLLVISPYDKTSV

ADIEKAI I AANLGVNPTS DGEVIRI AVPALTEERRKERV KDVKKIGEEAKVSVRNIRDMNDQLKKDEKNGDITE
DELRSGETEDVQKATDNSIK EIDQMIADKEKDIMSV

>HGS038 nusA (SEQ ID NO:7)

ATGGGGTCAAGTAATGAATTATTATTAGCTACTGAGTATTTAGAAAAAGAAAAGAAGATTCCTAGAGCAGTATTAATTGATGC
TATTGAAGCAGCTTTAATTACTGCATACAAAAAGAACTATGATAGTGCAAGAAATGTCCGTGTGGAAATTAATATGGATCAAG
GTACTTTCAAAGTTATCGCTCGTAAAGATGTTGTTGAAGAAGTATTTGACGACAGAGATGAAGTGGATTAAAGTACAGCGCTT
GTTAAAAACCTCGCATATGAAATTGGTGATATATACGAAGAAGATGTAACACCTAAAGATTTTGGTCGTGTAGGTGCTCAAGC
AGCGAAACAAGCAGTAATGCAACGTCTTCGTGATGCTGAACGTGAAATTTTATTGAAGAATTTATAGACAAAAGAAGAAGACA
TACTTACTGGAATTATTGACCGTGTGACCATCGTTATGTATATGTGAATTTAGGTCGTATCGAAGCTGTTTTATCTGAAGCA
GAAAGAAGTCCTAACGAAAAATATATTCCTAACGAACGTATCAAAGTATATGTTAACAAAGTGGAAACAAACGACAAAAGGTCC
TCAAATCTATGTTTCTCGTAGCCATCCAGGTTTATTTAAACGTTTATTGAAACAAGAAGTCCAGAAATTTACGATGGTACTG
TAATTGTTAAATCAGTAGCAGCTGAAGCTGGCGATCGCTCTAAAATTAGTGTCTTCTCTGAAAAACAATGATATAGATGCTGTT
GGTGCAATGTTGGTGCTAAAGGCGCACGTGTTGAAGCTGTTGTTGAAGAGCTAGGTGGTGAAAAAATCGACATCGTTCAATG
GAATGAAGATCCAAAAGTATTTGTA AAAAATGCTTTAAGCCCTTCTCAAGTTT TAGAAGTTATTGTTGATGAAACAAATCAAT
CTACAGTAGTTGTTGTTCTGATTATCAATTGTCATTAGCGATTGGTAAAAGAGGACAAAACGCACGTCTAGCTGCTAAATTA
ACCGGCTGGAAAATTGATATTAAATCAGAAACAGATGCGCGTGAAGCGGGTATCTATCCAGTAGTTGAAGCTGAAAAGTAAC
TGAAGAAGATGTTGCTTTAGAAGATGCTGACACAACAGAATCAACCGAAGAGGTAAATGATGTTTCAGTTGAAAACAAATGTAG
AGAAAGAATCTGAA

>HGS038 NusA (SEQ ID NO:8)

MGSSNELLLATEYLEKEKKIPRAVLIDALEAALITAYKKNYDSARNVRVELNMDQGTFFKVIARKDVVEEVFDDRD
EVDLSTALVKNPAYEIGDIYEEDVTPKDFGRVGAQAQAVMQRRLDAEREILFEFIDKEEDILTGIIIDRVDRH
YVYVNLGRIEAVLSEAERSPNKEYIPNERIKVYVNVKVEQTTKGPQIYVSRSHPLLKRLFEQEVPEIYDGTIVIK
SVAREAGDRSKISVFSENNIDAVGACVGAKGARVEAVVEELGGEKIDIVQWNEPDKVFKNALSPSQVLEIVD
ETNQSTVVVVPDYQLSLAIGRQGNARLAALKLTGWKIDIKSETDAREAGIYPVVEAEKVTEEDVALEDADTTTEST
EEVNDVSVETNVEKESE

>HGS039 nusG (SEQ ID NO:9)

ATGGGATCTGAAGAGTTGGCGCAAAGCGTTGGTATGCAGTGCATACATATTCTGGATATGAAAAATAAGTTAAAAAGAATTT
AGAAAAAGAGTAGAATCTATGAATATGACTGAACAAATCTTTAGAGTAGTCATACCGGAAGAAGAAGAACTCAAGTAAAAG
ATGGCAAAGCTAAACGACTGTTAAAAAACATTCCCTGGATATGTTTATAGTGAATTAATCATGACAGATGAATCATGGTAT
GTGGTAAGAAATACACCAAGCGTTACTGGTTTGTAGGTTCTGCAGGTGCAGGGTCTAAGCCAAATCCATTGTTACCAGAAGA
AGTTCGCTTCATCTTAAACAAATGGGTCTTAAAGAAAAGACTATCGATGTTGAACTCGAAGTTGGCGAGCAAGTTCGTATTA
AATCAGGTCCATTGCGAATCAAGTTGGTGAAGTTCAAGAAATTGAAACAGATAAGTTTAAAGCTAACAGTATTAGTAGATATG
TTTGGCCGAGAAACACCAAGTAGAAGTTGAATTCGATCAAATTGAAAAGCTG

>HGS039 NusG (SEQ ID NO:10)

MGSEEVGAKRWYAVHTYSGYENKVKKNLEKRVESMMNTEQIFRVVIPEEEETQVKDGAKTTVKKTFPGYVLVEL
IMTDESYYVVRNTPGVTFVGSAGAGSKPNLLPEEVRFLKQMLKEKTIIDVELEVGEQVRIKSGPFANQVGEV
QEIETDKFKLTLVDMFGRETPEVEFEFDQIEKL

>HGS041. nade (NH3-Dependent NAD Synthetase) (SEQ ID NO:11)

ATGGGTAGTAAATTACAAGACGTTATTGTACAAGAAATGAAAGTGAAAAAGCGTATCGATAGTGCTGAAGAAATTATGGAATT
AAAGCAATTTATAAAAAATTATGTACAATCACATTCAATTTATAAAATCTTTAGTGTTAGGTATTTTCAGGAGGACAGGATTCTA
CATTAGTTGGAAGAACTAGTACAAATGTCTGTTAACGAATTACGTGAAGAAGGCATTGATTGTACGTTTATTGCAGTTAAATTA
CCTTATGGAGTTCAAAAAGATGCTGATGAAGTTGAGCAAGCTTTGCGATTCAATTGAACCAGATGAAATAGTAACAGTCAATAT
TAAGCCTGCAGTTGATCAAAGTGTCATATCAAAGAAGCCGGTATTGTTCTTACAGATTTCCAAAAAGGAAATGAAAAAG
CGCGTGAACGTATGAAAGTACAATTTCAATTGCTTCAAACCGACAAGGTATTGTAGTAGGAACAGATCATTAGCTGAAAAT
ATAACTGGGTTTTATACGAAGTACGGTGATGGTGCTGCAGATATCGACCTATATTGGTTTGAATAAACGACAAGGTCGTCA
ATTATTAGCGTATCTGGTGCGCAAAGGAATTATATGAAAAACGCCAACTGCTGATTTAGAAGATGATAAACACAGCTTC
CAGATGAAGATGCATTAGGTGTAACCTATGAGGCGATTGATAATTATTTAGAAGGTAAGCCAGTTACGCCAGAAGAACAAAA
GTAATTGAAAATCATTATATACGAAATGCACACAAACGTGAACCTGCATATACAAGATACACGTGGCCAAAATCC

>HGS041 Nade (NH3-Dependent NAD Synthetase) (SEQ ID NO:12)

MGSKLQDVIVQEMKVKRIDSAAEIMELKQFIKNYVQSHSFIKSLVLGISGGQDSTLVGKLVQMSVNELREEGID
CTFIIVKLPGVQKDADEVEQALRFIEPDEIVTVNIKPAVDQSVQSLKEAGIVLTDFOKGNEKARERMKVQFSIA
SNRQGIIVGTDHSAENITGFYTKYGDGAADIAPIFGLNKRQGRQLLAYLGAPKELYEKTPTADLEDDKPLPDED
ALGVTYEAIDNYLEKPVTPPEEQVIENHYIRNAHKRELAYTRYTPKS

>HGS042 trxB (Thioredoxin Reductase) (SEQ ID NO:13)
ATGGGTACTGAAATAGATTTTGTATAGCAATTATCGGTGCAGGTCCAGTGGTATGACTGCTGCAGTATACGCATCAGGTGC
TAATTTAAAAACAGTTATGATTGAAAGAGGTATTCAGGCGGTCAAATGGCTAATACAGAAGAAGTAGAGAACTTCCCTGGTT
TCGAAATGATTACAGGTCCAGATTTATCTACAAAAATGTTTGAACACGCTAAAAAGTTTGGTGCAGTTTATCAATATGGAGAT
ATTAAATCTGTAGAAGATAAAGGCGAATATAAAGTGATTAACCTTTGGTAATAAAGAATTAACAGCGAAAGCGGTATTATTGTC
TACAGGTGCAGAATACAAGAAAATTGGTGTTCGGGTGAAACAAGAATTTGGTGGACGCGGTGTAAGTTATTGTGCAGTATGTG
ATGGTGCATTCTTTAAAAATAAACGCCCTATTTCGTTATCGGTGGTGGTATTACAGCAGTAGAAGAGGGAACATTCTTAATACTAA
TTTGCTGACAAAGTAACAATCGTTCACCGTCGTGATGAGTTACGTGCACAGCGTATTTTACAAGATAGAGCATTCAAAAATGA
TAAAATCGACTTTATTGGAGTCATACCTTTGAAATCAATTAATGAAAAAGACGGCAAAGTGGGTTCTGTGACATTAACGCTCTA
CAAAAGATGGTTTCAAGAAGAACACACGAGGCTGATGGTGTATTCTATCTATATTGGTATGAAACCATTAACAGCGCCATTAAAA
GACTTAGGTATTACAAATGATGTTGGTTATATTGTAACAAAAGATGATATGACAACATCAGTACCAGTATTTTTCGACGAGG
AGATGTTTCGCGACAAGGTTTACGCCAATTGTCACTGCTACTGGCGATGGTAGTATTGCAGCGCAAAGTGCAGCGGAATATA
TTGAACATTTAAACGATCAAGCT

>HGS042 TrxB (Thioredoxin Reductase) (SEQ ID NO:14)
MGTEIDFDIAIIGAGPAGMTAAVYASRANLKTVMIERGIPGGQMANTEEEVENFPGFEMITGPDLSKMFHAKKF
GAVYQYQDIKSVEDKGEYKVINFGNKELTAKAVIIATGAEYKKIGVPEQELGGRGVSYCAVCDGAFKKNRFLV
IGGGDSAVEEGTFLTKFADKVTIVHRRDELRAQRILQDRAFKNDKIDFIWSHTLKSINEKDGKVGSVTLTSTKDG
SEETHEADGVFIYIGMKPLTAPFKDLGITNDVGYIVTKDDMTTSVPGIFAAGDVDRDKGLRQIVTATGDGSIQAQS
AAEYIEHLNDQA

>HGS043 femD/glmM (Phosphoglucosamine Mutase) (SEQ ID NO:15)
ATGGGGGGAAAAATTTTGGTACAGACGGAGTAAGAGGTGTCGCAAACCAAGAACTAACACCTGAATTGGCATTAAATTAGG
AAGATACGGTGGCTATGTTCTAGCACATAATAAAGGTGAAAAACACCCACGTGACTTGTAGGTGCGGATACTAGAGTTTCAG
GTGAAATGTTAGAATCAGCATTAAATAGCTGGTTTGATTTCATTTGGTGCAGAAGTGATGCGATTAGGTATTATTTCACACCA
GGTGTTCATATTTAACACCGCATATGGGTGCAGAGTTAGGTGTAATGATTTACGCTCTCATAATCCAGTTGCAGATAATGG
TATTAATTTCTTTGGATCAGATGGTTTAAACTATCAGATGAACAAGAAAATGAAATTGAAGCATTATTGGATCAAGAAAACC
CAGAATTACCAAGACCAGTTGGCAATGATATTGTACATTATTCAGATTACTTTGAAGGGGCACAAAAATATTGAGCTATTTA
AAATCAACAGTAGATGTTAACTTTGAAGTTTGAATAATGCTTTAGATGGTGCAATGGTTCAACATCATCTAGCGCCATT
CTTATTTGGTGACTTAGAAGCAGATACTGAAACAATTGGATGTAGTCTGTATGGATATAATATCAATGAGAAATGTGGCTCTA
CACATCTGAAAAATAGCTGAAAAAGTAGTTGAACTGAAAGTGATTTTGGTTAGCATTGTGACGGCGATGGAGACAGAATC
ATAGCAGTAGATGAGAATGGTCAAATCGTTGACGGTGACCAAAATTATGTTTATTATTGGTCAAGAAATGCATAAAAAATCAAGA
ATTGAATAATGACATGATTGTTTCTACTGTTATGAGTAATTTAGGTTTTTACAAGCGCTTGAACAAGAAGGAATTAATCTA
ATAAACTAAAGTTGGCGACAGATATGTAGTAGAAGAAATGCGTCGCGGTAATTATACTTAGGTGGAGAACAATCTGGACAT
ATCGTTATGATGGATTACAATACACTGGTGATGGTTTATTAACCTGGTATTCAATTAGCTTCTGTAATAAAATGACTGGTAA
ATCACTAAGTGAAATTAGCTGGACAAATGAAAAATATCCACAATCATTAATTAACGTACGCGTAACAGATAAATATCGTGTG
AAGAAATGTTGACGTTAAAGAAGTTATGACTAAAGTAGAAGTAGAAATGAATGGAGAAGGTGCAATTTTAGTAAGACCTTCT
GGAACAGAACCATTAGTTCTGTGTCATGGTTGAAGCAGCACTGATGAAGATGCTGAAAGATTTCACAACAATAGCTGATGT
GGTTCAAGATAAAATGGGATTAGATAAA

>HGS043 FemD/GlmM (Phosphoglucosamine Mutase) (SEQ ID NO:16)
MGGKYFGTDGVRGVANQELTPELAFKLGRYGGYVLAHNKGEKHPRVLVGRDTRVSGEMLESALIAGLISIGAEVM
RLGIISTPGVAYLTRDMGAELGVMISASHNPVADNGIKFFGSDGFKLSDEQENEIEALLDQENPELPRPVGNDIV
HYSDFEQAQKYLKSTVDVNFELKIALDGGANGSTSSSLAPFLFGDLEADTETIGCSPDGYNINEKCGSTHPE
KLAEKVVETESDFGLAFDGDGRIIAVDENGQIVDGDQIMFIIQEMHKNQELNNDMIVSTVMSNLGFYKALEQE
GIKSNKTKVGDYVVEEMRRGNYNLGGEQSGHIVMMDYNTTGDLLTGIQLASVIKMTGKSLSELAGQMKYPQS
LINVRVTDKYRVEENVVKEVMTKVEVEMNGEGRILVVRPSGTEPLVRVMVEAATDEDAERFAQQIADVVQDKMGL
DK

>HGS044 glmU (Glucosamine N-acetyly/uridylylate transferase) (SEQ ID NO:17)
ATGGGTTTTCATGCGAAGACACGCGATAATTTGGCAGCAGGTAAAGGCACAAGAAATGAAATCTAAAAAGTATAAAGTGCTACA
CGAGGTGCTGGGAAACCTATGGTCAACATGTATTGGAAGTGTAAGGCTCTGGTGTGATCAAGTTGTAAACCATCGTAG
GACATGGTGTGAAAGTGTAAGGACATTTAGGCGAGCGTCTTTTATACAGTTTCAAGAGGAACAACCTCGGTACTGCGCAT
GCAGTGCAAAATGGCGAAATCACTTAGAAGACAAGGAAGGTACGACAATCGTTGTATGTGGTGACACACCGCTCATCAAAA
GGAACATTAGTAACATTGATTGCGCATCAGGAGGATGCTAATGCTCAAGCAACTGTATTATCTGCATCGATTCAACAACCAT
ATGGATACGGAAGAAATCGTTCGAAATGCGTCAGGTGCTTTAGAACGCATAGTTGAAGAGAAAGATGCAACGCAAGCTGAAAAG
GATATTAATGAAATTAGTTTCAAGTATTTTGGCTTTAATAATAAAACGTTGTTTGAATAATTAACAAGTGAATAATGATAA
TGCGCAAGGTGAATATTACCTCCCTGATGATTGTGCTTTAATAATAAAACGTTGTTTGAATAATTAACAAGTGAATAATGATAA
ATGTTGAAGAAATCATGGGTGTAAATGATCGTGTAAATGCTTAGTCAGGCTGAGAAGGCGATGCAACGTCGTACGAATCATTAT
CACATGCTAAATGGTGTGACAATCATCGATCCTGACAGCACTTATATTGGTCCAGACGTTACAATTGGTAGTGATACAGTCAT
TGAACCAGGCGTACGAATTAATGGTTCGTACAGAAATTTGCGGAAGATGTTGTTATTGGTCAGTACTCTGAAATTAACAATAGTA

CGATTGAAATGGTGCATGTATTCAACAGTCTGTTGTTAATGATGCTAGCGTAGGAGCGAATACTAAGGTCGGACCGTTTGCG
CAATTGAGACCAGGCGCGCAATTAGGTGCAGATGTTAAGGTTGGAAATTTTGTAGAAATTAAAAAGCAGATCTTAAAGATGG
TGCCAAGGTTTCACATTTAAGTTATATTGGCGATGCTGTAATTGGCGAACGTAATAATTGGTTGCGGAACGATTACAGTTA
ACTATGATGGTGAAAATAAAATTTAAACTATCGTCGGCAAAGATTCATTTGTAGGTTGCAATGTTAATTTAGTAGCACCTGTA
ACAATTGGTGATGATGTATTGGTGGCAGCTGGTCCACAATCACAGATGACGTACCAATGACAGTTTAGCTGTGGCAAGAGC
AAGACAAACAACAAAGAAGGATATAGGAAA

>HGS044 GlmU (Glucosamine N-acetyl/uridylyl transferase) (SEQ ID NO:18)
MGFMRRHAILAAGKGRMKSKEYVLHEVAGKPMVEHVLESVKSGVDQVVTIVGHGAESVKHGLGERSLYSFO
EEQLGTAHAVQMAKSHLEDKEGTTIVVCGDTPLITKETLVTLIAHHEDANAQATVLSASIQOPYGYGRIVRNASG
RLERIVEEKDATQAEKDINEISSGIFAFNNKTLFEKLTQVKNDAQGEYLPDVLSLIINDGGIVEVYRTNDVEE
IMGVNDVRMLSQAEKAMQRRTNHYHMLNGVTIIDPDSTYIGPDVTIGSDTVIEPGVRINGRTEIGEDVVIGQYSE
INNSTIENGACIQQSVVNDASVGANTKVGPFAQLRPGAQLGADVKNFVEIKKADLKDGAKVSHLSYIGDAVIG
ERTNIGCGTITVNYDGENKFKTIVGKDSFVGCNVNLVAPVTIGDDVLVAAGSTITDDVPNDLAVARARQTTKEG
YRK

>HGS045 coADR (CoenzymeA Disulfide Reductase) (SEQ ID NO:19)
ATGGGGCCCAAAATAGTCGTAGTCGGAGCAGTCGCTGGCGGTGCAACATGTGCCAGCCAAATTCGACGTTTGTAGATAAGAAAG
TGACATTATTATTTTGAAGAAAGATCGTGATAGAGCTTTGCTAATTGTGCATTGCCTTATGTCAATTGGCGAAGTTGTGAAG
ATAGAGATATGCTTTAGCGTATACACCTGAAAAATTTATGATAGAAAGCAAATTACAGTAAAAACTTTATCATGAAGTTATT
GCAATCAATGATGAAAGACAACTGTATCTGTATTAATAGAAAGACAAACGAACAATTGAAGAATCTTACGATAAACTCAT
TTTAAGCCCTGGTGCAAGTGCAAAATAGCCTTGGCTTTGAAAGTGATATTACATTTACACTTAGAAATTTAGAAGACACTGATG
CTATCGATCAATTCATCAAAGCAAATCAAGTTGATAAAGTATTGGTTGTAGGTGCAGGTTATGTTTCATTAGAAGTTCTTGAA
AATCTTTATGAACGTGGTTTACACCCTACTTTAATTCATCGATCTGATAAGATAAAATAATTAATGGATGCCGACATGAATCA
ACCTATACTTGATGAATTAGATAAGCGGGAGATTCCATACCGTTTAAATGAGGAAATTAATGCTATCAATGGAATGAAATTA
CATTTAAATCAGGAAAAGTTGACATTACGATATGATTATTGAAGGTGTCGGTACTCACCCCAATTCAAAATTTATCGAAAGT
TCAAATATCAAATCTGATCGAAAAGGTTTCATACCGGTAAACGATAAAATTTGAAACAAATGTTCCAAACATTTATGCAATAGG
CGATATTGCAACATCACATTATCGACATGTCGATCTACCGGCTAGTGTTCTCTTTAGCTTGGGGCGCTCACCGTGACGCAAGTA
TTGTGCGGCAACAAATTGCTGGAAATGACACTATTGAATCAAAGGCTTCTTAGGCAACAATATTGTGAAGTTCTTTGATTAT
ACATTTGCGAGTGTGCGGCTTAAACCAAACGAACATAAGCAATTTGACTATAAAATGGTAGAAGTCACTCAAGGTGCACACGC
GAATTATTACCCAGGAAATCCCTTTTACACTTAAGAGTATATTATGACACTTCAAACCGTCAGATTTTAAGAGCAGCTGCAG
TAGGAAAAGAAGGTGCAGATAAACGTATTGATGTACTATCGATGGCAATGATGAACCAGCTAACTGTAGATGAGTTAACTGAG
TTTGAAGTGGCTTATGCACCACCATATAGCCACCCTAAAGATTTAATCAATATGATTGGTTACAAGCTAAA

>HGS045 CoADR (CoenzymeA Disulfide Reductase) (SEQ ID NO:20)
MGPKIVVVGAVAGGATCASQIRRLDKESDIIIFEKDRDMSFANCALPYVIGEVVEDRRYALAYTPEKFDYRKQIT
VKTYHEVIAINDERQTVSVLNRKTNEQFEESYDKLILSPGASANS LGFESDITFTLRNLEDIDAIDQFIKANQVD
KVLVVGAGYVSVLEVLNLYERGLHPTLIHRSDKINKLMDADMNPILDELDKREIPYRLNEEINAINGNEITFKS
GKVEHYDMIIEGVNTHPNKFIESSNIKLDRKGFIPVNDKFETNPVNIYAIGDIATSHYRHVDLPASVPLAWGAH
RAASIVAEQIAGNDTIEFKGFLGNINIVKFFDYTFASVGVKPNELKQFDYKMEVVTQGAHANYYPGNSPLHLRVYY
DTSNRQILRAAVGKEGADKRIDVLSMAMMNQLTVDELTEFEVAYAPPYSHPKDLINMIGYKAK

>HGS046 SVR (SEQ ID NO:21)
ATGAAAGACGAACAATTATATTATTTTGAGAAATCGCCAGTATTTAAAGCGATGATGCATTTCTCATTGCCAATGATGATAGG
GACTTTATTAAGCGTTATTTATGGCATATTAATATTTACTTTATAGGATTTTGTAGAAGATAGCCACATGATTTCTGCTATCT
CTCTAACACTGCCAGTATTTGCTATCTTAATGGGGTTAGGTAATTTATTTGGCGTTGGTGACGAACTTATATTTACGTTTA
TTAGGTGCGAAAGACTATAGTAAGAGTAAATTTGTAAGTAGTTCTCTATTATGAGGTGTTATGCACTAGGACTTATCGTGAT
TTTAGTTACTTTACCATTCAAGTGATCAAATCGCAGCAATTTTAGGGGCGAGAGGTGAAACGTTAGCTTTAACAAGTAATTATT
TGAAAGTAATGTTTTAAGTGACCTTTTGTAAATTTGTTCTTCATATTAGAACAATTTGCACGTGCAATTGGGGCACCATG
GTTTCTATGATTGGTATGTTAGCTAGGTAGGCTTAAATATTATTTTAGATCCAATTTTAAATTTTGGTTTTGATTTAAACGT
TGTTGGTGACGCTTTGGGTACTGCAATCAGTAATGTTGCTGCTGCTCTGTTCTTTATCATTTATTTTATGAAAAATAGTGACG
TTGTGTCAGTTAATATTAACCTTGCGAAACCTAATAAGAAATGCTTTCTGAAATCTTTAAATCGGTATTCTGCAATTTTA
ATGAGTATCTTAATGGGATTACAGGATTAGTTTAAATTTATTTTAGCACATTATGGAACTTCGCGATTGCAAGTTATGG
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TTATGGCAAATAAAGCCGATGAAAGACGTTATCAAAGCAGTTATCATGTCTATCGGCGTTATCTTTGTGTATGATGAGT
GCTGTATTTACAATTGGACATCATATGGTCGGAATTTACTACTGATCAAGCCATTGTTGAGATGGCGACATTTATTTTGAA
AGTAACAAATGGCATATTATTATTAATGGTATAGGTTTCTGTTTACTGGTATGCTTCAAGCGACTGGGCAAGGTCGTGGTG
CTACAATTATGGCCATTTTACAAGGTGCAATTTATCATTCCAGTATTATTTATGAAATGCTTTGTTTGGACTAACAGGTGTC
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TGATACATCTGAATTAATAGAAGGT

>HGS046 SVR (SEQ ID NO:22)

MKDEQLYFEKSPVFKAMMHFSLPMMIGTLLSVIYGILNIYFIGFLED SHMISAI SLTLPVFAILMGLGNLFGVG
AGTYISRLLAGAKDYSKSKFVSSFSIYGGIALGLIVILVTLPFSDQIAAILGARGETLALTSNYLKVMFLSAPFVI
LFFILEQFARAIGAPMVSMIGMLASVGLNIILDPILIFGFDLNVVGAALGTAINVAAALFFIYFMKNSDVVSV
NIKLA KP NKEMLSEIFKIGIPAF LMSILMGFTGLVLNLF LAHYGNFAIASYGISFRLVQFP ELIIMGLCEGVVPL
IAYNFMANKGRMKDVIKAVIMSIGVIFVVCMSAVFTIGHH MVGLFTTDQAI VEMATFILKVTMASLLNLGIGFLF
TGMLQATGQGRGATIMAILQGAIIPVLFIMNALFGLTGVIVSLLIAESLCALAAMLIVYLLRDLTVDTS ELIE
G

>HGS049 murE (SEQ ID NO:23)

TGCGATGCAAGTACGTTGTTTAAAGAAAGTAAAGCGTGTATTGGGTTCTTTAGAACAAACAATAGATGATATCACTAC
TGATTCAGGTACAGCGAGAGAAAGGTAGCATT TTTGTCGCTTCAGTTGGATATACTGTAGACAGTCATAAGTTCTGTCAAATG
TAGCTGATCAAGGGTGTAAGTTGGTAGTGGTCAATAAAGAACCAATCATTACCAGCTAACGTAACACAAGTGGTTGCGCGGAC
ACATTAAGAGTAGCTAGTATTCTAGCACACACATTATATGATTATCCGAGTCATCAGTTAGTGACATT TGGTGTACCGGGTAC
AAATGGTAAACTTCTATTGCGACGATGATTCAATTAATCAAAGAAAGTTACAAAAAATAGTGCATATTTAGGAACATAATG
GTTTCCAAATTAATGAAACAAAGACAAAAGGTGCAAAATACGACACAGAACAGTTTCTTTAACTAAGAAAATTAAGAAGCA
GTTGATGCAGGCGCTGAATCTATGACATTAGAAGTATCAAGCCATGGCTTAGTATTAGGACGACTGCGAGGCGTTGAATTTGA
CGTTGCAATATTTTCAAATTTAACACAAGACCATTTAGATTTTTCATGGCACAAATGGAAGCATACGGACACGCGAAGTCTTTAT
TGTTTAGTCAATTAGGTGAAGATTGTGCGAAAGAAAGTATGTCGTGTTAAACAATGACGATTCATTTTCTGAGTATTTAAGA
ACAGTGACGCCTTATGAAGTATTAGTTATGGAATTGATGAGGAAGCCCCAATTATGGCTAAAAATATTCAAGAACTCTTTACA
AGGTGTCAGCTTTGATTTTGAACGCCTTTTGAACCTTACCCAGTAAATCGCCTTATGTTGGTAAGTTTAATATTTCTAATA
TTATGGCGGCAATGATTGCGGTGTGGAGTAAAGGTACATCTTTAGAAACGATTATTAAGCTGTGAAAAATTTAGAACCTGTT
GAAGGGCGATTAGAAGTTTATGATCCTTCGTTACCTATGATTAAATTCGATTATGCACATACAGCTGATGGTATGAACAA
ATTAATCGATGCAGTACAGCCTTTGTAAAGCAAAGTTGATATTTTATGTTGGTATGGCAGCGAAGCTGATTTAACTAAAA
CGCCTGAAATGGGGCAGTTGCTGTGTCGAGATTATGTCATTTTACACCGGATAATCCGGCAAATGATGACCCGAAAATG
TTAACGGCAGAAATTAGCCAAAGGTGCAACACATCAAACTATATTGAATTTGATGATCGTGCAGAGGATAAAACATGCAAT
TGACATAGCTGAGCCTGGGGATACTGTGCTTTTAGCATCAAAAGGAAGAGAACCATATCAAATCATGCCAGGGCATATTAAGG
TGCCACATCGAGATGATTAAATGGCCTTGAAGCAGCTTACAAAAAGTTCCGGTGGTGGCCCTGTTGAT

>HGS049 MurE (SEQ ID NO:24)

LDASTLFKKVKVRLVLSLEQQIDDITDTSRTAREGSIFVASVG YTVDSHKFCQNVADQGCKLVVVNKEQSLPANVTQVVVPD
TLRVASILAHTLYDYPHQVLTGVTGTNGKTSIATMIHLIQRKLQKNSAYLGTNGFQINETKTGANTTPETVSLTKKIKEA
VDAGAESMTLEVSSHGLVLGRLRGVEFDVAIFSNLTQDHLDFHGTMEAYGHAKSLLFSQLGEDLSKEKYVVLNNDSSFSEYLR
TVTPYEVFSY GIDEEAQFMAKNIQESLQGVSFDFVTPFGTYPVKSPYVGKFNISNIMAAMIAVWSKGTSLETIIKAVENLEPV
EGRLEVLDPSLPIDLIIDYAHTADGMNKLIDAVQP FVKQLIFLVGMAGERDLTKTPEMGRVACRADYVIFTDPNPANDDPKM
LTAELAKGATHQNYIEFDDRAEGIKHAIDIAEPGDTVVLASKGREPYQIMPGHIKVPHRDDLIGLEAAKYKFGGGPVD

>HGS050 MurF (SEQ ID NO:25)

ATGATTAAATGTTACATTAAAGCAAATTCATCATGGATTCTTGTGAAATTGAAGATCAATTTTAAATCAAGAGATAAATGG
AGTCACAATTGATTACAGGACCAATTTCTAAAAATATGTTATTTATACCATTTAAAGGTGAAATGTTGACGGTCACTCGCTTTG
TCTCTAAAGCATTACAAGATGGTGTGCTGGGGCTGCTTTTATCAAAGAGGGACACCTATAGATGAAATGTAAGCGGGCCTATT
ATATGGGTTGAAGACATATAACCGCATTACAACAATTGGCACAAGCTTACTTGAGACATGTAAACCCATAAGTAATTGCCGT
CACAGGCTCTAATGCTATAAACAACGACTAAAGATATGATTGAAAGTGATTGCATACCGAATTTAAAGTTAAGAAAACGCAAG
GTAATTACAATAATGAAATTGGTTTACCTTAACTATTTTGAATTAGATAATGATATACTGAAATATCAATATTGGAGATGGGG
ATGTCAGGTTTCCATGAAATGAATTTCTGTCAAACCTCGCTCAACCAGATATTGCAGTTATACTAATATTGGTGAGTCACA
TATGCAAGATTTAGGTTTCGCGCGAGGGGATTGCTAAAGCTAAATCTGAAATTACAATAGGTCTAAAGATAATGGTACGTTTA
TATATGATGGCGATGAACCATATTGAAACCATGTTAAAGAAGTTGAAATGCAAAATGATTAGTATTGGTGTGCTACT
GATAATGCATTAGTTTGTCTGTTGATGATAGAGATACTACAGGTATTTCAATTTACGATTAATAATAAAGAACATTACGATCT
GCCAATATTAGGAAAGCATAATATGAAAAATGCGACGATTGCCATTGCGGTTGGTCAATGAATTAGGTTTGACATATAACACAA
TCTATCAAAATTTAAAAAATGTCAGCTTAACTGGTATGCGTATGGAACAACATACATTAGAAAAATGATATTACTGTGATAAAT
GATGCTATAATGCAAGTCTACAAGTATGAGAGCAGCTATTGATACACTGAGTACTTTGACAGGGCGTCGCTATTCTAATTTT
AGGAGATGTTTTAGAAATTAGGTGAAAATAGCAAAGAAATGCATATCGGTGTAGGTAATTTATTTAGAAGAAAAGCATATAGATG
TGTTGTATACGTTTGGTAATGAAGCGAAGTATATTTATGATTCGGGCCAGCAACATGTCGAAAAAGCACAACTTCAATTCT
AAAGACGATATGATAGAAGTTTAAATAAAGCATTAAAAGCGCATGACCGTGTATTAGTTAAAGGATCAGTGGTATGAAATT
AGAAGAAGTGGTAAATGCTTTAATTTCA

>HGS050 MurF (SEQ ID NO:26)

MINVTLKQIQSWIPCEIEDQFLNQEINGVTIDSR AISKNMFLIPFKGENVDGHRFVSKALQDGAGAAFYQRGTFIDENVSGPI
IWVEDTLTALQQLAQAYLRHVNPKVIAVTGSNGKTTTKDMIESVLHTEFKVKKTGQNYNNEIGLPLTILELDNDTEISILEMG
MSGFHEIEFLSNLAQPDIAVITNIGESHMQDLGSRREGIAKAKSEITIGLKDNGTFIYDGDEPLLKPHVKEVENAKCISIGVAT
DNALVCSVDDRDTTGISFTINKEHYDLPILGKHNMKNATIAIAVGHELGLTYNTIYQNLKNVSLTGMRMEQHTLENDITVIN

DAYNASPTSMRAAIDTLSTLTGRRILILGDVLELGENSEMHIGVGNYLEEKHIDVLYTFGNEAKYIYDSGOQHVEKAQHFNS
KDDMIEVLINDLKAHDRVLVKSGRGMKLEEVNALIS

>HGS052 Ribosomal Protein S8 (SEQ ID NO:27)

ATGACAATGACAGATCCAATCGCAGATATGCTTACTCGTGTAAGAAACGCAACATGGTGCCTCACGAGAAGTTAGAATTACC
TGCATCAAAATATTAAAAAAGAAATGCTGAAATCTTAAAGAGTGAAGGTTTCATTAAAAATGTTGAATACGTAGAAGATGATA
AACAAGGTGTAAGTTCGTTTATTCTTAAATATGGTCAAAACGATGAGCGTGTATCACAGGATTAAACCGTATTTCAAAACCA
GGTTTACGTGTTTATGCAAAAGCTAGCGAAATGCCTAAAGTATTAAATGGTTTAGGTATTGCATTAGTATCAACTTCTGAAGG
TGTAATCACTGACAAAGAAGCAAGAAAACGTAATGTTGGTGGAGAAATTATCGCATACTGTTG

>HGS052 Ribosomal Protein S8 (SEQ ID NO:28)

MTMTDPIADMLTRVRNANMVRHEKLELPASNIKKEIAEILKSEGFKNVEYVEDDKQGVLRFLKYGQNDERVIT
GLKRISKPLRVYAKASEMPKVLNGLGIALVSTSEGVITDKARKRNVGGEIIAYVW

>HGS053 Ribosomal Protein S15 (SEQ ID NO:29)

ATGGCAATTTACAAGAACGTAACGAAATCATTAAAGAATACCGGTGACACGAACTGATACTGGTTCACCAGAAGTACA
AATCGCTGTACTTACTGCGAGAAATCAACGCAGTAAACGACACTTACGTACACACAAAAAGACCACCATTCACGTCGTGGAT
TATTAAAAATGGTAGGTCGTCGTAGACATTTATTAACTACTTACGTAGTAAAGATATTCAACGTTACCGTGAATTAATTA
TCACTTGGCATCCGTCGT

>HGS053 Ribosomal Protein S15 (SEQ ID NO:30)

MAISQERKNEIIKEYRVHETDTGSPEVQIAVLTAEINAVNEHLRTHKKDHHSRRGLLMVGRRRHLLNLYRSKDI
QRYRELIKSLGIRR

>HGS055 Ribosomal Protein S3 (SEQ ID NO:31)

TAAGGAGGGAATACTGTGGGTCAAAAAATTAATCCAATCGAAGTTCGTTGGTATTATCCGTGATTGGGAAGCTAAATGGTA
TGCTGAAAAAGACTTCGCTTCACTTTTACACGAAGATTTAAAAATCCGTAAATTTATGATAATGAATTAAGAAGCATCAG
TTTCTCACGTAGAGATTGAACGTGTGCAACCGTATCAACATTGCAATTCATACTGGTAAACCTGGTATGGTAATTGGTAAA
GGCGGTTCAAGAAATCGAAAAATTACGCAACAAATTAATGCGTTAACTGATAAAAAAGTACACATCAACGTAATTGAAATCAA
AAAAGTTGATCTTGACGCTCGTTTAGTAGCTGAAAACATCGCACGTCAATTAGAAAACCGTGCTTCATTCCGTCGTGTACAAA
AACAAGCAATCACTAGAGCTATGAACTTGGTGCTAAAGGTATCAAACTCAAGTATCTGGTTCGTTAGGCGGAGCTGACATC
GCTCGTGCTGAACAATATTGAGAAGAACTGTTCCACTTCATACGTTACGTGCTGACATCGATTATGCACACGCTGAAGCTGA
CACTACTTACGTTAAATTAGGCGTTAAAGTATGGATTATCGTGGAAGATTCTTCTACTAAGAACACTAGTGGAGGAGGAA
AA

>HGS055 Ribosomal Protein S3 (SEQ ID NO:32)

VGQKINPIGLRVGIIRDWEAKWYAEKDFASLLHEDLKIRKFDIDNELKEASVSHVEIERAANRINIAIHTGKPGMVIGKGGSEI
EKLRLNKLNALTDKVVHINVIEIKKVDLDARLVAENIARQLENRASFRVQKQAITRAMKLGAKGIKTQVSGRLGGADIARAEO
YSEGTVPHLTLRADIDYAHAEADTTYGKLGKVVWYIRGEVLPKNTSGGGK

>HGS056 Ribosomal Protein S5 (SEQ ID NO:33)

ATGGCTCGTAGAGAAGAAGAGACGAAAGAATTTGAAGAACCGGTTGTTACAATCAACCGTGTAGCAAAAGTTGTAAGGTGG
TCGTGCTTTCCGTTTCACTGCATTAGTTGTAGTTGGAGACAAAATGGTCGTGTAGGTTTCGGTACTGGTAAAGCTCAAGAGG
TACCAGAAGCAATCAAAAAAGCTGTTGAAGCAGCTAAAAAGATTTAGTAGTTGTTCCACGTGTTGAAGGTACAACCTCACAC
ACAATTACTGGCCGTTACGGTTCAGGAAGCGTATTTATGAAACCGGCTGCACCTGGTACAGGAGTTATCGCTGGTGGTCCTGT
TCGTGCCGTACTTGAATTAGCAGGTATCACTGATATCTTAAGTAAATCATTAGGATCAAACACACCAATCAACATGGTTCGTG
CTACAATCGATGGTTTACAAAACCTTAAAAATGCTGAAGATGTTGCGAAATTACGTGGCAAAACAGTAGAAGAATTATACAAT

>HGS056 Ribosomal Protein S5 (SEQ ID NO:34)

MARREEETKEFEERVVTINRVAKVVGGRFRFTALVVVGDKNVRVFGTGKAQEVPEAIKKAVEAAKKDLVVVP
RVEGTTPTHITGRYSGSVFMKPAAPGTGVIAGGPVRAVLELAGITDILSKSLGSNTPINMVRATIDGLQNLKNA
EDVAKLRGKTVEELYN

>HGS057 Ribosomal Protein S9 (SEQ ID NO:35)

ATGGCACAAGTTGAATATAGAGGCACAGGCCGTCGTAACCACTCAGTAGCACGTGTACGTTTGTAGTACCAGGTGAAGGTAACAT
CACAGTTAATAACCGTGACGTACGCGAATACTTACCATTGCAATCATTAAATTTAGACTTAAACCAACCATTTGATGTAAGT
AACTAAAGGTAAGTATGATGTTTGTAGTTAACGTTTATGGTGGTGGTTTCACTGGACAAGCTCAAGCTATCCGTACCGGAATC
GCTCGTGCAATTATTAGAAGCAGATCCTGAATACAGAGGTTCTTTAAACCGCGCTGGATTACTTACTCGTGACCCACGTATGAA
AGAACATAAAAAACAGGTCTTAAAGCAGCTCGTCGTTCACTCAATTCTCAAAACGT

>HGS057 Ribosomal Protein S9 (SEQ ID NO:36)

MAQVEYRGTGRRKNSVARVRLVPGEGNITVNNRDVREYLPFESLILDNLNQPFDVTETKGNVDVLNVHGGGFTGQ
AQAIRHGFIARALLEADPEYRGSLSKRALLLTRDPRMKEHKKPGLKAARRSPQFSKR

>HGS058 Ribosomal Protein S10 (SEQ ID NO:37)

ATGGCAAACAAAAAATCAGAATCAGATTAAAGGCTTATGATCACCGCGTAATTGATCAATCAGCAGAGAAGATTGTAGAAAC
AGCGAAACGTTCTGGTGCAGATGTTTCTGGACCAATTCCGTTACCAACTGAGAAATCAGTTTACACAATCATCCGTGCCGTGC
ATAAGTATAAGATTACGCTGAACAATTGGAACAACGTACACACAAACGTTTAAATCGATATTGTAAACCAACACCAAAAACA
GTTGACGCTTTAATGGGCTTAAACTTACCATCTGGGTAGACATCGAAATCAAATTA

>HGS058 Ribosomal Protein S10 (SEQ ID NO:38)

MAKQKIRIRLKAYDHRVIDQSAEKIVETAKRSGADVSGPIPLPTEKSVYTIIRAVHKYKDSREQFEQRTHKRLID
IVNPTPKTVDALMGLNLPSGV DIEIKL

>HGS059 Ribosomal Protein S14 (SEQ ID NO:39)

ATGGCTAAGAAATCTAAAATAGCAAAAGAGAGAAAAAGAGAAGAGTTAGTAAATAAATATTACGAATTACGTAAAGAGTTAAA
AGCAAAAGGTGATTACGAAGCGTTAAGAAAATTACCAAGAGATTACACCTACACGTTTAACTAGAAGATGTAAAGTAACTG
GAAGACCTAGAGGTGATTACGTAAATTTGAAATGTCCTGATTGCGTTTAGAGAACATGCGCACAAAGGACAAATTCAGGT
GTTAAAAAATCAAGTTGG

>HGS059 Ribosomal Protein S14 (SEQ ID NO:40)

MAKKSIAKERKREELVNKYELRKELKAKGDYEALRLPRDSSPTRLTRRCKVTGRPRGVLRKFEMSRIAFREH
AHKGQIPGVKKSSW

>HGS060 Ribosomal Protein S19 (SEQ ID NO:41)

ATGGCTCGTAGTATTAAAAAAGGACCTTTCGTCGATGAGCATTTAATGAAAAAAGTTGAAGCTCAAGAAGGAAGCGAAAAGAA
ACAAGTAATCAAAACATGGTCACGTCGTTCTACAATTTCCCTAATTTTCATCGACATACTTTTGCAGTATACGACGGACGTA
AACACGTACCTGTATATGTAAGTATGGTAGGTCATAAATTAGGTGAGTTTGCTCCTACTCGTACATTCAAAGGACAC
GTTGCAGACGACAAGAAAACAAGAAGA

>HGS060 Ribosomal Protein S19 (SEQ ID NO:42)

MARSIKKGPVDEHLMKKVEAQEGSEKKQVIKTWSRRSTIFPNFIGHTFAVYDGRKHVPVYVTEDMVGHKLGEFA
PTRTFKHVADDDKTRR

>HGS062 Ribosomal Protein S14 Homolog (SEQ ID NO:43)

ATGGCTAAAACCTCAATGGTTGCTAAGCAACAAAAAACAATAATATGCAGTTTCGTGAATACACTCGTTGTGAACGTTGTGG
CCGTCCACATTCTGTATATCGTAAATTTAAATTATGCCGTATTGTTTCCGTGAATTAGCTTACAAAGGCCAAATCCCTGGCG
TTCGTAAAGCTAGCTGG

>HGS062 Ribosomal Protein S14 Homolog (SEQ ID NO:44)

MAKTSMVAKQKKQKYAVREYTRCERCGRPHSVYRKFKLCRICFRELAYKGQIPGVRKASW

>HGS064 YycF (SEQ ID NO:45)

ATGGCTAGAAAAGTTGTTGTAGTTGATGATGAAAAACCGATTGCTGATATTTAGAATTTAACTTAAAAAAGAAGGATACGA
TGTGTACTGTGCATACGATGGTAATGATGCAGTCGACTTAATTTATGAAGAAGAACCAGACATCGTATTACTAGATATCATGT
TACCTGGTCGTGATGGTATGGAAGTATGTCGTGAAGTGCGCAAAAAATACGAAATGCCAATAATAATGCTTACTGCTAAAGAT
TCAGAAATTGATAAAGTGCTTGGTTTAGAACTAGGTGCAGATGACTATGTAACGAAACCGTTTAGTACGCGTGAATTAATCGC
ACGTGTGAAAGCGAACTTACGTCGTCTAATCTACAACCAGCACAAAGACTGGAAATGTAACGAATGAAATCACAAATTAAG
ATATTGTGATTTATCCAGACGCATATTCTATTAATAAAACGTCGCGAAGATATTGAATTAACACATCGTGAATTTGAATTGTTC
CATTATTATCAAAACATATGGGACAAGTAATGACACGTGAACATTTATTACAAACAGTATGGGGCTATGATTACTTTGGCGA
TGTACGTACGTCGATGTAACGATTTCGTTTACGTGAAAAGATTGAAGATGATCCGTCACATCCTGAATATATTGTGACGC
GTAGAGGCGTTGGATATTTCTCCAACAACATGAG

>HGS064 YycF (SEQ ID NO:46)

MARKVVVVDEKPIADILEFNLKKEGYDVYCAVDGNDVLDLIYEEPDIVLLDIMLPGRDGMVCREVRKKYEMP
IIMLTAKDSEIDKVLGLGELGADDYVTKPFSTRELIARVKANLRRHYSQPAQDTGNVTNEITIKDIVIYPDAYSIK
KRGEDIETHREFELPHYLSKHMGMQVMTREHLLQTVWGYDYFGDVRTVDVTIRRLREKIEDDPSPHEYIVTRRGV
GYFLQQHE

>HGS063 (SEQ ID NO:47)

ATGCCATTATTTTACAACCAATTTAAAAACAAAATTATGGGGCGGTCAACGTCTAAGTGAGTTTGATATCAATTAGACAA
TGATACAACCTGGGGGAATGTTGGTGTGTGTCAGCACATCCAAATGGTACGAGCGAGATTATTAATGGACCATATCAAGGTCAA

ACATTAGACCGTATTGGTCAGAACATCGTGAATTGTTGGTGATTTCCTCAAGCAAAGATTTCCGCTTCTAACTAAAATAGT
GGATGCAAGAGAATCACTTTCTATTTCATGTGCACCCGTGATAATTCTTATGCTTATGAGCATGAAAACGGGCAATATGGCAAAT
CTGAATGTTGGTATATTATAGATGCAGAGAAGATGCAGAAATAGTTATAGGGACATTAGCAGAGTCTAGAGAAGAAGTTGCG
AATCATGTTCAACACGGAACGATAGAGTCGATACCTTAGATATATTAAAGTAAAACCTGGAGAATTCATTTTATTCCAGCAGG
AACAGTWCATACTATTTCTTCAGGAATATTAGCATACGAAACGATGCAATCGTCAGACATTACATATAGACTTTTATGATTTCA
ATCGTCAAGATAATCAATATAATGATAGACCGTTAAATATTGAAAAAGCTTTAGACGTTATTGAGTACAATGCACCATTACCT
AATATTTTGCCTGAAAGCGAAATTATTGAAAACCATAGTGATACACACATTGTATCGAATGATTTCTTTACATTGGTTAAATG
GGAAATTTCTGGCAGCTTAAATTATATGAAGCCTAGAGAGTTCTGTTTATGTTACAGTGTGGAAGGCGAAGGGCAAATGATTG
TCTATGGTGAAATTTCAAAGTACTACTGGTACAACTTTATTTGACTTCTGAAGATTGGATAGTGTCTTTGAAGGTGAT
TTCACATTGATGATTAGCTATGTG

>HGS063 (SEQ ID NO:48)

MPLFLQPIILKTKLWGGQRLSEFGYQLDNDTTEGECWCVSAHPNGTSEIINGPYQGQTLDRIWSEHRELFDFGDFPSKD
FPLLTKIVDARESLSIHVHPDINSYAYEHENGQYKSECWYIIDAEEDAEIVIGTLAESREEVANHVQHTIESIL
RYIKVKPGEFYFIPAGTVHTISSGILAYETMQSSDIYRLYDFNRQDNQYNDRLNIEKALDVIQYNAPLPNLP
ESEI IENHKCTHIVSNDFFTLVKWEISGTLNMYKPREFLVTVLEGGQMIVDGEIFKLTTGTNFILTSDDLDSV
FEGDFTLMISYV

>HGS065 (SEQ ID NO:49)

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ACGAACATAACAAGGATAAGCAGACTGCTTTTATCATTGAACAGTTAGAATTAGGTCTTGACGTTGCGCTCGTATCTGATGCT
GGATTGCCCTTAATTAGTGATCCTGGATACGAATTAGTAGTGGCAGCCAGAGAAGCTAATATTAAAGTAGAGACTGTGCCTGG
ACCTAATGCTGGGCTGACGGCTTTGATGGCTAGTGGATTACCTTCATATGTATATACATTTTATAGGATTTTGGCCACGAAAAG
AGAAAGAAAAAGTGCTGTATTAGAGCAACGTATGCATGAAAATAGCACATTAATTATATACGAATCACCGCATCGTGTGACA
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TGTAAGTATGATGTAAACACAATTACAAGCATTGATTACGCAAGGCGATGTACCATTGAAAGGCGAATTCGTTATCTTAATTG
AAGGTGCTAAAGCGAACAATGAGATATCGTGGTTTGTATGATTATCTATCAATGAGCATGTTGATCATTATATTCAAACCTCA
CAGATGAAACCAAAACAAGCTATTAAAAAGTTGCTGAAGAACGACAACCTAAAACGAATGAAGTATATAATATTATCATCA
AATAAGT

>HGS065 (SEQ ID NO:50)

MAVLYLVGTPIGNLADITYRAVDVLKRVDMIACEDTRVTSKLCNHYDIPTPLKSYHEHNKDKQTAFIIEQLELGL
DVALVSDAGLPLISDPGYELVVAAREANIKVETVPGPNAGLTALMASGLPSYVYTFGLFLPRKEKEKSAVLEQRM
HENSTLI IYESPHRVTDLTAKIDATRQVSLGRELTKKFEQIVTDDVTQLQALIQQGDVPLKGEFVILIEGAK
ANNEISWFDLDSINEHVDHYIQTSMKPKQAIKKVAEERQLKTNVYNIYHQIS

>HGS066 (SEQ ID NO:51)

ATGAAATTGGAAAAACAATCGCAGTAGTATTAGCATCTAGTGTCTGCTTGCAGGATGTACTACGGATAAAAAAGAAATTA
GGCATATTTAAAGCAAGTGGATAAAATTAAGATGATGAAGAACCAATTAAGTCTGTTGTAAGAAATTTGCTGAATTAGATG
AGAAAAAGAAAAATTAAGTGAAGATGTCAATAGTAAGATACAGCAGTTGCGCGTAAAGCAGTAAAGGATTTAATTAAAAAT
GCGGATGATCGTCTAAAGGAATTTGAAAAAGAAGAAGACGCAATTAAGAAGTCTGAACAAGACTTTAAGAAAGCAAAAGTCA
CGTTGATAACATTGATAATGATGTTAAACGTAAAGAAGTAAACAATTAGATGATGATTAAAAAGAAAAATATAAGTTACACA
GTGATTACGCGAAAGCATATAAAAAAGGCTGTAAACTCAGAGAAAACATTATTAAATATTAAATCAAATGACGCGACACAA
CAAGGTGTTAAACGAAAAATCAWAGCAATAGAACAGAACTATAAAAAAGTTAAAGAAGTATCAGATAAGTATACAAAAGTACT
AAATAAGGTTGGTAAAGAAAAGCAAGACGTTGATCAATTTAAA

>HGS066 (SEQ ID NO:52)

MKFGKTIIVVLLASSVLLAGCTTDKKEIKAYLKQVDKIKDDEEPIKTVGKKIAELDEKKKKLTEDVNSKDTAVRGK
AVKDLIKNADDRLKEFEKEEDAIIKKSEQDFKKAKSHVDNIDNDVKKRKEVKQLDDVLKEKYKLHSDYAKAYKKAVN
SEKTLFKYLNQNDATQQGVNEKSXAIEQNYKKLKEVSDKYTKVLNKGKQDQVDFK

>HGS067 (SEQ ID NO:53)

ATCGAGGACAGAATATTGTTAAAGTATGAACATATTGCTAAGCAGCTTAATGCGTTTATACATCAATCTAATTTCAAACCCGG
TGATAAATTGCCAAGCGTGACGCAATTAAGAGAAGCTTATCAAGTAAGTAAAGTACTATCATTAAAGCATTAGGCTTATTGG
AACAGATGGTTTTGATCTATCAAGCACAGGCGAGTGGTATTTATGTGAGAAATATTGCTGATGCCAATCGTATCAACGCTCTT
AAGACTAATGGTTTCTCTAAAGTTTTAGGTGAACACCGAATGACAAGTAAGGTACTTGTTTTTAAGGAGATTGCAACGCCACC
TAAATCTGTACAAGATGAGCTCCAATTAAATGCAGATGATACCGTCTACTATTTAGAGCGATTAAAGATTCTGTGGACGATGATG
TTTTATGTATCGAATATTCTTATTATCAAAAGAAATCGTGAAATATTAAATGATGATATTGCTAAGGGCTCTATCTTCGAC
TATTTAGAATCAAACATGAACTTCGTATTGGTTTTTCAGATATTTCTTAAATGTAGATCAACTCAAGTGAAGCTTC

ATTACTACAATTGTCTACAGGTGAACCATGTTTACGTTACCACCAGACTTTTTATACAATGACTGGCAAACCCTTTGATTCAT
CTGACATCGTATTTTATTATCGTCATGCACAGTTTTATATTCTTAGTAAAAAG

>HGS067 (SEQ ID NO:54)

IEDRILLKYEHIKQLNAFIHQSNFKPGDKLPVSVTLKERYQVSKSTIIKALGLLEQDGLIYQAQSGSIYVRNIA
DANRINVFKNFGSKSLGEHRMTSKVLVFKEIATPPKSVQDELQLNADDTVYYLERLRFVDDDLVLCIEYSYYHKE
IVKYLNDIDAKGSIFDYLESNMKLRIGFSDIFFNVDQLTSSEASLLQLSTGEPCLRYHQTFYMTGKPFSSDIV
FHYRHAQFYIPSKK

>HGS068 (SEQ ID NO:55)

ATGACTGTAGAATGGTTAGCAGAACAATTAAAAGAACAATAATTCAATTAAGTGAAGTCAAAAAACAACAGTTTCAAACATA
TTATCGTTTACTTGTGAATGGAATGAAAAGATGAATTTGACAAGTATTACAGATGAACACGATGTATATTTGAAACATTTT
ATGATTCATTGCACCTAGTTTTTATTTGATTTTAAATCAGCCTATAAGTATATGTGATGTAGGCGCTGGAGCTGTTTTCCA
AGTATTCGGTTAAAAATAATGTTCCGCAGTTAAAAGTGACGATTGTTGATTCATTAAATAAGCGTATTCAATTTTAAACCA
TTAGCGTCAGAATTACAATTACAGGATGTCAGCTTTATACACGATAGAGCAGAAACATTTGGTAAGGGTGTCTACAGGGAGT
CTTATGATGTTGTTACTGCAAGAGCAGTAGCTAGATTATCCGTGTTAAGTGAATGTGTTTACCGCTAGTTAAAAAAGGTGGA
CAGTTTGTGTCATTAAATCTTCAAAAGGTGAAGAAGAATTAGAAGAAGCAAAATTTGCAATTAGTGTGTTAGGTGGTAATGT
TACAGAAAAACATACCTTTGAATTGCCAGAAGATGCTGGAGAGCGCCAGATGTTTATTGATAAAAAAAGACAGACGCCGA
AAAAGTATCCAAGAAAACAGGGACGCTAATAAGACTCCTTTACTTGA AAAA

>HGS068 (SEQ ID NO:56)

MTVEWLAELKEHNIQLTETQKQFQTYRLLVEWNEKMNLTSTDEHDVYLKHFYDSIAPSFYFDENQPIISICD
VGAGAGFPSIPLKIMFPQLKVTIVDSLNRKIQFLNLHASELQLQDVSIHRAETFGKGVYRESYDVVTARAVAR
LSVLSELCLPLVKKGGQFVALKSSKGEELKEAKFAISVLGGNVTEHTFELPEDAGERQMFIIKKRQTPKKYP
RKPGETPNKTPLEK

>HGS069 (SEQ ID NO:57)

ATGGCACATACCATTACGATTGTTGGCTTAGGAACTATGGCATTGATGATTGCGGCTAGGGATATATAAATTTTAAAGAC
ACAAGATAAAGTTTATGCAAGAACGTTAGATCATCCAGTTATAGAAATCATTGCAAGATGAATTAACATTTTCAAGTTTTGACC
ATGTTTATGAAGCACATAACCAATTTGAAGATGTCTATATTGATATTGTGGCGCAATTGGTTGAAGCTGCTAATGAAAAAGAT
ATTGTCTATGCGGTTCCGGTCATCCTAGAGTTGCTGAGACAACACTACAGTGAATTAAGTGGCTTTAGCAAAGGACAATACTGA
TATAGATGTGAAGTTTATAGGTGGGAAAAGCTTTATTGATGATGTGTTGAAGCAGTTAATGTAGATCCAAATGATGGCTTCA
CACTGTTAGATGCGACATCATTACAAGAAGTAACACTTAATGTTAGAAGCATACATTGATTACGCAAGTTTATAGTGCAATG
GTTGCTGCTAATTTGAAAATCACTTTAATGGAACGATATCCTGATGATTACCTGTTTCAAATGTCACTGGTGACGAAGCGA
TGGTGCGGATAACGTTGTGACATGCCCATATATGAATGGATCATGATGAAAATGCATTCAATAATTTGACGAGTGATTTGCG
TACCAAAAATCATAACATCGACATATTTGTATCATGACTTTGATTTGCAACGGAAGTGATTGATCTTTAGTTGATGAAGAT
AAAGTTTGTCCATGGGATAAAGTGCAACGCGATGAAGCTAAAGCGTTATTTACTTGAAGAAACATTTGAATGTTTGAAGC
TATTGACAATGAAGATGATTGGCATATGATTGAAGAACTAGGAGATATTTTATTACAAGTGTTATTGCATACACTAGTATTGGTA
AAAAAGAAGGGTATATCGACATTAAAGAAGTGATTACAAGCTTAAATGCTAAAATGATTGCTAGACACCCACACATATTTGGT
GATGCCAATGCTGAACTATCGATGACTTAAAGAAATTTGGTCTAAGGCGAAAGATGCTGAAGGTAACACAGCCAAGAGTTAA
ATTTGAAAAAGTATTTGCAGAGCATTTTAAATTTATATGAGAAGACGAAGGATAAGTCATTTGATGAGGCCGCGTTAAAGC
AGTGGCTAGAAAAAGGGGAGAGTAATACA

>HGS069 (SEQ ID NO:58)

MAHTITIVGLGNYGIDDLPLGIYKFLKTQDKVYARTLDHPVIESLQDELTFQSFHDVYEAHNQFEDVYIDIVAQL
VEANEKDIVYAVPGHPRVAETTTVKLLALAKDNTDIDVKVLGGSFIDDVFEAVNVDPNDFLLDATSLQEV
LNVRTHTLITQVYSAMVAANLKITLMERYPDDYPVQIVTGARS DGADNVVTCPLYELDH DENAFNNLT SVFVPKI
ITSTYLHDFDFATEVIDTLVDEDKGCPWDKVQTHXLT LKRYLLEETFEIDNEDDWHMIEELGDILLQVLLH
TSIGKKEGYIDIKEVITSLNAKMIRHPHIFGDANAETIDDLKEIWSKAKDAEGKQPRVKFEKVFAEHFLNLYEK
TKDKSFDEAALKQWLEKGESNT

>HGS070 (SEQ ID NO:59)

AATGTAAATCATTTCTAATAAAACGACAACCTGTGTCTTCTTTACTTGTATATGTTACATATATTTACGATAGAGAGGATAAGAA
AATGGCTCAAATTTCTAAATATAAACGTTAGTTTGTAACTAAGTGGTGAAGCGTTAGCTGGAGAAAAAGGATTTGGCATAA
ATCCAGTAATTATTAAGTGTGCTGAGCAAGTGGCTGAAGTTGCTAAAATGGACTGTGAAATCGCAGTAATCGTTGGTGGC
GGAACATTTGGAGAGGTAAAACAGGTAGTGACTTAGGTATGGACCGTGAAGTCTGATTACATGGGTATGCTTGAACCTGT
AATGAATGCCTTAGCATTACAAGATAGTTTAGAACAATTTGGATGTGATACACGAGTATTAACATCTATTGAAATGAAGCAAG
TGGCTGAACCTTATATTCGTCGTCGTCGAATTAGACACTTAGAAAAGAAACGCGTAGTTATTTTGTGTCAGGTATTGGAAC
CCATCTTCTCTACAGATACACTACGCGCTTACGTGCTGCAGAAGTTGAAGCAGATGTTATTTAATGGGCAAAAATAATGT
AGATGGTGTATATCTGCAGATCCTAAAGTAAACAAAGATGCGGTAAAATATGAACATTTAACGCATATTCAAATGCTTCAAG

AAGGTTTACAAGTAATGGATTCAACAGCATCCTCATTCTGTATGGATAATAACATTCCGTTAACTGTTTTCTCTATTATGGAA
GAAGGAAATATTAAACGTGCTGTTATGGGTGAAAAGATAGGTACGTTAATTACAAAA

>HGS070 (SEQ ID NO:60)

NVNHSNKTITVSSLLVYVYIHDREDKKMAQISKYKRVVLKLSGEALAGEKGFGINPVIKSVAEQVAEVAKMDC
EIAVIVGGGNWVRGKTGSDLGMDRGTDYMGMLATVMNALALQDSLEQLDCDTRVLTISIEMKQVAEPYIRRAIR
HLEKKRVVIFAAGIGNPYFSTDTTAALRAAEVEADVILMGKNVNDGVYSADPKVNKDAVKYEHLLTHIQMLQEGLO
VMDSTASSFCMDNNIPLTVFSIMEEGNIKRAVMGEKIGTLITK

>HGS071 D-alanyl-alanine ligase (DdlA) (SEQ ID NO:61)

ATGACAAAAGAAAATATTTGTATCGTTTTTGGAGGGAAAAGTGCAGAACACGAAGTATCGATTCTGACAGCACAAAATGTATT
AAATGCAATAGATAAAGACAAATATCATGTTGATATCATTATATTACCAATGATGGTGTAGGAGAAAAGCAAAATAATATTA
CAGCTGAAATTAATCTACTGATGAGCTTCATTTAGAAAAATGGAGAGGCGCTTGAGATTTACAGCTATTGAAAGAAAGTAGT
TCAGGACAACCATACGATGCGATTATCCATTATTACATGGTCTTAATGGTGAAGATGGCAGGATTCAAGGGCTTTTTGAAGT
TTTGGATGTACCATATGTAGGAAATGGTGTATTGTCAGCTGCAAGTTCTATGGACAACTTGTAATGAAACAATTATTTGAAC
ATCGAGGGTTACACAGTTACCTTATATTAGTTCTTACGTTCTGAATATGAAAAATATGAACATAACATTTTAAATTAGTA
AATGATAAATTAATTAACCCAGTCTTTGTTAAACCTGCTACTTAGGTCAGTGTAGGTATCAGTAAATGTAATAATGAAGC
GGAACCTAAAGAAGGTATTAAGAAGCATTCCAATTTGACCGTAAGCTTGTTATAGAACAAGGCGTTAACGCACGTGAAATTG
AAGTAGCAGTTTATGAAATGACTATCCTGAAGCGACATGGCCAGGTGAAGTCGTAAGATGTCGCGTTTTACGATTACAAA
TCAAAATATAAGATGGTAAGGTTCAATTACAAATCCAGCTGACTTAGACGAAGATGTTCAATTAACGCTTAGAAATATGGC
ATTAGAGGCATTCAAAGCGACAGATTGTTCTGGTTTAGTCCGTGCTGATTTCTTTGTAACAGAGACAACCAATATATATTA
ATGAAACAAATGCAATGCCTGGATTTACGGCTTTCAGTATGTATCCAAGTTATGGGAAAATATGGGCTTATCTTATCCAGAA
TTGATTACAAACTTATCGAGCTTGCTAAAGAAGCTCACCAGGATAAACAGAAAAATAATACAAAATTGAC

>HGS071 D-alanyl-alanine ligase (DdlA) (SEQ ID NO:62)

MTKENICIVFGKSAEHEVSILTAQNVLNAIDKDKYHVDIYITNDGDWRKQNNITAEIKSTDELHLENGEALEI
SQLLKESSSGQPYDAVFPPLHGPNGEDGTIQGLFEVLDPVYVGNVLSAASSMDKLVKQLFEHRLPQLPYISF
LRSEYKEYEHNILKLVNDKLNYPVFKPANLGSSVGISKNNAEELKEGKEAFQFDRKLVEQGVNAREIEVAV
LGNDYPEATWPGEVVKDVAFYDYKSKYKDGKVLQIPADLDEDVQLTLRNMALFAFKATDCSGLVRADFFVTEDN
QIYINETNAMPGFTAFSMYPKLWENMGLSYPELITKLIELAKERHQDKQKNKYKID

>HGS072 Farnesyl diphosphatesynthase (IspA) (SEQ ID NO:63)

ATGACGAATCTACCGATGAATAAATTAATAGATGAAGTCAATAATGAATTATCGGTTGCGATAAATAAATCAGTAATGGATAC
TCAGCTAGAAGAAAGTATGTTGTATTCATTAATGCTGGAGGTAAACGCATCCGACCAAGTTCTGTTATTACTCACTTTAGATT
CACTAAATACCGAGTATGAGTTAGGTATGAAGAGCGCAATTGCACTAGAAATGATTTCATACATATTCATTATTATGATGAC
CTACCAGCGATGGATAATGATGATTATCGACGAGGAAAAATTAACAAATCATAAAGTATATGGTGAGTGGAAGTATGATAGC
AGGTGATGCTTTATTAATACTAAAGCATTGAACTTATTTCAAGTGATGATAGATTAACTGATGAAGTAAAAATAAAGTTCTAC
AACGGCTGTCAATAGCAAGTGGTTCATGTTGGAATGGTCCGCGGTCAAATGTTAGATATGCAAAGCGAAGGCCAACCAATTGAT
CTTGAAACTTTGGAAATGATACACAAACAAAAACAGGAGCATTATTAACCTTTTGCAGTTATGAGTGCAGCAGATATCGCTAA
TGTCGATGATACAACTAAAGAACATTGAGAAAGTTATAGTTATCATTTAGGTATGATGTTCCAGATTAAAGATGATTATTAG
ACTGCTATGGTGATGAAGCAAAGTTAGGTAAAAAGTGGGCAGCGATCTTGAAATAATAAAGTACGTACGTGAGTTTATTA
GGGAAAGATGGCGCAGAAAGATAAATTGACTTATCATAGAGACGCAGCAGTGATGAACAAACGCAATTTGATGAACAATTCAA
TACAAACACTTATTAGAAATCGTTGATTA

>HGS072 Farnesyl diphosphate synthase (IspA) (SEQ ID NO:64)

MTNLPMNKLIDEVNNELSVAINKSVMQTQLEESMLYSLNAGGKRIRPVLLLLLTLDSLNTYEYELGMKSAIALEMIH
TYSLIHDDLPAAMDNDYRRGKLTNHNKVGWETAILAGDALLTKAFELISSDRLTDEVKIKVLQRLSIA SGHVMG
VGGQMLDMQSEGQPIDLETLEMIHKTKTGALLTFAVMSAADIANVDDTKEHLESYSYHLGMMFQIKDDLDCYG
DEAKLGKKVGSDELNNKSTYVSLLGKDGAEKLTYHRDAVDELTDIDEQFNTKHLLEIVDL

>HGS073 Diphosphate Synthase (IspB) (SEQ ID NO:65)

TTTGTATTCTGAGTAGCAATTTGGCAAAGATGAACAAACGTCTGAACAAACGTATCAAGTTGCAGTCGCATTAGAGTTAAT
TCATATGGCAACACTTGTTTCATGATGACGTTATTGATAAAGCGACAAGCGTCGAGGCAAGTTAACCATATCAAAGAAATGGG
ATCAGACAACCTGCTATTTTAACTGGGAATTTTTTATTGGCATTAGGACTTGAACACTTAATGGCCGTTAAAGATAATCGTGTA
CATCAATTGATATCTGAATCTATCGTTGATGTTGTAGAGGGGAACCTTTCCAATTTCAAGACCAATTTAACAGTCAACAGAC
AATTATTAAATTTTACGACGTATCAATCGCAAAACAGCACTGTTAATTCAAATATCAACTGAAGTTGGTGCAATTACTTCTC
AATCTGATAAAGAGAGCTGACGAAAAATGAAAAATGATGGTCATTATATAGGTATGAGCTTCCAAATCATTGATGATGTATTA
GACTTCACAAAGTACCGGAAAGAAATAGGTAAAGCCGCTCGGAAGTATTTGCTTAATGGTCATATTACGTTACCGATTTATT
AGAAATGCGTAAAAATCCAGACTTCAAATTTGAAAAATCGAACAGTTACGTCGTGATAGTGAACGCAAGAAATTTGAAGAATGTA

TCCAAATCATTAGAAAATCTGACAGCATCGATGAGGCTAAGGCAGTAAGTTCTGAAGTATTTAAGTAAAGCCTTGAATTTGATT
TCyGAGTTACCAGATGGACATCCGAGATCACTACyTTTAAGTTTGACGAAAAAAATGGGTTCAAnAAACACG

>HGS073 Diphosphate Synthase (IspB) (SEQ ID NO:66)

FVILSSQFGKDEQTSEQTYQVALELIHMATLVHDDVIDKSDKRRGKLTISKKWDQTTAILTGNFLLALGLEHL
MAVKDNRVHQLISESIVDVCRGELFQFQDQFNSQQTIIINYLRRINRKTALLIQISTEVGAITSQSDKETVRKLKM
IGHYIGMSFQIIDVDLFTSTEEKLKGKPVGSDLLNGHITLPIILEMRKNPDKLKIIEQLRRDSEKFEFECIQII
RKSDSIDEAKAVSSKYLKALNLISELPDGHPRSLXLSTLTKMGSXNT

>HGS074 Undecaprenyl Pyrophosphate Synthetase (UppS) (SEQ ID NO:67)

GTAAATTATATTATGAATTTGCTGTCAATTTCTTAAAGACATTTTACCGGAACCTAATTGAAAAAATGTCAAAGTTGAAAC
AATTGGATTTACTGATAAGTTGCCAAAATCAACGATAGAAGCAATTAATAATGCyMAAGAAAAGACAGCTAATAATACCGCT
TAAAAATTAATATTGCAATTAATTATGGTGGCAGAGCAGAACTTGTTTCATAGTATTAAAAATATGTTTGACGAGCTTCATCAA
CAAGGTTTAAATAGTATGATCATAGATGAAACATATATAAACCAATCAATTAATGACAAAAGACTATCCTGATCCAGAGTTGTT
AATTCGTACTTCAGGAGAACAAAGAATAAGTAATTTCTTGATTGGCAAGTTTCGTATAGTGAATTTATCTTTAATCAAAAAT
TATGGCTGACTTTGACGAAGATGAATTAATTAATGTATAAAAATTTATCAGTCACGTCAAAGACGCTTTGGCGGATTGAGT
GAGGAG

>HGS074 Undecaprenyl Pyrophosphate Synthetase (UppS) (SEQ ID NO:68)

VNYIMNLPVNFLKTLPELIEKNVKVETIGFTDKLPKSTIEAINNAXEKTANNTGLKLIFAINYGGRAELVHSIK
NMFDELHQQLNSDIIIDETYINNHLMTKDYPPELLIRTSGEQRISNFLIWQVSYSEFIFNQKLWPDFDEDELIX
CIKIYQSRQRRFGLSEE

>HGS075 YycG (SEQ ID NO:69)

ATGAAGTGGCTAAAACAACCTACAATCCCTTCATACTAAATTTGTAATTGTTTATGTATTACTGATTATCATTGGTATGCAAT
TATCGGGTTATATTTTACAAATAACCTTGAAAAAGAGCTGCTTGATAATTTAAGAAGAATATTACGCAGTACCGGAAACAAT
TAGAAATTAGTATTGAAAAAGTATATGACGAAAAGGGCTCCGTAAATGCACAAAAAGATATTCAAAATTTATTAAGTGAGTAT
GCCAACCGTCAAGAAATTGGAGAAATTCGTTTTATAGATAAAGACCAAAATTATTATTGCGACGACGAAGCAGTCTAACCGTAG
TCTAATCAATCAAAAAGCGAATGATAGTTCTGTCCAAAAGCACTATCACTAGGACAATCAAAAGATCATTTAATTTTAAAG
ATTATGGCGGTGGTAAGGACCGTGTCTGGGTATATAATATCCCACTTAAAGTCGATAAAAAGGTAATTGGTAATATTTATATC
GAATCAAAAATTAATGACGTTTATAACCAATTAATAATATAAATCAAAATATTCATTGTGGTACAGCTATTTTATTATTAAT
CACAGTCATCCTAGGATTCTTTATAGCGCGAAGCATTACCAACCAATCACCGATATGCGTAACCGAGACGGTCGAAATGTCCa
GAGGTAACCTATACGCAACGTGTGAAGATTTATGGTAATGATGAAATTGGCGAATTAGCTTTAGCATTTAATAACTTGTCTAAA
CGTGTACAAGAAGCGCAGGCTAATACTGAAAGTGAGAAACGTAGACTGGACTCAGTTATCACCCATAGAGTGATGGTATTAT
TGCAACAGACCCCGTGGACGTATTCTGTATCGTCAATGATATGGCACTCAAGATGCTTGGTATGGCGAAAGAAGACATCATCG
GATATTACATGTTAAGTGATTAAGTCTTGAAGATGAATTTAACTGGAAGAAATTCAAGAGAATAATGATAGTTTCTTTATTA
GATTTAAATGAAGAAGAAGGTCTAATCGCACGTGTTAACTTTAGTACGATTGTGCAGGAAACAGGATTTGTAACGTGGTTATAT
CGCTGTGTTACATGACGTAACGAACAACAAGTTGAACGTGAGCGTCTGTAATTTGTTGCCAATGTATCACATGAGTTAC
GTACACCTTTAACTTCTATGAATAGTTACATTGAAGCACTTGAAGAAGGTGCATGGAAGATGAGGAACTTCGCCACAAATTT
TTATCTGTACCCGTGAAGAAACAGAAAGATGATTGCACTGGTCAATGACTTGCTACAGTTATCTAAAATGGATAATGAGTC
TGATCAAAATCAACAAAGAAATTACGACTTTAACATGTTTCATTAATAAAATTATTAATCGACATGAAATGCTCGGAAAGATAC
AACATTTATTTCGAGATATTCGAAAAAGACGATTTTACAGAATTTGATCCTGATAAAATGACGCAAGTATTTGATAATGTCA
TTACAAATGCGATGAAATATTCTAGAGGCGATAAACGTGTCGAGTTCCACGTGAAACAAAATCCACTTTATAATCGAATGACG
ATTCGTATTAAAGATAATGGCATTGGTATTCCTATCAATAAAGTCGATAAGATATTGACCGATTCTATCGTGTAGATAAGGC
ACGTACGCGTAAATGGGTGGTACTGGATTAGGACTAGCCATTTGAAAGAGATTGTGGAAGCGCACAAATGGTCGTATTGGG
CAACAGTGTAGAAGGTCAAGGTACATCTATCTTTATCACACTTCCATGTGAAGTCATTGAAGACCGTGATTGGGATGAA

>HGS075 YycG (SEQ ID NO:70)

MKWLKQLQSLHTKFVIVYVLLIIIGMQIIIGLYFTNNLEKELLDNFKKNITQYAKQLEISIEKVYDEKGSVNAQKD
IQNLLSEYANRQEIGEIRFIDKDQIIIIATTKQSNRSLINQKANDSSVQKALSLGQSNHLLKDYGGGKDRVWVY
NIPVKVDKVKVIGNIYIESKINDVYNQLNNINQIFIVGTAISLLITVILGFFIARTITKPIIDMRNQTVEMSRGNY
TORVKIYGNDEIGELALAFNNLSKRVQEAQANTESEKRRLLDSVITHMSDGI IATDRGRIRIVNDMALKMLGMK
EDIIGYYMLSVLSLEDEFKLEEIQENNDSFLLDLNEEGLIARVNFSTIVQETGFTVGYIAVLHVDTEQQQVERE
RREFVANVSHELRTPLTSMNSYIEALEEGAWKDEELAPFLSVTREETERMIRLVNDLLQLSKMDNESDQINKEI
IDFMFINKI INRHEMSAKDTTFIRDIPKKTIFTEFPDKMTQVFDNVITNAMKYSRGDKRVEFHVKNPLYNRM
TIRIKDNGIGIPINKVDKI PDRFYRVDKARTRKMGGTGLGLAISKEIVEAHNGRIWANSVEGQGTSTIFITLPCEV
IEDGDWDE

>pbp1 (SEQ ID NO:71)

ATGGCGAAGCAAAAAATTAAATTAATAAATAAATAGGGGCGAGTCTACTTGTGGTTTATTTCGGACTGCTCTTTTTTAT
ATTGGTTTTAAGAAATTTTCATATATCATGATTACTGGACATTCTAATGGTCAAGATTTAGTCATGAAGGCAATGAAAAGTATT
TAGTTAAGAATGCACAACAACAGAACGAGGAAAGATATATGATCGTAATGGTAAAGTGCTAGCAGAAAGATGTAGAAAGATAT
AAACTTGTTCAGTAATAGATAAAAAAGGCGAGTGCCAATTCTAAAAAACCTAGGCATGTAGTTGATAAAAAAGAGACTGCAAA
GAAATTATCTACAGTCATTAATATGAAGCCAGAGGAAATGAAAAGAGACTTAGTCAAAAGAAAGCTTTCCAAATTGAATTTG
GACGCAAGGAACAAATTTAACGTATCAGGACAAATTTGAAAAATAGAGAAAATGAATTTGCCTGGTATTCTTTATTGCGCTGAA
ACAGAACGCTTTTATCCAAATGGCAATTTTGCATCACACTTAATTGGTAGAGCTCAGAAAAATCCGGATACTGGTGAACTTAA
AGGTGCACTTGGAGTTGAAAAGATTTTGTAGATTATTTAAGTGGATCTAAAGGATCATTGAGATATATTTCATGATATTTGGG
GATATATCGCACCATACTAAAAAGAGAAGCAGCCTAAACGTGGTGATGATGCCATTTAACAATCGATTCAAATATTCAA
GTATTTGTTGAAGAAGCTTTAGATGGCATGGTTGAAAGATACCAGCCGAAAGATTTATTTCGGTTGTATGGATGCCAAAAC
TGGAGAAATTTTAGCATACAGTCAGCGACCAACATTTAATCCTGAACTGGTAAAGACTTTGGTAAAAAGTGGGCAATGACC
TTTATCAAAACACATACGAGCCTGGATCAACATTTAATCATATGGGTTAGCAGCTGCTATTCAAGAAGGTGCTTTTGATCCT
GATAAGAAATATAAATCTGCACATAGATATTATGGGTTACGTTATTTCAGACTGGAATAGAGTCGGTGGGGTGAAATCCC
AATGTCACTCGGATTTACTTTATCTAATACATTGATGATGATTACAAGATTTAGTTGGTGCAAGCAAAAATGAAATCTT
GGTATGAACGATTGGATTGGAAAACTAAAGGTATGTTTGTATGGAGAAGCACCTGGTCAAATGGATGGAGTAATGAG
TTGCAACAAAAACGTATCATTTGGTCAATCGACAACAGTAACACCTGTTCAAATGTTACAAGCGCAATCAGCGTTCTTTAA
TGATGGTAATATGTTAAACCATGGTTTGTGAATAGCGTTGAAAATCCTGTTAGTAAAGACAATTTTATAAAGGGCAAAAAC
AAATCGCAGGCAAAACCAATAACAAAAGATACTGCTGAAAAAGTTGAAAAGCAATTGGATTAGTTGTGAATAGTAAGAAGAGT
CAGCTGCAAACTATCGTATTGATGGTTATGAGGTGCAAGGTAAAGTGGTACAGCACAAAGTCGCTGCACCTAATGGTGGTGG
ATACGTTAAAGGTCCAAACCCATATTTGTAAAGTTTGTGGGTTGACGCGCGCAAGAAAAATCCTAAAGTTATTGTATACGCTG
GTATGAGCTTGGCACAATAAAGGATGACCAAGAAGCTTATGAATTAGGTGTTAGTAAAGCGTTTAAACCAATAATGGAATACT
TTGAAATATTTAATGTAGGTAAATCAAAGATGACACATCTAATGCAGAGTATAGTAAAGTGCCAGATGTTGAAGGTCAAGA
CAAACAAAAAGCTATTGATAATGTGAGTGCAAAATCATTAGAACCAGTTACTATTGGTTCTGGCACACAATAAAGCACAAT
CTATAAAGCAGGGAATAAAGTCTTACCTCATAGTAAAGTACTGTTATTAACAGATGGAGACTTAACATATGCCTGACATGTCA
GGATGGACGAAGAAGATGTCATTGCTTTTGAACCTTAACAAATATTAAAGTAAATTTAAAGGTAGCGGTTTGTGTCCCA
CCAATCAATTAGTAAGGGACAAAACTTACTGAAAAAGATAAAATAGACGTAGAATTTTCATCAGAGAATGTAGACAGCAATT
CGACGAATAATTCTGATTCAAATTCAGATGATAAGAAGAAATCTGACAGTAAACTGACAAGGATAAGTCCGAC

>Pbp1 (SEQ ID NO:72)

MAKQKIKIKKNKIGAVLLVGLFGLFFILVLRISYIMITGHSNGQDLVMKANEKYLVKNAQQPERGKIYDRNGKV
LAEDVERYKLVAVIDKKASANSKKPRHVVDKKE TAKKLSTVINMKPEEIEKRLSOKKAFQIEFGRKGTNLTYQDK
LKIEKMNLPGISLLPETERFYPNGNFASHLIGRAQKNPDTELKALGVEKIFDSYLSGSKGSLRYIHDWGYIA
PNTKKEKQPKRGDDVHLTIDSNIOVFVEEALDGMVERYQKDLFAVMDAKTGEILAYSQRPTFNPETGKDFGKK
WANDLYQNTYEPGSTFKSYGLAAAIQEGAFDPDKKYKSGHRDIMGSRISDWNRVGWGEIPMSLGFTYSSNTLMMH
LQDLVGADKMSWYERFPGKSTKGMFDGEAPGQIGWSNELQOKTSSFGQSTTVTPVQMLQAQSAFFNDGNMLKP
WVFNVENPVSKRQFYKQKQIAGKPI TKDTAEKVEKQLDLVNSKKSHAANYRIDGYEVEGKTGTQAQAAPNGG
GYVKGPNPYFVFSMGDAPKKNPKVIVYAGMSLAQKNDQEAYELGVSKAFKPI MENTLKYLNVGKSKDDTSNAEYS
KVPDVEGQDKQKIDNVSAKSLEPVTIGSGTQIKAQSIKAGNKVLPKSVLLLTGDLTMDPSGWTKEDVIAFE
NLTNIKVNLKSGSFVSHQSIKSGQKLTEDKIDVEFSSENVDSNSTNNSDSNDDKKSDSKTDKDKSD

>dead (SEQ ID NO:73)

ATTCGCAAAATGCTTTATTGCGATTAAATTTTTTTGGTGGTACTATATAGAAGTTGATGAAATATTAATGAACCTTATATGCAA
AAGTATATTGAGAAATAACAGGTAAAAAGGAGAATTATTTTGCAAAATTTTAAAGAACTAGGGATTTCGGATAATACGGTTC
AGTCACTTGAATCAATGGGATTTAAAGAGCCGACACCTATCCAAAAGACAGTATCCCTTATGCGTTACAAGGAATTGATATC
CTTGGGCAAGCTCAAACCGGTACAGGTAAACAGGAGCATCGGTATTCTTTAATTGAGAAAAGTAGTAGGGAACAAGGGGT
TCAATCGTTTGATTTTAGCACCTACAAGAGAATTGGCAATGCAGGTAGCTGAACAATTAGAGAATTTAGCCGTGGACAAGGTG
TCCAAGTTGTTACTGTATTGCGGTGGTATGCCTATCGAACGCCAAATTAAGCCTTGAAAAAGGCCCAAAATCGTAGTCGGA
ACACCTGGGCGTGTATCGACCATTTAAATCGTCGCACATTAAAAACGGACGGAATTCATACTTTGATTTTAGTAGAGCTGA
TGAAATGATGAATATGGGATTCATCGATGATATGAGATTTATTATGGATAAAATTCAGCAGTACAACGTCAAACAATGTTGT
TCTCAGCTACAATGCCTAAAGCAATCCAAGCTTTAGTACAACAATTTATGAAATCACCAAAAATCATTAAGACAATGAATAAT
GAAATGTCTGATCCACAAATCGAAGAATCTATACAATTTGTTAAAGAATTAGAGAAATTTGATACATTTACAAATTTCTTAGA
TGTTCAATCAACCTGAATTAGCAATCGTATTTCGACGTACAAAACGTCGTGTTGATGAATTAACAAGTGCTTTGATTTCTAAAG
GATATAAAGCTGAAGGTTTACATGGTGATATTACACAAGCGAAACGTTTGAAGTATTAAAGAAATTTAAAAATGACCAAAAT
AATATTTTAGTCGTACTGATGTAGCAGCAAGAGGCTAGATATTTCTGGTGTGAGTCATGTTTATACTTTGATATACCTCA
AGATACTGAAAGCTATACACACCGTATTGGTTCGTACGGGTCGTGCTGGTAAAGAAGGTATCGCTGTAAACGTTGTTAAATCCAA
TCGAAATGGATTATATCAGACAAATTGAAGATGCAACGGTAGAAAAATGAGTGCACCTTCGTCCACCACATCGTAAAGAAAGTA
CTTCAAGCACGTGAAGATGACATCAAAGAAAAAGTTGAAAACCTGGATGTCTAAAGAGTCAGAATCAGCTTGAAACGCATTTC
TACAGAGTTGTTAAATGAATATAACGATGTTGATTTAGTTGCTGCACTTTTACAAGAGTTAGTAGAAGCAACGATGAAGTTG
AAGTTCAATTAACCTTTTGA AAAACCATTTATCTCGCAAGGCCGTAAACGGTAAACCAAGTGGTCTCGTAACAGAAATAGTAAG
CGTGGTAATCCTAAATTTGACAGTAAGAGTAAACGTTCAAAAGGATACTCAAGTAAGAAGAAAAGTACAAAAAATTCGACCG

TAAAGAGAAGAGCAGCGGTGGAAGCAGACCTATGAAAGGTGCGACATTTGCTGACCATCAAAAATAATTTATAGATTAAGAGC
TTAAAGATGTAATGTCT

>DeaD (SEQ ID NO:74)

NINELICKSILRNKQVKRRIILQNFKELGISDNTVQSLESMGFKEPTPIQKDSIPYALQGIDILGQAQTGTGKTG
AFGIPLIEKVVGKQGVQSLILAPTRELAMQVAEQLREFSRGQGVVTVFGGMPIERQIKALKKGPQIVVGTPGR
VIDHLNRRTLKTDGIHTLILDEADEMMNMGFIDDMRFIMDKIPAVQRQTMLEFSATMPKAIQALVQQFMKSPKIIK
TMNNEMSDPQIEEFYTIVKELEKFDFTFNFLDVHQPELAIVFGRTKRRVDELTSALISKGYKAEGHLHGDITQAKR
LEVLKKFKNDQINILVATDVAARGLDISGVSHVYNFDIPQDTESYTHRIGRTGRAGKEGIAVTFVNP IEMDYIRQ
IEDANGRKMSALRPPHRKEVLQAREDDIKEKVENWMSKESESRLKRISTELLNEYNDVDLVAALLQELVEANDEV
EVQLTFEKPLSRKGRNGKPSGSRNRNSKRCNPKFDSKSKRSKGYSSKKKSTKKFDRKEKSSGSRPMKGRFTADH
Q

Nucleic acid molecules of the present invention may be in the form of RNA, such as mRNA, or in the form of DNA, including, for instance, DNA and genomic DNA obtained by cloning or produced synthetically. The DNA may be double-stranded or single-stranded. Single-stranded DNA or RNA may be the coding strand, also known as the sense strand, or it may be the non-coding strand, also referred to as the anti-sense strand.

The present invention further encompasses nucleic acid molecules of the present invention that are chemically synthesized, or reproduced as peptide nucleic acids (PNA), or according to other methods known in the art. The use of PNAs would serve as the preferred form if the nucleic acid molecules of the invention are incorporated onto a solid support, or gene chip. For the purposes of the present invention, a peptide nucleic acid (PNA) is a polyamide type of DNA analog and the monomeric units for adenine, guanine, thymine and cytosine are available commercially (Perceptive Biosystems). Certain components of DNA, such as phosphorus, phosphorus oxides, or deoxyribose derivatives, are not present in PNAs. For general review, see, e.g., P. E. Nielsen, M. Egholm, R. H. Berg and O. Buchardt, *Science* 254, 1497 (1991); and M. Egholm, O. Buchardt, L. Christensen, C. Behrens, S. M. Freier, D. A. Driver, R. H. Berg, S. K. Kim, B. Norden, and P. E. Nielsen, *Nature* 365, 666 (1993), hereby incorporated by reference herein.

PNAs bind specifically and tightly to complementary DNA strands and are not degraded by nucleases. In fact, a PNA binds more strongly to DNA than does DNA itself. This is probably because there is no electrostatic repulsion between the two strands, and also the polyamide backbone is more flexible. Because of this, PNA/DNA duplexes bind under a wider range of stringency conditions than DNA/DNA duplexes, making it easier to perform multiplex hybridization. Smaller probes can be used than with DNA due to the strong binding. In addition, it is more likely that single base mismatches can be determined with PNA/DNA hybridization because a single mismatch in a PNA/DNA 15-mer lowers the melting point (T^m) by 8°-20° C, vs. 4°-16° C for the DNA/DNA 15-mer duplex. Also, the absence of charge groups in PNA means that hybridization can be done at low ionic strengths and reduce possible interference by salt during the analysis.

By "isolated" polynucleotide sequence is intended a nucleic acid molecule, DNA or RNA, which has been removed from its native environment. This includes segments of DNA comprising the *S. aureus* polynucleotides of the present invention isolated from the native chromosome. These fragments include both isolated fragments consisting only of *S. aureus* DNA and fragments comprising heterologous sequences such as vector sequences or other

foreign DNA. For example, recombinant DNA molecules contained in a vector are considered isolated for the purposes of the present invention which may be partially or substantially purified to exclude RNA or heterologous DNA. Isolated polynucleotides may be at least 10%, 20%, 30%, 40%, 50%, 60%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99%, or 100% pure relative to heterologous polynucleotides (e.g., DNA or RNA) or relative to all materials and compounds other than the carrier solution. Further examples of isolated DNA molecules include recombinant DNA molecules introduced and maintained in heterologous host cells or purified (partially or substantially) DNA molecules in solution. Isolated RNA molecules include *in vivo* or *in vitro* RNA transcripts of the DNA molecules of the present invention. Isolated nucleic acid molecules according to the present invention further include such molecules produced synthetically which may be partially or substantially purified. The term "isolated" does not refer to genomic or cDNA libraries, whole cell mRNA preparations, genomic DNA digests (including those gel separated by electrophoresis), whole chromosomes, or sheared whole cell genomic DNA preparations or other compositions where the art demonstrates no distinguishing features of the polynucleotides sequences of the present invention.

In addition, isolated nucleic acid molecules of the invention include DNA molecules which comprise a sequence substantially different from those described above but which, due to the degeneracy of the genetic code, still encode a *S. aureus* polypeptides and peptides of the present invention (e.g., polypeptides of Table 1). That is, all possible DNA sequences that encode the *S. aureus* polypeptides of the present invention. This includes the genetic code and species-specific codon preferences known in the art. Thus, it would be routine for one skilled in the art to generate the degenerate variants described above, for instance, to optimize codon expression for a particular host (e.g., change codons in the bacterial mRNA to those preferred by a mammalian or other bacterial host such as *E. coli*).

The invention further provides isolated nucleic acid molecules having the nucleotide sequence shown in Table 1 or a nucleic acid molecule having a sequence complementary to one of the above sequences. Such isolated molecules, particularly DNA molecules, are useful as probes for gene mapping and for identifying *S. aureus* in a biological sample, for instance, by PCR or hybridization analysis (e.g., including, but not limited to, Northern blot analysis). In specific embodiments, the polynucleotides of the present invention are less than 300kb, 200kb, 100kb, 50kb, 10kb, 7.5kb, 5kb, 2.5kb, and 1kb. In another embodiment, the polynucleotides comprising the coding sequence for polypeptides of the present invention do not contain genomic flanking gene sequences or contain only genomic flanking gene sequences having regulatory control sequences for the said polynucleotides.

In further embodiments, polynucleotides of the invention comprise at least 15, at least

30, at least 50, at least 100, or at least 250, at least 500, or at least 1000 contiguous nucleotides comprising the coding sequence for polypeptides of the present invention, but consist of less than or equal to 1000 kb, 500 kb, 250 kb, 200 kb, 150 kb, 100 kb, 75 kb, 50 kb, 30 kb, 25 kb, 20 kb, 15 kb, 10 kb, or 5 kb of genomic DNA that flanks the 5' or 3' coding nucleotide set forth in Table 1. In further embodiments, polynucleotides of the invention comprise at least 15, at least 30, at least 50, at least 100, or at least 250, at least 500, or at least 1000 contiguous nucleotides comprising the coding sequence for polypeptides of the present invention. In another embodiment, the nucleic acid comprising coding sequence for polypeptides of the present invention does not contain coding sequences of a genomic flanking gene (i.e., 5' or 3' to the Table 1 sequences in the genome). In other embodiments, the polynucleotides of the invention do not contain the coding sequence of more than 1000, 500, 250, 100, 50, 25, 20, 15, 10, 5, 4, 3, 2, or 1 genomic flanking gene(s).

The present invention is further directed to nucleic acid molecules encoding portions or fragments of the nucleotide sequences described herein. Uses for the polynucleotide fragments of the present invention include, but are not limited to, probes, primers, molecular weight markers and expressing the polypeptide fragments of the present invention. Fragments include portions of the nucleotide sequences of Table 1, at least 10 contiguous nucleotides in length selected from any two integers, one of which representing a 5' nucleotide position and a second of which representing a 3' nucleotide position, where the first nucleotide for each nucleotide sequence in Table 1 is position 1. That is, every combination of a 5' and 3' nucleotide position that a fragment at least 10 contiguous nucleotides in length could occupy is included in the invention as an individual species. "At least" means a fragment may be 10 contiguous nucleotide bases in length or any integer between 10 and the length of an entire nucleotide sequence minus 1. Therefore, included in the invention are contiguous fragments specified by any 5' and 3' nucleotide base positions of a nucleotide sequences of Table 1 wherein the contiguous fragment is any integer between 10 and the length of an entire nucleotide sequence minus 1.

The polynucleotide fragments specified by 5' and 3' positions can be immediately envisaged using the clone description and are therefore not individually listed solely for the purpose of not unnecessarily lengthening the specifications.

Although it is particularly pointed out that each of the above described species may be included in or excluded from the present invention. The above species of polynucleotides fragments of the present invention may alternatively be described by the formula "a to b"; where "a" equals the 5' nucleotide position and "b" equals the 3' nucleotide position of the polynucleotide fragment, where "a" equals as integer between 1 and the number of nucleotides of the polynucleotide sequence of the present invention minus 10, where "b"

equals an integer between 10 and the number of nucleotides of the polynucleotide sequence of the present invention; and where "a" is an integer smaller than "b" by at least 10.

Again, it is particularly pointed out that each species of the above formula may be specifically included in, or excluded from, the present invention.

Further, the invention includes polynucleotides comprising sub-genuses of fragments specified by size, in nucleotides, rather than by nucleotide positions. The invention includes any fragment size, in contiguous nucleotides, selected from integers between 10 and the length of an entire nucleotide sequence minus 1. Preferred sizes of contiguous nucleotide fragments include at least 20 nucleotides, at least 30 nucleotides, at least 40 nucleotides, at least 50 nucleotides, at least 60 nucleotides, at least 70 nucleotides, at least 80 nucleotides, at least 90 nucleotides, at least 100 nucleotides, at least 125 nucleotides, at least 150 nucleotides, at least 175 nucleotides, at least 200 nucleotides, at least 250 nucleotides, at least 300 nucleotides, at least 350 nucleotides, at least 400 nucleotides, at least 450 nucleotides, at least 500 nucleotides, at least 550 nucleotides, at least 600 nucleotides, at least 650 nucleotides, at least 700 nucleotides, at least 750 nucleotides, at least 800 nucleotides, at least 850 nucleotides, at least 900 nucleotides, at least 950 nucleotides, at least 1000 nucleotides, at least 1050 nucleotides, at least 1100 nucleotides, and at least 1150 nucleotides. Other preferred sizes of contiguous polynucleotide fragments, which may be useful as diagnostic probes and primers, include fragments 50-300 nucleotides in length which include, as discussed above, fragment sizes representing each integer between 50-300. Larger fragments are also useful according to the present invention corresponding to most, if not all, of the polynucleotide sequences of the sequence listing, shown in Table 1, or deposited clones. The preferred sizes are, of course, meant to exemplify not limit the present invention as all size fragments, representing any integer between 10 and the length of an entire nucleotide sequence minus 1 of the sequence listing or deposited clones, may be specifically included in or excluded from the invention. Additional preferred nucleic acid fragments of the present invention include nucleic acid molecules encoding epitope-bearing portions of the polypeptides (e.g., including but not limited to, nucleic acid molecules encoding epitope-bearing portions of the polypeptides which are shown in Table 4).

In another aspect, the invention provides an isolated nucleic acid molecule comprising a polynucleotide which hybridizes under stringent hybridization conditions to a portion of a polynucleotide in a nucleic acid molecules of the invention described above, for instance, nucleotide sequences of Table 1. By "stringent hybridization conditions" is intended overnight incubation at 42°C in a solution comprising: 50% formamide, 5x SSC (750 mM NaCl, 75 mM trisodium citrate), 50 mM sodium phosphate (pH 7.6), 5x Denhardt's solution, 10% dextran sulfate, and 20 µg/ml denatured, sheared salmon sperm DNA, followed by

washing the filters in 0.1x SSC at about 65°C. Hybridizing polynucleotides are useful as diagnostic probes and primers as discussed above. Portions of a polynucleotide which hybridize to a nucleotide sequence in Table 1, which can be used as probes and primers, may be precisely specified by 5' and 3' base positions or by size in nucleotide bases as described above or precisely excluded in the same manner. Preferred hybridizing polynucleotides of the present invention are those that, when labeled and used in a hybridization assay known in the art (e.g., Southern and Northern blot analysis), display the greatest signal strength with the polynucleotides of Table 1 regardless of other heterologous sequences present in equimolar amounts

The nucleic acid molecules of the present invention, which encode a *S. aureus* polypeptide, may include, but are not limited to, nucleic acid molecules encoding the full length *S. aureus* polypeptides of Table 1. Also included in the present invention are nucleic acids encoding the above full length sequences and further comprise additional sequences, such as those encoding an added secretory leader sequence, such as a pre-, or pro- or prepro-protein sequence. Further included in the present invention are nucleic acids encoding the above full length sequences and portions thereof and further comprise additional heterologous amino acid sequences encoded by nucleic acid sequences from a different source.

Also included in the present invention are nucleic acids encoding the above protein sequences together with additional, non-coding sequences, including for example, but not limited to non-coding 5' and 3' sequences. These sequences include transcribed, non-translated sequences that may play a role in transcription, and mRNA processing, for example, ribosome binding and stability of mRNA. Also included in the present invention are additional coding sequences which provide additional functionalities.

Thus, a nucleotide sequence encoding a polypeptide may be fused to a marker sequence, such as a sequence encoding a peptide which facilitates purification of the fused polypeptide. In certain preferred embodiments of this aspect of the invention, the marker amino acid sequence is a hexa-histidine peptide, such as the tag provided in a pQE vector (QIAGEN, Inc., 9259 Eton Avenue, Chatsworth, CA, 91311), among others, many of which are commercially available. For instance, hexa-histidine provides for convenient purification of the fusion protein. See Gentz et al. (1989) Proc. Natl. Acad. Sci. 86:821-24. The "HA" tag is another peptide useful for purification which corresponds to an epitope derived from the influenza hemagglutinin protein. See Wilson et al. (1984) Cell 37:767. As discussed below, other such fusion proteins include the *S. aureus* fused to Fc at the N- or C-terminus.

Variant and Mutant Polynucleotides

The present invention further relates to variants of the nucleic acid molecules which encode portions, analogs or derivatives of a *S. aureus* polypeptides of Table 1, and variant polypeptides thereof including portions, analogs, and derivatives of the *S. aureus* polypeptides. Variants may occur naturally, such as a natural allelic variant. By an "allelic variant" is intended one of several alternate forms of a gene occupying a given locus on a chromosome of an organism. See, e.g., B. Lewin, Genes IV (1990). Non-naturally occurring variants may be produced using art-known mutagenesis techniques.

Such nucleic acid variants include those produced by nucleotide substitutions, deletions, or additions. The substitutions, deletions, or additions may involve one or more nucleotides. The variants may be altered in coding regions, non-coding regions, or both. Alterations in the coding regions may produce conservative or non-conservative amino acid substitutions, deletions or additions. Especially preferred among these are silent substitutions, additions and deletions, which do not alter the properties and activities of a *S. aureus* protein of the present invention or portions thereof. Also preferred in this regard are conservative substitutions.

Such polypeptide variants include those produced by amino acid substitutions, deletions or additions. The substitutions, deletions, or additions may involve one or more residues. Alterations may produce conservative or non-conservative amino acid substitutions, deletions, or additions. Especially preferred among these are silent substitutions, additions and deletions, which do not alter the properties and activities of a *S. aureus* protein of the present invention or portions thereof. Also especially preferred in this regard are conservative substitutions.

The present invention also relates to recombinant vectors, which include the isolated nucleic acid molecules of the present invention, and to host cells containing the recombinant vectors, as well as to methods of making such vectors and host cells and for using them for production of *S. aureus* polypeptides or peptides by recombinant techniques.

The present application is directed to nucleic acid molecules at least 90%, 95%, 96%, 97%, 98%, 99% or 100% identical to a nucleic acid sequence shown in Table 1. The above nucleic acid sequences are included irrespective of whether they encode a polypeptide having *S. aureus* activity. This is because even where a particular nucleic acid molecule does not encode a polypeptide having *S. aureus* activity, one of skill in the art would still know how to use the nucleic acid molecule, for instance, as a hybridization probe or primer. Uses of the nucleic acid molecules of the present invention that do not encode a polypeptide having *S. aureus* activity include, *inter alia*, isolating an *S. aureus* gene or allelic variants thereof from

a DNA library, and detecting *S. aureus* mRNA expression in biological or environmental samples, suspected of containing *S. aureus* by hybridization analysis (e.g., including, but not limited to, Northern Blot analysis) or PCR.

For example, one such method involves assaying for the expression of a polynucleotide encoding *S. aureus* polypeptides in a sample from an animal host (e.g., including, but not limited to, human, bovine, rabbit, porcine, murine, chicken, and/or avian species). The expression of polynucleotides can be assayed by detecting the nucleic acids of Table 1. An example of such a method involves the use of the polymerase chain reaction (PCR) to amplify and detect *Staphylococcus* nucleic acid sequences in a biological or environmental sample.

The present invention also relates to nucleic acid probes having all or part of a nucleotide sequence described in Table 1 which are capable of hybridizing under stringent conditions to *Staphylococcus* nucleic acids. The invention further relates to a method of detecting one or more *Staphylococcus* nucleic acids in a biological sample obtained from an animal, said one or more nucleic acids encoding *Staphylococcus* polypeptides, comprising: (a) contacting the sample with one or more of the above-described nucleic acid probes, under conditions such that hybridization occurs, and (b) detecting hybridization of said one or more probes to the *Staphylococcus* nucleic acid present in the biological sample.

The invention also includes a kit for analyzing samples for the presence of members of the *Staphylococcus* genus in a biological or environmental sample. In a general embodiment, the kit includes at least one polynucleotide probe containing a nucleotide sequence that will specifically hybridize with a *S. aureus* nucleic acid molecule of Table 1 and a suitable container. In a specific embodiment, the kit includes two polynucleotide probes defining an internal region of the *S. aureus* nucleic acid molecule of Table 1, where each probe has one strand containing a 31' mer-end internal to the region. In a further embodiment, the probes may be useful as primers for polymerase chain reaction amplification.

The method(s) provided above may preferably be applied in a diagnostic method and/or kits in which *S. aureus* polynucleotides of Table 1 are attached to a solid support. In one exemplary method, the support may be a "gene chip" or a "biological chip" as described in US Patents 5,837,832, 5,874,219, and 5,856,174. Further, such a gene chip with *S. aureus* polynucleotides of Table 1 attached may be used to diagnose *S. aureus* infection in an animal host, preferably a human. The US Patents referenced above are incorporated herein by

reference in their entirety.

The present invention is further directed to nucleic acid molecules having sequences at least 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99% or 100% identical to the nucleic acid sequence shown in Table 1, which do, in fact, encode a polypeptide having *S. aureus* protein activity. By "a polypeptide having *S. aureus* activity" is intended polypeptides exhibiting activity similar, but not necessarily identical, to an activity of the *S. aureus* protein of the invention, as measured in a particular biological assay suitable for measuring activity of the specified protein. The biological activity of some of the polypeptides of the presents invention are listed in Table 1, after the name of the closest homolog with similar activity. The biological activities were determined using methods known in the art for the particular biological activity listed. For the remaining polypeptides of Table 1, the assays known in the art to measure the activity of the polypeptides of Table 2, sharing a high degree of identity, may be used to measure the activity of the corresponding polypeptides of Table 1.

Of course, due to the degeneracy of the genetic code, one of ordinary skill in the art will immediately recognize that a large number of the nucleic acid molecules having a sequence at least 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99%, or 100% identical to the nucleic acid sequences shown in Table 1 will encode a polypeptide having biological activity. In fact, since degenerate variants of these nucleotide sequences all encode the same polypeptide, this will be clear to the skilled artisan even without performing the above described comparison assay. It will be further recognized in the art that, for such nucleic acid molecules that are not degenerate variants, a reasonable number will also encode a polypeptide having biological activity. This is because the skilled artisan is fully aware of amino acid substitutions that are either less likely or not likely to significantly effect protein function (e.g., replacing one aliphatic amino acid with a second aliphatic amino acid), as further described below.

By a polynucleotide having a nucleotide sequence at least, for example, 95% "identical" to a reference nucleotide sequence of the present invention, it is intended that the nucleotide sequence of the polynucleotide is identical to the reference sequence except that the polynucleotide sequence may include up to five point mutations per each 100 nucleotides of the reference nucleotide sequence encoding the *S. aureus* polypeptide. In other words, to obtain a polynucleotide having a nucleotide sequence at least 95% identical to a reference nucleotide sequence, up to 5% of the nucleotides in the reference sequence may be deleted, inserted, or substituted with another nucleotide. The query sequence may be an entire sequence shown in Table 1, the ORF (open reading frame), or any fragment specified as described herein.

Other methods of determining and defining whether any particular nucleic acid

molecule or polypeptide is at least 90%, 95%, 96%, 97%, 98%, 99%, or 100% identical to a nucleotide sequence of the present invention can be done by using known computer programs. A preferred method for determining the best overall match between a query sequence (a sequence of the present invention) and a subject sequence, also referred to as a global sequence alignment, can be determined using the FASTDB computer program based on the algorithm of Brutlag et al. See Brutlag et al. (1990) Comp. App. Biosci. 6:237-245. In a sequence alignment the query and subject sequences are both DNA sequences. An RNA sequence can be compared by first converting U's to T's. The result of said global sequence alignment is in percent identity. Preferred parameters used in a FASTDB alignment of DNA sequences to calculate percent identity are: Matrix=Unitary, k-tuple=4, Mismatch Penalty=1, Joining Penalty=30, Randomization Group Length=0, Cutoff Score=1, Gap Penalty=5, Gap Size Penalty 0.05, Window Size=500 or the length of the subject nucleotide sequence, whichever is shorter.

If the subject sequence is shorter than the query sequence because of 5' or 3' deletions, not because of internal deletions, a manual correction must be made to the results. This is because the FASTDB program does not account for 5' and 3' truncations of the subject sequence when calculating percent identity. For subject sequences truncated at the 5' or 3' ends, relative to the query sequence, the percent identity is corrected by calculating the number of bases of the query sequence that are 5' and 3' of the subject sequence, which are not matched/aligned, as a percent of the total bases of the query sequence. Whether a nucleotide is matched/aligned is determined by results of the FASTDB sequence alignment. This percentage is then subtracted from the percent identity, calculated by the above FASTDB program using the specified parameters, to arrive at a final percent identity score. This corrected score is what is used for the purposes of the present invention. Only nucleotides outside the 5' and 3' nucleotides of the subject sequence, as displayed by the FASTDB alignment, which are not matched/aligned with the query sequence, are calculated for the purposes of manually adjusting the percent identity score.

For example, a 90 nucleotide subject sequence is aligned to a 100 nucleotide query sequence to determine percent identity. The deletions occur at the 5' end of the subject sequence and therefore, the FASTDB alignment does not show a matched/alignment of the first 10 nucleotides at 5' end. The 10 unpaired nucleotides represent 10% of the sequence (number of nucleotides at the 5' and 3' ends not matched/total number of nucleotides in the query sequence) so 10% is subtracted from the percent identity score calculated by the FASTDB program. If the remaining 90 nucleotides were perfectly matched the final percent identity would be 90%. In another example, a 90 nucleotide subject sequence is compared with a 100 nucleotide query sequence. This time the deletions are internal deletions so that

there are no nucleotides on the 5' or 3' of the subject sequence which are not matched/aligned with the query. In this case the percent identity calculated by FASTDB is not manually corrected. Once again, only nucleotides 5' and 3' of the subject sequence which are not matched/aligned with the query sequence are manually corrected for. No other manual
5 corrections are made for the purposes of the present invention.

TABLE 2. Closest matching sequence between the polypeptides of the present invention an sequences in GenSeq and GenBank databases

Sequence ID.	Antigen Accession No.	Match Gene Name	High Score	Smallest Sum Probability P (N)
<i>GenSeq</i>				
HGS010	W87771	UDP-N-acetylmuramate:L-alanine ligase (MurC polype...	2238	9.10E-308
HGS010	W89199	Partial sequence of the MurC polypeptide. New isol...	1067	1.20E-144
HGS010	W55120	Streptococcus pneumoniae SP0070 protein. Nucleic a...	451	2.30E-142
HGS010	W20606	H. pylori cytoplasmic protein, O1ep30520orf27. Hel...	147	1.10E-29
HGS010	W77686	Staphylococcus aureus protein of unknown function....	185	7.30E-19
HGS010	W20102	H. pylori cytoplasmic protein, 11253.aa. Helicobac...	122	1.10E-12
HGS010	W24585	H. pylori cytoplasmic protein, 11253.aa. Helicobac...	122	1.10E-12
HGS010	W29454	Streptococcus pneumoniae MurD protein. Streptococc...	99	7.50E-10
HGS010	W68551	S. pneumoniae MurD protein. Streptococcus pneumoni...	99	7.50E-10
HGS010	W55117	Streptococcus pneumoniae SP0067 protein. Nucleic a...	99	4.80E-09
HGS027	W29380	S. pneumoniae peptide releasing factor RF-1. DNA e...	593	1.00E-141
HGS027	W38592	S. pneumoniae peptide chain release factor 1. Nove...	593	1.00E-141
HGS029	W71494	Helicobacter polypeptide GHPO 805. Helicobacter po...	440	4.20E-55
HGS029	R14036	Ribosome releasing factor. Novel peptide promoting...	437	1.10E-54
HGS029	W69755	Ribosome recycling factor protein. Expression and ...	411	3.20E-51
HGS029	W69754	Ribosome recycling factor protein. Expression and ...	410	4.40E-51
HGS029	W78188	Human secreted protein encoded by gene 63 clone HP ...	109	2.10E-14
HGS038	W79340	Staphylococcus aureus nusA protein homologue. New ...	667	1.00E-89
HGS038	W98760	H. pylori GHPO 1087 protein. New isolated Helicoba...	260	1.70E-36

HGS039	W80656	<i>S. pneumoniae</i> transcription elongation factor. Str...	246	2.80E-33
HGS039	W27997	Amino acid sequence of transcription antitermini...	272	1.00E-32
HGS041	R58587	Nicotineamide adenine dinucleotide synthetase N-te...	181	8.10E-38
HGS042	W21022	<i>H. pylori</i> cytoplasmic protein, hp5e15440orf21. Hel...	295	1.90E-80
HGS042	R47583	NADH oxidase. DNA encoding NADH oxidase - used in ...	229	1.90E-39
HGS042	R60863	Hydrogen peroxide-generating NADH oxidase. A DNA f...	309	6.30E-35
HGS042	W28236	Amino acid sequence of a mercuric reductase. Novel...	91	1.60E-15
HGS042	W29772	<i>Malassezia fungus</i> MF-5 antigenic protein. Antigeni...	80	5.60E-14
HGS042	R43074	<i>Aspergillus niger</i> Sulphydryl oxidase (SOX). DNA en...	92	2.10E-12
HGS042	W53251	<i>Candida albicans</i> fungal antigen - allergen SEQ ID ...	76	9.80E-11
HGS042	W98700	<i>H. pylori</i> GHPO 698 protein. New isolated <i>Helicobac...</i>	65	3.60E-09
HGS043	W71558	<i>Helicobacter</i> polypeptide GHPO 1252. <i>Helicobacter p...</i>	437	1.60E-108
HGS043	W98793	<i>H. pylori</i> GHPO 1252 protein. New isolated <i>Helicoba...</i>	437	1.60E-108
HGS043	W20598	<i>H. pylori</i> protein. <i>Helicobacter pylori</i> nucleic aci...	434	2.20E-108
HGS043	W28298	<i>Staphylococcus aureus</i> protein of unknown function...	584	1.20E-75
HGS043	W20206	<i>H. pylori</i> derived protein. <i>Helicobacter pylori</i> nuc...	268	1.20E-37
HGS043	W88304	<i>E. coli</i> O111 antigen gene cluster ORF5 (manB) prot...	130	9.00E-32
HGS043	W88322	<i>E. coli</i> O157 antigen pathway ORF11 (manD) protein ...	128	7.60E-29
HGS043	R04578	Part of protein with urease activity. New nucleoti...	175	2.60E-17
HGS043	W88333	<i>Salmonella enterica</i> O antigen gene cluster manB pr...	69	2.90E-11
HGS043	W20803	<i>H. pylori</i> cytoplasmic protein, O9ap11406orf8. Heli...	84	5.60E-09
HGS044	W19930	N-acetylglucosamine 1-phosphate uridylyltransferase...	2281	8.60E-308
HGS044	W19929	N-acetylglucosamine 1-phosphate uridylyltransferase...	2275	5.70E-307
HGS044	W89182	<i>S. pneumoniae</i> GlimU polypeptide. New <i>Streptococcus</i> ...	1111	3.50E-148
HGS044	W89183	<i>S. pneumoniae</i> GlimU ORF polypeptide sequence. New S...	926	7.30E-123
HGS044	W98337	<i>H. pylori</i> GHPO 142 protein. New isolated <i>Helicobac...</i>	284	1.70E-101

HGS045	W18209	Staphylococcus aureus Coenzyme A disulphide reduct...	2236	7.40E-305
HGS045	W77578	Staphylococcus aureus protein of unknown function...	548	1.50E-71
HGS045	W06425	Water-forming NADH oxidase. DNA encoding water-for...	101	4.10E-34
HGS045	W94460	NADH:H ₂ O oxidase activity protein. Increasing the ...	75	5.00E-22
HGS045	W02649	Ascorbate-free-radical-reductase. New isolated tom...	86	4.40E-11
HGS045	W83401	Human thioredoxin reductase mature protein. Prepar...	82	2.10E-08
HGS045	R92050	KM31-7 precursor. Clover yellow vein virus nuclear...	82	3.50E-08
HGS045	W83404	Human KM-102-derived reductase like factor. Prepar...	82	3.50E-08
HGS046	W98618	H. pylori GHPO 231 protein. New isolated Helicobac...	108	3.20E-14
HGS046	W20305	H. pylori surface membrane protein 24409577.aa. He...	136	1.40E-11
HGS046	W20809	H. pylori surface or membrane protein, 09cp10502or...	134	2.10E-11
HGS049	W20733	H. pylori cell envelope protein, 06cp11722orf15. H...	156	2.70E-51
HGS049	W26775	Peptidoglycan biosynthetic enzyme MurE. Streptococ...	173	1.10E-31
HGS049	W20436	H. pylori protein. Helicobacter pylori nucleic aci...	98	6.00E-17
HGS049	W29454	Streptococcus pneumoniae MurD protein. Streptococc...	64	4.90E-08
HGS049	W68551	S. pneumoniae MurD protein. Streptococcus pneumoni...	64	4.90E-08
HGS050	W34453	MurF protein. Brevibacterium flavum murF gene - us...	513	4.10E-133
HGS050	W20826	H. pylori cytoplasmic protein 11ep12011orf9. Helic...	99	3.00E-18
HGS050	W71543	Helicobacter polypeptide GHPO 208. Helicobacter po...	92	2.30E-16
HGS050	W98302	H. pylori GHPO 208 protein. New isolated Helicobac...	92	2.30E-16
HGS053	W88645	Secreted protein encoded by gene 112 clone HUKFC71...	114	4.60E-10
HGS055	W62677	Streptococcus pneumoniae polypeptide. Streptococcu...	853	2.10E-113
HGS057	W38664	S. pneumoniae 30S ribosomal protein S9. Novel Stre...	387	2.10E-49
HGS059	W38499	S. pneumoniae ribosomal protein S14 (pS14). Novel...	292	4.30E-36

HGS060	P81003	Sequence encoding protein uniquely expressed by hu...	106	3.90E-08
HGS062	W38499	S. pneumoniae ribosomal protein S14 (rpS14). Novel...	167	9.90E-24
HGS064	W89791	Staphylococcus aureus protein SEQ ID #5239. Polynu...	1219	7.10E-166
HGS064	W38174	Response regulator amino acid sequence from S. pne...	399	3.30E-105
HGS064	W57633	S. pneumoniae response regulator protein. New isol...	399	3.30E-105
HGS064	W18219	Staphylococcus aureus response regulator protein. ...	266	5.30E-85
HGS064	W68415	Mycobacterium bovis regX3 protein. Mycobacterial n...	333	1.10E-71
HGS064	W38175	Response regulator amino acid sequence. DNA encodi...	353	9.70E-64
HGS064	W57634	S. pneumoniae response regulator protein. New isol...	353	5.70E-64
HGS064	W19274	Staphylococcus aureus novel response regulator pro...	303	1.90E-61
HGS064	W13272	Rhodococcus erythropolis SK92-B1 regulatory factor...	298	6.40E-58
HGS064	W80799	Rhodococcus nitrile hydratase gene fragment produc...	298	6.40E-58
HGS065	W80663	S. pneumoniae protein of unknown function. Strepto...	368	1.00E-46
HGS065	W38482	Streptococcus pneumoniae protein of unknown functi...	256	1.60E-29
HGS066	W77583	Staphylococcus aureus protein of unknown function...	332	4.80E-40
HGS067	W77630	Staphylococcus aureus protein of unknown function....	453	6.60E-59
HGS067	R34719	Bacillus subtilis srfA operon ORF8 prod. Multi-enz...	144	5.00E-12
HGS068	W74405	S. aureus gidB protein sequence. New Staphylococcu...	1229	4.70E-166
HGS068	W74406	S. aureus gidB protein sequence. New Staphylococcu...	1174	1.90E-158
HGS068	W89447	A gidB polypeptide sequence. New nucleic acid enco...	244	1.70E-56
HGS068	W77522	Glucose inhibited division protein B. New nucleic ...	269	1.60E-31
HGS070	W98338	H. pylori GHPO 250 protein. New isolated Helicobac...	274	1.10E-51
HGS070	W20646	H. pylori cytoplasmic protein, 02cp11822orf26. Hel...	291	1.00E-46

HGS070	W38565	S. pneumoniae uridylylate kinase. Novel Streptococcus...	246	5.00E-28
HGS070	W20147	H. pylori cytoplasmic protein, 14574201.aa. Helico...	75	3.00E-08
HGS071	W37743	S. pneumoniae DDL protein. Streptococcus pneumonia...	558	1.20E-113
HGS071	W46752	D-alanine-D-alanine ligase sequence of Mycobacteri...	182	1.00E-87
HGS071	R57151	Enterococcus faecalis vanB protein. New protein Va...	176	4.50E-72
HGS071	R24298	D-alanine-D-alanine ligase VanA from E. faecium. Po...	184	6.40E-70
HGS071	R24303	D-Ala-D-Ala ligase VanC involved in antibiotic res...	281	2.70E-66
HGS071	R24305	Translation of ORF 1 contg. E. faecium proteins Van...	184	1.00E-58
HGS071	R57150	Enterococcus faecalis vanB protein internal fragme...	155	2.00E-30
HGS071	W98614	H. pylori GHPO 205 protein. New isolated Helicobac...	92	7.00E-17
HGS072	W00285	Mutant farnesyl diphosphate synthase (4). Productio...	339	1.70E-86
HGS072	W00286	Native farnesyl diphosphate synthase. Production of...	335	2.30E-86
HGS072	W47444	Bacillus stearothermophilus farnesyl diphosphate s...	333	4.30E-86
HGS072	W62532	Farnesyl diphosphate synthase of B. stearothermophi...	333	4.30E-86
HGS072	W00283	Mutant farnesyl diphosphate synthase (2). Productio...	332	5.90E-86
HGS072	R35047	FPS. New thermally stable farnesyl pyrophosphate s...	333	1.10E-85
HGS072	W00284	Mutant farnesyl diphosphate synthase (3). Productio...	333	1.50E-85
HGS072	W00282	Mutant farnesyl diphosphate synthase (1). Productio...	328	7.30E-85
HGS072	W62535	Mutant farnesyl diphosphate synthase of B. stearoth...	331	8.90E-84
HGS072	W62537	Mutant farnesyl diphosphate synthase of B. stearoth...	331	8.90E-84
HGS073	R92060	Heptaprenyl diphosphate synthetase ORFIII product...	506	1.10E-81
HGS073	W47422	Bacillus stearothermophilus prenyl diphosphate syn...	506	1.10E-81
HGS073	W47420	Micrococcus luteus prenyl diphosphate synthetase s...	493	4.30E-70
HGS073	W53922	Decaprenyl diphosphate synthase #3. Production of ...	292	1.20E-36
HGS073	W53920	Decaprenyl diphosphate synthase #1. Production of ...	292	2.80E-36
HGS073	W53921	Decaprenyl diphosphate synthase #2. Production of ...	292	6.80E-36
HGS073	W12389	Geranylgeranyl diphosphate synthase F77S mutant. N...	172	4.30E-34
HGS073	W12386	Geranylgeranyl diphosphate synthase. New mutant ge...	177	6.90E-34

HGS073	W12388	Geranylgeranyl diphosphate synthase F118L mutant. ...	173	6.90E-34
HGS073	R79969	Geranylgeranyl diphosphate synthase. DNA encoding ...	174	2.70E-33
HGS074	W60977	Streptococcus pneumoniae encoded polypeptide. New ...	253	2.10E-58
HGS074	W80710	S. pneumoniae protein of unknown function. Strepto...	248	9.80E-58
HGS075	W83372	Streptococcus pneumoniae histidine kinase. New Str...	175	1.10E-33
HGS075	W68414	Mycobacterium bovis senX3 protein. Mycobacterial n...	159	3.50E-27
HGS075	R24296	Regulatory protein VanS involved in glycopeptide r...	143	2.60E-25
HGS075	W68522	N. crassa os1p protein. New assay for histidine ki...	185	4.30E-24
HGS075	W83377	Streptococcus pneumoniae histidine kinase. New Str...	135	8.40E-23
HGS075	W89427	S. pneumoniae histidine kinase polypeptide. New hi...	135	8.40E-23
HGS075	W89432	Streptococcus pneumoniae histidine kinase. New Str...	135	8.40E-23
HGS075	W81600	Candida albicans CaNIK1 protein involved in phenot...	168	5.30E-22
HGS075	R24306	Translation of ORF 2 contg. E. faecium protein VanS...	142	8.90E-22
HGS075	W68523	Partial C. albicans cos1p protein. New assay for h...	151	5.90E-21
Pbp1	W98771	H. pylori GHPO 1134 protein. New isolated Helicoba...	87	3.80E-12
Pbp1	R27253	Penicillin binding protein PBP2A-epi. Polynucleoti...	78	1.90E-08
deaD	W60667	E. coli cold shock protein CsdA. Modulating protein...	395	2.40E-119
deaD	W24291	Lmelf4A. Compositions comprising LbelF4A and Lmelf...	321	9.80E-81
deaD	R77503	Leishmania sp. antigen LbelF4A. DNA encoding prote...	317	2.90E-80
deaD	W24290	LbelF4A. Compositions comprising LbelF4A and Lmelf...	317	2.90E-80
deaD	W70213	Leishmania antigen LbelF4A protein. New immunogeni...	317	2.90E-80
deaD	W92743	L. braziliensis EIF4A protein. New Leishmania braz...	317	2.90E-80
deaD	W81502	Dead Box X (DBX) gene short transcript amino acid ...	276	7.40E-80
deaD	W81501	Dead Box X (DBX) gene long transcript amino acid s...	276	7.40E-80
deaD	W11218	Leishmania braziliensis LbelF4A antigen. Polypepti...	313	1.20E-79
deaD	W81503	Dead Box Y (DBY) gene product. Novel genes in the ...	279	5.20E-74

Genbank

HGS010	gij2642659	(AF034076) UDP-N-acetylmuramoyl-L-alanin...	2255	0.00E+00
HGS010	gnlPID e1185852	UDP-N-acetyl muramate-alanine ligase [Ba...	1438	7.30E-196
HGS010	gij2688761	(AE001180) UDP-N-acetylmuramate--alanine...	169	2.30E-56
HGS010	gij2983764	(AE000736) UDP-N-acetylmuramate-alanine ...	183	8.80E-54
HGS010	gnlPID d1035273	(AB015023) MurC [Corynebacterium glutami...	108	1.30E-52
HGS010	gij42056	(UDP-N-acetylmuramate: L-alanine ligase)...	191	2.70E-51
HGS010	gij2177094	UDP-MurNAc:L-alanine ligase [Escherichia...	191	2.70E-51
HGS010	gij3322616	(AE001213) UDP-N-acetylmuramate--alanine...	165	1.10E-45
HGS010	gij1574695	UDP-N-acetylmuramate--alanine ligase (mu...	175	4.80E-44
HGS010	gnlPID d1025270	MurC [Porphyromonas gingivalis] >sp Q518...	182	6.40E-44
HGS027	gnlPID e1184607	peptide chain release factor 1 [Bacillus...	888	8.80E-160
HGS027	gnlPID d1009421	Peptide Termination Factor [Mycoplasma c...	715	1.10E-126
HGS027	gnlPID d1019559	peptide chain release factor [Synechocys...	539	4.00E-121
HGS027	gij2688096	(AE001130) peptide chain release factor ...	628	1.40E-115
HGS027	gnlPID e1342822	(AJ235272) PEPTIDE CHAIN RELEASE FACTOR ...	569	1.50E-115
HGS027	gnlPID d1015453	Peptide chain release factor 1 (RF-1) [E...	467	4.30E-113
HGS027	gij968930	peptide chain release factor 1 [Escheric...	463	1.50E-112
HGS027	gij3328413	(AE001277) Peptide Chain Releasing Facto...	430	1.50E-112
HGS027	gij3322309	(AE001190) peptide chain release factor ...	604	1.70E-112
HGS027	gij147567	peptide chain release factor 1 [Escheric...	467	3.60E-112
HGS029	gij2645713	(AF033018) ribosome recycling factor [St...	895	2.60E-115
HGS029	gnlPID e1185243	ribosome recycling factor [Bacillus subt...	633	2.90E-80
HGS029	gnlPID d1019290	ribosome releasing factor [Synechocystis...	486	1.40E-60
HGS029	gnlPID e248763	fr [Mycobacterium tuberculosis]	445	4.10E-55
HGS029	gij2314423	(AE000631) ribosome releasing factor (fr...	440	1.90E-54
HGS029	gij1573820	ribosome releasing factor (rrf) [Haemoph...	438	3.60E-54
HGS029	gij147771	ribosome releasing factor (gtg start cod...	437	4.90E-54
HGS029	gij3322898	(AE001235) ribosome recycling factor [Tr...	433	1.70E-53

HGS029	gnl PID e327819	ribosome recycling factor [Mycobacterium...	431	3.10E-53
HGS029	gil4155787	(AE001545) RIBOSOME RECYCLING FACTOR (RI...	430	4.20E-53
HGS038	gnl PID e1185251	nusA [Bacillus subtilis] >pir B69668 B6...	1218	1.50E-160
HGS038	gil49316	ORF2 gene product [Bacillus subtilis] >...	1210	1.80E-159
HGS038	gnl PID e1342846	(AJ235272) N UTILIZATION SUBSTANCE PROT...	602	6.90E-97
HGS038	gil642364	NusA protein [Thermus aquaticus thermop...	502	6.30E-92
HGS038	gnl PID e1299837	nusA [Mycobacterium tuberculosis] >sp O...	333	2.40E-89
HGS038	gil3323210	(AE001259) N utilization substance prot...	288	3.20E-87
HGS038	gil606109	L factor [Escherichia coli] >gil1789560...	412	5.60E-86
HGS038	gil515637	transcription factor [Salmonella typhim...	409	1.90E-85
HGS038	pir D64114 D64114	transcription termination-antiterminati...	418	2.00E-83
HGS038	gnl PID e1172585	NusA protein (nusA) [Escherichia coli]	608	3.40E-78
HGS039	gil2078377	NusG [Staphylococcus aureus] >sp O08386 ...	924	4.00E-121
HGS039	gil426473	nusG gene product [Staphylococcus carnos...	894	4.80E-117
HGS039	gnl PID d1003063	transcription antitermination factor Nus...	648	1.30E-83
HGS039	gnl PID e306572	nusG [Mycobacterium tuberculosis]	289	1.90E-60
HGS039	spiP96930 P96930	TRANSCRIPTION ANTITERMINATION PROTEIN NUSG.	289	1.90E-60
HGS039	gnl PID d1007561	nusG [Streptomyces coelicolor] >pir S547...	290	1.20E-53
HGS039	gil457386	transcription factor [Thermus aquaticus ...	148	1.50E-53
HGS039	gnl PID d1004802	NusG [Streptomyces coelicolor] >pir S410...	283	1.10E-52
HGS039	gnl PID d1004801	NusG [Streptomyces griseus] >pir S41061 ...	282	1.50E-52
HGS039	gnl PID e349728	NusG [Streptomyces griseus] >pir S32234 ...	282	1.50E-52
HGS041	gnl PID d1016252	NH(3)-dependent NAD(+) synthetase (EC 6...	620	2.60E-105
HGS041	gil146974	NH3-dependent NAD synthetase [Escherich...	620	3.30E-99
HGS041	gil143279	outB [Bacillus subtilis] >gnl PID d1009...	410	1.50E-87
HGS041	gnl PID d1030194	(AP000001) 257aa long hypothetical NH(3...	156	1.20E-21
HGS041	pir S77778 S77778	probable NH(3)-dependent NAD(+) synthet...	153	1.80E-20
HGS041	gil2649596	(AE001035) NH(3)-dependent NAD+ synthet...	167	3.60E-19

HGS041	gi 2622628	(AE000911) NH(3)-dependent NAD+ synthet...	167	1.90E-18
HGS041	gi 3844972	NH(3)-dependent NAD+ synthetase, putati...	142	3.20E-18
HGS041	gi 1673951	(AE000027) Mycoplasma pneumoniae, proba...	140	6.30E-16
HGS041	gi 1591995	NH(3)-dependent NAD+ synthetase (nadE) ...	162	1.30E-14
HGS042	gnl PID e1320012	(AJ223781) thioredoxin reductase [Staph...	1592	1.50E-214
HGS042	gnl PID e1313024	hypothetical protein [Bacillus subtilis...	1162	1.80E-155
HGS042	gi 2246749	(AF009622) thioredoxin reductase [Liste...	1060	1.80E-141
HGS042	gi 1353197	thioredoxin reductase [Eubacterium acid...	404	3.80E-98
HGS042	pir S38988 D35156	thioredoxin reductase (NADPH) (EC 1.6.4...	404	3.80E-98
HGS042	gi 3323124	(AE001252) thioredoxin reductase (trxB)...	353	1.80E-96
HGS042	gi 1171125	thioredoxin reductase [Clostridium lito...	397	3.80E-95
HGS042	gi 2262173	(AC002329) NADPH thioredoxin reductase ...	193	1.80E-84
HGS042	pir S44027 S44027	thioredoxin reductase (NADPH) (EC 1.6.4...	188	7.10E-84
HGS042	gnl PID d1008681	Thioredoxin Reductase (NADPH) [Neurospo...	145	5.20E-82
HGS043	gnl PID e283110	femD [Staphylococcus aureus] >gnl PID e1...	2299	0.00E+00
HGS043	gnl PID e284993	phosphoglucosamine mutase [Staphylococcu...	2295	0.00E+00
HGS043	gnl PID e1182110	similar to phosphoglucosamine mutase (glycolysi...	1419	8.30E-211
HGS043	gnl PID d1034036	(AB006424) ybbT [Bacillus subtilis] >sp ...	1419	5.50E-210
HGS043	gnl PID e1318460	(AL031317) putative phospho-sugar mutase...	744	1.90E-145
HGS043	gi 467124	ured; B229_C3_234 [Mycobacterium leprae]...	643	9.70E-133
HGS043	gnl PID e316048	mrsA [Mycobacterium tuberculosis] >sp O0...	655	1.40E-132
HGS043	gi 1574798	mrsA protein (mrsA) [Haemophilus influen...	349	4.50E-129
HGS043	gnl PID d1018426	hypothetical protein [Synecocystis sp.]...	422	7.00E-119
HGS043	gi 3329284	(AE001354) Phosphoglucosamine mutase [Chlamydia...	598	5.70E-117
HGS044	gnl PID d1005827	temperature sensitive cell division [Bac...	1338	1.70E-178
HGS044	gi 40217	tms gene product (AA 1-456) [Bacillus su...	1330	2.10E-177
HGS044	gnl PID e304562	gimU [Mycobacterium tuberculosis] >sp P9...	765	1.50E-122
HGS044	gi 2983227	(AE000698) UDP-N-acetylglucosamine pyrop...	349	5.10E-118

HGS044	gil975206	uridylyltransferase [Neisseria gonorrhoeae...	373	2.90E-117
HGS044	gnl PID d1011507	UDP-N-acetylglucosamine pyrophosphorylas...	486	2.30E-114
HGS044	gil43267	Eco urf 1 protein [Escherichia coli]	413	3.50E-111
HGS044	gil1790168	(AE000450) N-acetyl glucosamine-1-phosph...	413	6.40E-111
HGS044	gil1573640	UDP-N-acetylglucosamine pyrophosphorylas...	381	1.80E-110
HGS044	gil2313807	(AE000581) UDP-N-acetylglucosamine pyrop...	264	8.00E-101
HGS045	gil2792490	(AF041467) coenzyme A disulfide reductas...	2243	3.70E-305
HGS045	gil2688656	(AE001172) NADH oxidase, water-forming (...	194	7.30E-91
HGS045	gil1591361	NADH oxidase (nox) [Methanococcus jannas...	153	1.60E-51
HGS045	gnl PID d1031560	(AP000006) 440aa long hypothetical NADH ...	113	8.10E-44
HGS045	gil2650233	(AE001077) NADH oxidase (noxA-3) [Archae...	139	3.20E-40
HGS045	gil2650234	(AE001077) NADH oxidase (noxA-2) [Archae...	100	2.30E-37
HGS045	gil642030	NADH oxidase [Serpulina hydrosentieriae] ...	109	4.10E-36
HGS045	gnl PID d1030604	(AP000002) 445aa long hypothetical NADH ...	115	1.90E-35
HGS045	gil2622461	(AE000898) NADH oxidase [Methanobacteriu...	130	4.70E-35
HGS045	gil49023	NADH peroxidase [Enterococcus faecalis] ...	96	1.60E-33
HGS046	gnl PID e1183495	similar to hypothetical proteins from B....	192	8.00E-39
HGS046	gil2688416	(AE001152) conserved hypothetical integr...	263	8.30E-35
HGS046	gil2649097	(AE001001) conserved hypothetical protei...	132	3.00E-30
HGS046	gnl PID d1030609	(AP000002) 449aa long hypothetical damag...	253	3.50E-29
HGS046	gnl PID d1030607	(AP000002) 472aa long hypothetical prote...	226	1.40E-26
HGS046	gil1591425	conserved hypothetical protein [Methanoc...	159	4.60E-26
HGS046	gil2314344	(AE000624) conserved hypothetical integr...	230	1.90E-25
HGS046	gil4155699	(AE001538) putative [Helicobacter pylori]...	224	6.80E-25
HGS046	gil2621368	(AE000816) conserved protein [Methanobac...	122	1.80E-22
HGS046	gnl PID e340160	DinF protein [Streptococcus pneumoniae] ...	219	2.70E-22
HGS050	dbjl AB001488_41	(AB001488) PROBABLE UDP-N-ACETYLMURAMOYL...	513	5.60E-133
HGS050	gil4009466	(AF068901) D-Ala-D-Ala adding enzyme [St...	341	1.30E-90

HGS050	gil1574689	UDP-MurNAC-pentapeptide synthetase (murF...	386	5.80E-87
HGS050	gil2177096	UDP-MurNAC-Tripeptide-D-Ala-D-Ala-Adding...	265	1.40E-65
HGS050	gil1743865	UDP-MurNAC-Tripeptide-D-Ala-D-Ala-Adding...	265	1.40E-65
HGS050	gil42048	UDP-MurNAC-pentapeptide presynthetase (A...	263	5.20E-64
HGS050	gil2983375	(AE000709) UDP-MURNAC-pentapeptide sythe...	244	3.50E-63
HGS050	gil3322664	(AE001217) UDP-N-acetylmuramoylalanyl-D...	212	8.30E-55
HGS050	gil575416	UDP-N-acetylmuramoylalanyl-D-glutamyl-2...	309	1.70E-54
HGS050	gnlPID d1018904	UDP-N-acetylmuramoylalanyl-D-glutamyl-2...	309	1.70E-54
HGS052	gil1044978	ribosomal protein S8 [Bacillus subtilis]...	564	8.20E-73
HGS052	gnlPID d1011637	ribosomal protein S8 [Bacillus subtilis]...	551	5.00E-71
HGS052	gil44429	S8 protein [Micrococcus luteus] >pilS29...	339	9.10E-50
HGS052	gnlPID e1358535	ribosomal protein S8 [Thermotoga maritima]...	205	9.60E-50
HGS052	gnlPID e293129	rpsH [Mycobacterium tuberculosis] >sp P9...	386	2.20E-48
HGS052	gnlPID e337975	ribosomal protein S8 [Mycobacterium leprae]	385	3.00E-48
HGS052	gil1276767	30S ribosomal protein S8 [Porphyra purpu...	231	1.00E-47
HGS052	dbj AB000111_15	(AB000111) 30S ribosomal protein S8 [Syn...	225	3.50E-47
HGS052	gil498771	ribosomal S8 protein [Thermus aquaticus]...	197	2.20E-46
HGS052	gil48108	ribosomal protein S8 [Thermus aquaticus]...	190	5.70E-46
HGS053	gnlPID e269878	ribosomal protein S15 [Bacillus subtilis]...	365	5.10E-46
HGS053	gnlPID e1173915	(AL008967) rpsO [Mycobacterium tuberculo...	290	1.40E-35
HGS053	gnlPID e335030	30s ribosomal protein s15 [Mycobacterium...	286	5.00E-35
HGS053	gnlPID e1315092	(AL031231) 30S ribosomal protein S15 [St...	270	7.90E-33
HGS053	gnlPID e118966	ribosomal protein S15 [Thermus thermophi...	266	3.00E-32
HGS053	gnlPID d1017615	30S ribosomal protein S15 [Synechocystis...	259	2.80E-31
HGS053	gil147748	ribosomal protein S15 [Escherichia coli]...	257	5.40E-31
HGS053	gnlPID e1342799	(AJ235272) 30S RIBOSOMAL PROTEIN S15 (rp...	256	7.30E-31
HGS053	gnlPID e321499	rpsO [Yersinia enterocolitica]	246	1.60E-29
HGS053	gil2982947	(AE000679) ribosomal protein S15 [Aquife...	245	2.50E-29

HGS055	gil1165309	S3 [Bacillus subtilis]	872	2.30E-115
HGS055	gnlPIDjd1009470	Ribosomal Protein S3 [Bacillus subtilis]...	870	4.30E-115
HGS055	gil580921	ribosomal protein S3 [Bacillus stearothermophilus]...	860	1.00E-113
HGS055	gil456688	ribosomal protein S3 [Acholeplasma palmae]...	486	1.70E-87
HGS055	gil3047158	ribosomal protein S3 [Phytolasma sp. ST]...	492	1.90E-85
HGS055	gil149869	rps3 [Mycoplasma-like organism] >pir[B41]...	494	8.90E-85
HGS055	gil456692	ribosomal protein S3 [Anaerobaculum abacti]...	468	8.00E-84
HGS055	gil1573793	ribosomal protein S3 (rps3) [Haemophilus]...	568	9.20E-84
HGS055	gnlPIDjd1011609	ribosomal protein S3 [Actinobacillus actinomycetum]...	562	6.10E-83
HGS055	gil141818	5' end of coding region undetermined [Acetivibrio]...	465	1.60E-81
HGS056	gil1044981	ribosomal protein S5 [Bacillus subtilis]...	626	5.10E-81
HGS056	gil143575	spc ORF1; S5 [Bacillus subtilis] >pir[S1]...	565	6.90E-80
HGS056	gil143417	ribosomal protein S5 [Bacillus stearothermophilus]...	613	3.00E-79
HGS056	gnlPIDje1254448	S5 ribosomal protein [Streptomyces coelicolor]...	487	4.10E-62
HGS056	gil44432	S5 protein [Micrococcus luteus] >pir[S29]...	422	4.20E-60
HGS056	gil606237	30S ribosomal subunit protein S5 [Escherichia coli]...	470	1.00E-59
HGS056	gil1573805	ribosomal protein S5 (rps5) [Haemophilus]...	467	2.70E-59
HGS056	gnlPIDje1234851	(AJ232327) ribosomal protein S5 [Salmonella]...	460	2.40E-58
HGS056	gil44226	ribosomal protein S5 (AA 1-250) [Mycoplasma]...	455	7.70E-58
HGS056	gil3322469	(AE001202) ribosomal protein S5 (rpsE) [Mycoplasma]...	451	3.90E-57
HGS057	gnlPIDjd1011647	ribosomal protein S9 [Bacillus subtilis]...	505	4.20E-65
HGS057	pirS08564R3BS9	ribosomal protein S9 - Bacillus stearothermophilus...	482	6.30E-62
HGS057	gil1673892	(AE000022) Mycoplasma pneumoniae, ribosomal protein S9...	293	1.10E-42
HGS057	gil1276757	30S ribosomal protein S9 [Porphyra purpurea]...	234	5.20E-42
HGS057	gnlPIDjd1018054	30S ribosomal protein S9 [Synecocystis]...	241	7.10E-42
HGS057	gnlPIDje1316459	(AL031317) 30S ribosomal protein S9 [Stramonium]...	220	5.20E-41
HGS057	gil606169	30S ribosomal subunit protein S9 [Escherichia coli]...	325	2.90E-40
HGS057	gil3323359	(AE001270) ribosomal protein S9 (rpsI) [Mycoplasma]...	318	2.70E-39
HGS057	gil3845009	ribosomal protein S9 (rps9) [Mycoplasma]...	273	1.40E-38

HGS057	gil2688239	(AE001140) ribosomal protein, S9 (rpsI) [...]	308	6.20E-38
HGS058	gnlPID d1011664	ribosomal protein S10 [Bacillus subtilis...	479	3.40E-61
HGS058	gil1165302	S10 [Bacillus subtilis]	472	3.10E-60
HGS058	gil467321	S10 ribosomal protein [Streptococcus mut...	422	2.40E-53
HGS058	gnlPID e260119	ribosomal protein S10 [Planobispora rose...	393	2.30E-49
HGS058	gnlPID e1192296	rpsX [Mycobacterium bovis BCG] >gnlPID ...	385	3.00E-48
HGS058	gil681340	ribosomal protein S10 [Mycobacterium lep...	384	4.10E-48
HGS058	gil437922	ribosomal protein S10 [Thermotoga mariti...	369	4.60E-46
HGS058	gil44208	ribosomal protein S10 (AA 1-102) [Mycopl...	367	8.80E-46
HGS058	gil3328867	(AE001317) S10 Ribosomal Protein [Chlamy...	333	2.40E-44
HGS058	gil1573786	ribosomal protein S10 (rps10) [Haemophil...	343	1.50E-42
HGS059	gnlPID e1182877	similar to ribosomal protein S14 [Bacill...	344	1.30E-42
HGS059	gil3329252	(AE001351) S14 Ribosomal Protein [Chlamy...	187	1.60E-29
HGS059	gil606241	30S ribosomal subunit protein, S14 [Esche...	181	5.50E-29
HGS059	gil1016092	ribosomal protein S14 [Cyanophora paradoxa]	167	5.60E-29
HGS059	gil1573801	ribosomal protein S14 (rps14) [Haemophil...	186	1.00E-28
HGS059	gil414859	30s ribosomal protein S14 [Astasia longa...	174	2.70E-28
HGS059	gil42982	S14 (rpsN) (aa 1-99) [Escherichia coli]	165	8.40E-27
HGS059	gnlPID d1007159	ribosomal protein S14 [Acyrtosiphon kon...	164	2.10E-26
HGS059	gil11670	rps14 [Marchantia polymorpha] >pir A0273...	155	2.10E-26
HGS059	gnlPID e1299756	(AL021899) rpsN2 [Mycobacterium tubercul...	167	5.40E-26
HGS060	gnlPID d1009468	Ribosomal Protein S19 [Bacillus subtilis...	409	4.90E-52
HGS060	gnlPID e316791	rpsS [Mycobacterium bovis BCG] >gnlPID ...	401	6.20E-51
HGS060	gil40106	ribosomal protein S19 [Bacillus stearoth...	400	8.60E-51
HGS060	bbs137759	S19=30S ribosomal protein [Mycobacterium...	395	4.20E-50
HGS060	gnlPID e337967	ribosomal protein S19 [Mycobacterium lep...	395	4.20E-50
HGS060	gil1016142	ribosomal protein S19 [Cyanophora parado...	344	5.20E-43
HGS060	dbj AB000111_6	(AB000111) 30S ribosomal protein S19 [Sy...	342	9.90E-43

HGS060	gil606250	30S ribosomal subunit protein S19 [Esche...	332	2.40E-41
HGS060	gil11715	rps19 [Marchantia polymorpha] >pir A0274...	330	4.50E-41
HGS060	gnl PID d1021585	(AB001684) 30S ribosomal protein S19 [Ch...	329	6.20E-41
HGS062	gil580930	S14 protein (AA 1-61) [Bacillus subtili...	285	1.60E-35
HGS062	pir S48688 S48688	ribosomal protein S14 - Bacillus stearo...	279	1.10E-34
HGS062	gil2766516	(AF036708) ribosomal protein S14 [Mycop...	240	3.80E-29
HGS062	gil4155818	(AE001547) 30S RIBOSOMAL PROTEIN S14 [H...	232	5.20E-28
HGS062	gnl PID e1358534	ribosomal protein S14 [Thermotoga marit...	230	1.00E-27
HGS062	gil44222	ribosomal protein S14 (AA 1-61) [Mycopl...	228	1.90E-27
HGS062	gil2314484	(AE000633) ribosomal protein S14 (rpS14...	228	1.90E-27
HGS062	bbs168339	ribosomal protein S14 [Thermus thermoph...	219	3.60E-26
HGS062	gnl PID e293291	rpsN [Mycobacterium tuberculosis] >sp P...	218	5.00E-26
HGS062	gil48107	ribosomal protein S14 [Thermus aquaticus]	217	7.00E-26
HGS064	gnl PID d1005715	unknown [Bacillus subtilis] >gnl PID d10...	950	8.60E-128
HGS064	gil4104602	(AF036966) putative response regulator [...	469	4.80E-121
HGS064	gnl PID e1299427	(AJ001103) arca [Lactococcus lactis] >sp...	794	4.40E-106
HGS064	gil1575577	DNA-binding response regulator [Thermoto...	278	1.50E-82
HGS064	gnl PID d1011205	regulatory components of sensory transdu...	239	1.30E-72
HGS064	gnl PID e321544	RegX3 [Mycobacterium bovis BCG] >sp O071...	333	5.00E-71
HGS064	gnl PID e321547	RegX3 [Mycobacterium tuberculosis] >gnl ...	333	5.00E-71
HGS064	gnl PID d1002953	SphR [Synecococcus sp.] >pir S32931 S32...	198	1.10E-70
HGS064	gnl PID e314479	mtaA [Mycobacterium tuberculosis] >sp Q5...	329	8.10E-69
HGS064	gnl PID e1181525	(AJ002571) YkoG [Bacillus subtilis] >gnl...	193	4.80E-68
HGS063	gnl PID d1037676	(AB016431) Hypothetical protein [Staphyl...	870	9.00E-116
HGS063	gnl PID d1004537	Mannosephosphate isomerase [Streptococcu...	302	2.80E-102
HGS063	gnl PID d1020490	B. subtilis mannose-6-phosphate isomeras...	662	4.80E-96
HGS063	gnl PID e1183222	similar to mannose-6-phosphate isomerase...	724	7.00E-96
HGS063	gil476092	unknown [Bacillus subtilis] >gnl PID d10...	659	1.10E-94

HGS063	gi 3043889	(AF015751) Cyp4 [Lactococcus lactis] >sp...	168	5.60E-16
HGS065	gnl PID d1005813	unknown [Bacillus subtilis] >gnl PID e11...	545	1.10E-70
HGS065	gnl PID d1018846	hypothetical protein [Synechocystis sp.]...	430	1.60E-69
HGS065	gi 606086	ORF_1286 [Escherichia coli] >gi 1789535 ...	422	3.60E-66
HGS065	gi 1574503	conserved hypothetical protein [Haemophi...	426	2.00E-63
HGS065	gnl PID d1002952	ORF3 [Micromonospora olivasterospora] >p...	455	2.50E-58
HGS065	gi 1045730	conserved hypothetical protein [Mycoplas...	163	1.00E-56
HGS065	gnl PID e1343020	(AJ235273) unknown [Rickettsia prowazeki...	409	1.80E-56
HGS065	gi 1673738	(AE000010) Mycoplasma pneumoniae, hypothe...	152	3.50E-54
HGS065	gi 2983597	(AE000724) hypothetical protein [Aquifex...	313	2.60E-51
HGS065	gi 2688342	(AE001148) conserved hypothetical protel...	377	1.60E-47
HGS066	gnl PID e1185047	alternate gene name: ykrC; similar to h...	152	1.70E-14
HGS066	gi 143376	ORF5 [Bacillus subtilis]	147	7.60E-14
HGS066	pir A42771 A42771	reticulocyte-binding protein 1 - Plasm...	55	5.30E-09
HGS066	gi 160626	reticulocyte binding protein 1 [Plasmod...	55	5.70E-09
HGS067	gnl PID d1011933	yydK [Bacillus subtilis] >sp Q45591 Q455...	278	1.20E-62
HGS067	gi 2668604	(AF015453) GNTR transcriptional regulato...	143	6.10E-21
HGS067	gnl PID e1186191	similar to transcriptional regulator (Gn...	108	3.30E-16
HGS067	gi 290533	similar to E. coli ORF adjacent to suc o...	104	4.30E-16
HGS067	gi 1000453	TreR [Bacillus subtilis] >gi 2626829 Tre...	167	7.10E-15
HGS067	gnl PID d1020488	K. aerogenes, histidine utilization repr...	118	9.40E-15
HGS067	gi 41519	P30 protein (AA 1-240) [Escherichia coli...	108	9.90E-15
HGS067	gi 1763080	PhnR [Salmonella typhimurium] >sp P96061...	99	1.90E-14
HGS067	gnl PID e1184335	similar to transcriptional regulator (Gn...	87	5.00E-12
HGS067	gi 396486	ORF8 [Bacillus subtilis] >gnl PID d10096...	144	2.30E-11
HGS068	gi 40027	homologous to E.coli gidB [Bacillus subt...	444	1.50E-102
HGS068	gi 950065	methyltransferases [Mycoplasma capricolu...	164	2.60E-47

HGS068	gnlPIDjd1011190	glucose inhibited division protein B [Sy...	157	3.20E-38
HGS068	gil290589	glucose inhibited division protein [Esch...	136	1.60E-33
HGS068	gil1573466	glucose-inhibited division protein (gidB...	117	1.10E-31
HGS068	gnlPIDje290777	orf256; translated orf similarity to SWI...	136	1.80E-26
HGS068	gil581464	homologous to E.coli gidB [Pseudomonas p...	139	6.40E-23
HGS068	gil2983927	(AE000746) glucose inhibited division pr...	117	3.70E-21
HGS068	gil2898105	(AF031590) GidB-like [Streptomyces coeli...	130	2.90E-20
HGS068	gil2314206	(AE000613) glucose-inhibited division pr...	121	2.40E-17
HGS069	gnlPIDjd1005835	unknown [Bacillus subtilis] >gnlPIDje11...	328	6.00E-100
HGS069	gil2983025	(AE000684) hypothetical protein [Aquifex...	281	9.30E-31
HGS069	gil2708269	unknown [Brucella abortus] >sp[O54384]O5...	259	1.80E-27
HGS069	gnlPIDje242767	product is homologous to Streptococcus c...	253	6.60E-27
HGS069	gil416198	homologous to a Streptomyces cacaoi beta...	251	2.80E-26
HGS069	gil882675	CG Site No. 33299 [Escherichia coli] >gi...	251	2.90E-26
HGS069	gnlPIDjd1017776	regulatory protein for beta-lactamase [S...	232	1.70E-23
HGS069	gil1573434	mazG protein (mazG) [Haemophilus influen...	223	3.20E-22
HGS069	gnlPIDjd1001238	regulatory protein for beta-lactamase [S...	133	5.50E-18
HGS069	gil3323087	(AE001249) mazG protein (mazG) [Treponem...	124	2.40E-13
HGS070	gnlPIDje1185242	uridylyate kinase [Bacillus subtilis] >pi...	920	2.20E-122
HGS070	gnlPIDjd1019291	uridine monophosphate kinase [Synechocys...	530	3.80E-96
HGS070	gnlPIDje1296663	(AL023797) uridylyate kinase [Streptomyce...	678	4.50E-89
HGS070	gnlPIDje248883	pyrH [Mycobacterium tuberculosis]	416	1.30E-88
HGS070	gnlPIDje327783	uridylyate kinase [Mycobacterium leprae] ...	403	1.60E-85
HGS070	gil473234	uridine 5'-monophosphate (UMP) kinase [E...	384	4.00E-72
HGS070	gil1552748	uridine 5'-monophosphate (UMP) kinase [E...	375	6.80E-71
HGS070	gil1574616	uridylyate kinase (pyrH) [Haemophilus inf...	409	6.90E-71
HGS070	gnlPIDjd1033306	(AB010087) UMP kinase [Pseudomonas aerug...	355	7.70E-66
HGS070	gnlPIDje1342466	(AJ235270) URIDYLATE KINASE (pyrH) [Rick...	461	3.60E-59

HGS071	dbj AB001488_40	(AB001488) PROBABLE D-ALANINE--D-ALANINE...	812	2.70E-123
HGS071	gil1244574	D-alanine:D-alanine ligase [Enterococcus...	732	2.60E-114
HGS071	gil460080	D-alanine:D-alanine ligase-related prote...	742	2.70E-114
HGS071	gnl PID e304921	unnamed protein product [unidentified] >...	742	2.70E-114
HGS071	gil4009465	(AF068901) D-Ala-D-Ala ligase [Streptoco...	554	9.40E-112
HGS071	gnl PID d1018410	D-alanine:D-alanine ligase-related prote...	219	2.00E-107
HGS071	gil153943	D-alanine:D-alanine ligase (EC 6.3.2.4) ...	256	1.90E-103
HGS071	gil145722	D-alanine:D-alanine ligase A [Escherichi...	240	4.60E-100
HGS071	gil1244572	D-alanine:D-alanine ligase [Enterococcus...	625	2.00E-98
HGS071	gnl PID e1359179	(AL034447) D-alanine-D-alanine ligase [S...	239	2.70E-92
HGS072	gnl PID d1003054	farnesyl diphosphate synthase [Bacillus ...	333	2.00E-85
HGS072	gnl PID e1185696	similar to geranyltransferase [Baci...	335	2.10E-82
HGS072	gnl PID d1026193	(AB003187) farnesyl diphosphate synthase...	340	3.70E-82
HGS072	gil1016225	CrtE (Cyanophora paradoxa) >sp P48368 CR...	302	2.40E-69
HGS072	gnl PID d1017423	geranylgeranyl pyrophosphate synthase [S...	293	8.20E-69
HGS072	gil3885426	(AF020041) geranylgeranyl pyrophosphate ...	302	1.70E-65
HGS072	sp O81099 O81099	GERANYLGERANYL PYROPHOSPHATE SYNTHASE.	302	3.10E-65
HGS072	gil1574277	geranyltransferase (ispA) [Haemophi...	344	5.10E-65
HGS072	gil1773105	geranyltransferase [Escherichia coli] >g...	228	1.80E-64
HGS072	gil1063276	geranylgeranyl pyrophosphate synthase [C...	288	1.30E-63
HGS073	gil143803	GerC3 [Bacillus subtilis] >gnl PID e118...	517	1.20E-82
HGS073	gnl PID d1009341	component II of heptaprenyl diphosphate...	506	5.20E-81
HGS073	gnl PID d1026196	(AB003188) component B of hexaprenyl di...	493	2.00E-69
HGS073	gil1813470	spore germination protein C3 [Bacillus ...	467	3.10E-60
HGS073	gil336639	prephytoene pyrophosphate dehydrogenase...	338	9.30E-50
HGS073	pir S76966 S76966	geranylgeranyl pyrophosphate synthase c...	352	1.20E-47
HGS073	dbj AB001997_1	(AB001997) solanesyl diphosphate syntha...	211	7.40E-44
HGS073	gil1276734	prenyl transferase [Porphyrin purpurea] ...	306	3.40E-42
HGS073	gnl PID d1031114	(AP000004) 342aa long hypothetical gera...	202	4.80E-42

HGS073	gil1573899	octaprenyl-diphosphate synthase (ispB) ...	296	1.40E-40
HGS074	gnlPID1d1032955	(AB004319) undecaprenyl diphosphate synt...	533	2.00E-69
HGS074	gnlPID1e1185244	similar to hypothetical proteins [Bacill...	450	5.80E-58
HGS074	gnlPID1d1011480	hypothetical protein [Synechocystis sp.]...	340	1.00E-42
HGS074	gil3328883	(AE001319) YaeS family [Chlamydia tracho...	324	1.60E-40
HGS074	gil1786371	(AE000127) orf, hypothetical protein [Es...	323	2.20E-40
HGS074	gnlPID1d1012616	unknown [Escherichia coli]	315	4.40E-39
HGS074	gil1573941	conserved hypothetical protein [Haemophi...	307	3.90E-38
HGS074	gil32242704	(AC003040) hypothetical protein [Arabido...	220	3.70E-37
HGS074	gnlPID1e1342726	(AJ235271) unknown [Rickettsia prowazeki...	188	1.20E-36
HGS074	gnlPID1e315162	hypothetical protein Rv2361c [Mycobacter...	301	1.20E-35
HGS075	gil4104603	(AF036966) putative histidine kinase [La...	426	1.60E-185
HGS075	gnlPID1d1011961	homologous to sp:PHOR_BACSU [Bacillus su...	517	8.60E-180
HGS075	gil2182992	histidine kinase [Lactococcus lactis cre...	300	1.30E-90
HGS075	gil410142	ORFX18 [Bacillus subtilis] >gnlPID1e118...	373	1.20E-63
HGS075	gil1575578	histidine protein kinase [Thermotoga mar...	248	6.50E-51
HGS075	gil143331	alkaline phosphatase regulatory protein ...	360	5.30E-49
HGS075	gil3687664	(AF049873) sensor protein [Lactococcus l...	202	5.40E-49
HGS075	gil288420	drug sensory protein A [Synechocystis PC...	114	5.90E-44
HGS075	gil2352098	histidine protein kinase; KinB [Pseudomo...	118	3.10E-38
HGS075	gil1276858	hypothetical chloroplast ORF 26 [Porphy...	102	3.20E-38
dead	gil1573195	ATP-dependent RNA helicase (dead) [H...	419	2.10E-121
dead	gil145727	dead [Escherichia coli]	405	2.00E-120
dead	gil149184	RNA helicase [Klebsiella pneumoniae]...	403	2.10E-120
dead	gnlPID1d1011207	ATP-dependent RNA helicase Dead [Syn...	810	7.20E-117
dead	gil606102	two frameshifts relative to ECODEAD ...	405	3.30E-116
dead	spIP23304 DEAD_EC	OLI ATP-DEPENDENT RNA HELICASE DEAD: >gi...	405	6.70E-116
dead	gnlPID1e254889	dead [Mycobacterium tuberculosis] >s...	356	1.60E-112

deaD	gjl2313340	(AE000544) ATP-dependent RNA helicase...	421	4.20E-112
deaD	gjl2621248	(AE000807) ATP-dependent RNA helicase...	437	1.70E-111
deaD	gjl4154758	(AE001461) ATP-DEPENDENT RNA HELICASE...	420	8.40E-108

Vectors and Host Cell

The present invention also relates to vectors which include the isolated DNA molecules of the present invention, host cells comprising the recombinant vectors, and the
5 production of *S. aureus* polypeptides and peptides of the present invention expressed by the host cells.

Recombinant constructs may be introduced into host cells using well known techniques such as infection, transduction, transfection, transvection, electroporation and transformation. The vector may be, for example, a phage, plasmid, viral or retroviral vector.
10 Retroviral vectors may be replication competent or replication defective. In the latter case, viral propagation generally will occur only in complementing host cells.

The *S. aureus* polynucleotides may be joined to a vector containing a selectable marker for propagation in a host. Generally, a plasmid vector is introduced in a precipitate, such as a calcium phosphate precipitate, or in a complex with a charged lipid. If the vector is
15 a virus, it may be packaged *in vitro* using an appropriate packaging cell line and then transduced into host cells.

Preferred are vectors comprising *cis*-acting control regions to the polynucleotide of interest. Appropriate *trans*-acting factors may be supplied by the host, supplied by a complementing vector or supplied by the vector itself upon introduction into the host.

20 In certain preferred embodiments in this regard, the vectors provide for specific expression, which may be inducible and/or cell type-specific. Particularly preferred among such vectors are those inducible by environmental factors that are easy to manipulate, such as temperature and nutrient additives.

Expression vectors useful in the present invention include chromosomal-, episomal-
25 and virus-derived vectors, *e.g.*, vectors derived from bacterial plasmids, bacteriophage, yeast episomes, yeast chromosomal elements, viruses such as baculoviruses, papova viruses, vaccinia viruses, adenoviruses, fowl pox viruses, pseudorabies viruses and retroviruses, and vectors derived from combinations thereof, such as cosmids and phagemids.

The DNA insert should be operatively linked to an appropriate promoter, such as the
30 phage lambda PL promoter, the *E. coli lac*, *trp*, *phoA* and *tac* promoters, the SV40 early and late promoters and promoters of retroviral LTRs, to name a few. Other suitable promoters will be known to the skilled artisan. The expression constructs will further contain sites for

transcription initiation, termination and, in the transcribed region, a ribosome binding site for translation. The coding portion of the mature transcripts expressed by the constructs will preferably include a translation initiating site at the beginning and a termination codon (UAA, UGA or UAG) appropriately positioned at the end of the polypeptide to be translated.

5 As indicated, the expression vectors will preferably include at least one selectable marker. Such markers include dihydrofolate reductase, G418 or neomycin resistance for eukaryotic cell culture and tetracycline, kanamycin, or ampicillin resistance genes for culturing in *E. coli* and other bacteria. Representative examples of appropriate hosts include, but are not limited to, bacterial cells, such as *E. coli*, *Streptomyces* and *Salmonella*
10 *typhimurium* cells; fungal cells, such as yeast cells (e.g., *Saccharomyces cerevisiae* or *Pichia pastoris* (ATCC Accession No. 201178)); insect cells such as *Drosophila* S2 and *Spodoptera* Sf9 cells; animal cells such as CHO, COS and Bowes melanoma cells; and plant cells. Appropriate culture mediums and conditions for the above-described host cells are known in the art.

15 Among vectors preferred for use in bacteria include pQE70, pQE60 and pQE9, pQE10 available from Qiagen; pBS vectors, Phagescript vectors, Bluescript vectors, pNH8A, pNH16a, pNH18A, pNH46A available from Stratagene Cloning Systems, Inc.; pET series of vectors available from Novagen; and ptrc99a, pKK223-3, pKK233-3, pDR540, pRITS available from Pharmacia Biotech, Inc. Among preferred eukaryotic vectors are pWLNEO,
20 pSV2CAT, pOG44, pXT1 and pSG available from Stratagene; and pSVK3, pBPV, pMSG and pSVL available from Pharmacia. Preferred expression vectors for use in yeast systems include, but are not limited to pYES2, pYD1, pTEF1/Zeo, pYES2/GS, pPICZ, pGAPZ, pGAPZalph, pPIC9, pPIC3.5, pHIL-D2, pHIL-S1, pPIC3.5K, pPIC9K, and PAO815 (all available from Invitrogen, Carlsbad, CA). Other suitable vectors will be readily apparent to the
25 skilled artisan.

Among known bacterial promoters suitable for use in the present invention include the *E. coli lacI* and *lacZ* promoters, the T3, T5 and T7 promoters, the *gpt* promoter, the lambda PR and PL promoters and the *trp* promoter. Suitable eukaryotic promoters include the CMV immediate early promoter, the HSV thymidine kinase promoter, the early and late
30 SV40 promoters; the promoters of retroviral LTRs, such as those of the Rous sarcoma virus (RSV), and metallothionein promoters, such as the mouse metallothionein-I promoter.

Introduction of the construct into the host cell can be effected by competent cell transformation, calcium phosphate transfection, DEAE-dextran mediated transfection, cationic lipid-mediated transfection, electroporation, transduction, infection or other methods. Such methods are described in many standard laboratory manuals (for example, Davis, *et al.*,
5 *Basic Methods In Molecular Biology* (1986)). It is specifically contemplated that the polypeptides of the present invention may in fact be expressed by a host cell lacking a recombinant vector.

Transcription of DNA encoding the polypeptides of the present invention by higher eukaryotes may be increased by inserting an enhancer sequence into the vector. Enhancers
10 are *cis*-acting elements of DNA, usually about from 10 to 300 nucleotides that act to increase transcriptional activity of a promoter in a given host cell-type. Examples of enhancers include the SV40 enhancer, which is located on the late side of the replication origin at nucleotides 100 to 270, the cytomegalovirus early promoter enhancer, the polyoma enhancer on the late side of the replication origin, and adenovirus enhancers.

15 For secretion of the translated polypeptide into the lumen of the endoplasmic reticulum, into the periplasmic space or into the extracellular environment, appropriate secretion signals may be incorporated into the expressed polypeptide, for example, the amino acid sequence KDEL. The signals may be endogenous to the polypeptide or they may be heterologous signals.

20 The polypeptide may be expressed in a modified form, such as a fusion protein, and may include not only secretion signals, but also additional heterologous functional regions. For instance, a region of additional amino acids, particularly charged amino acids, may be added to the N-terminus of the polypeptide to improve stability and persistence in the host cell, during purification, or during subsequent handling and storage. Also, peptide moieties
25 may be added to the polypeptide to facilitate purification. A preferred fusion protein comprises a Hexa-Histidine peptide fused inframe to the polypeptide of the invention. Such regions may be removed prior to final preparation of the polypeptide. The addition of peptide moieties to polypeptides to engender secretion or excretion, to improve stability and to facilitate purification, among others, are familiar and routine techniques in the art. A
30 preferred fusion protein comprises a heterologous region from immunoglobulin that is useful to solubilize proteins. For example, EP-A-O 464 533 (Canadian counterpart 2045869) discloses fusion proteins comprising various portions of constant region of immunoglobulin

molecules together with another human protein or part thereof. In many cases, the Fc part in a fusion protein is thoroughly advantageous for use in therapy and diagnosis and thus results, for example, in improved pharmacokinetic properties (EP-A 0232 262). On the other hand, for some uses it would be desirable to be able to delete the Fc part after the fusion protein has been expressed, detected and purified in the advantageous manner described. This is the case when Fc portion proves to be a hindrance to use in therapy and diagnosis, for example when the fusion protein is to be used as antigen for immunizations. In drug discovery, for example, human proteins, such as, hIL-5-receptor has been fused with Fc portions for the purpose of high-throughput screening assays to identify antagonists of hIL-5. See Bennett, D. et al. (1995) J. Molec. Recogn. 8:52-58 and Johanson, K. et al. (1995) J. Biol. Chem. 270 (16):9459-9471.

The *S. aureus* polypeptides can be recovered and purified from recombinant cell cultures by well-known methods including ammonium sulfate or ethanol precipitation, acid extraction, anion or cation exchange chromatography (e.g. a Nickel anion exchange column can be used to bind the Hexa-His tagged fusion protein), phosphocellulose chromatography, hydrophobic interaction chromatography, affinity chromatography, hydroxylapatite chromatography, lectin chromatography and high performance liquid chromatography ("HPLC") is employed for purification. Polypeptides of the present invention include naturally purified products, products of chemical synthetic procedures, and products produced by recombinant techniques from a prokaryotic or eukaryotic host, including, for example, bacterial, yeast, higher plant, insect and mammalian cells.

Depending upon the host employed in a recombinant production procedure, the polypeptides of the present invention may be glycosylated or may be non-glycosylated. In addition, polypeptides of the invention may also include an initial modified methionine residue, in some cases as a result of host-mediated processes. Thus, it is well known in the art that the N-terminal methionine encoded by the translation initiation codon generally is removed with high efficiency from any protein after translation in all eukaryotic cells. While the N-terminal methionine on most proteins also is efficiently removed in most prokaryotes, for some proteins, this prokaryotic removal process is inefficient, depending on the nature of the amino acid to which the N-terminal methionine is covalently linked.

In one embodiment, the yeast *Pichia pastoris* is used to express any plasma membrane associated protein of the invention in a eukaryotic system. *Pichia pastoris* is a

methylophilic yeast which can metabolize methanol as its sole carbon source. A main step in the methanol metabolism pathway is the oxidation of methanol to formaldehyde using O_2 . This reaction is catalyzed by the enzyme alcohol oxidase. In order to metabolize methanol as its sole carbon source, *Pichia pastoris* must generate high levels of alcohol oxidase due, in part, to the relatively low affinity of alcohol oxidase for O_2 . Consequently, in a growth medium depending on methanol as a main carbon source, the promoter region of one of the two alcohol oxidase genes (*AOX1*) is highly active. In the presence of methanol, alcohol oxidase produced from the *AOX1* gene comprises up to approximately 30% of the total soluble protein in *Pichia pastoris*. See, Ellis, S.B., et al., *Mol. Cell. Biol.* 5:1111-21 (1985); Koutz, P.J., et al., *Yeast* 5:167-77 (1989); Tschopp, J.F., et al., *Nucl. Acids Res.* 15:3859-76 (1987). Thus, a heterologous coding sequence, such as, for example, a plasma membrane associated polynucleotide of the present invention, under the transcriptional regulation of all or part of the *AOX1* regulatory sequence is expressed at exceptionally high levels in *Pichia* yeast grown in the presence of methanol.

In one example, the plasmid vector pPIC9K is used to express DNA encoding a plasma membrane associated polypeptide of the invention, as set forth herein, in a *Pichea* yeast system essentially as described in "*Pichia* Protocols: Methods in Molecular Biology," D.R. Higgins and J. Cregg, eds. The Humana Press, Totowa, NJ, 1998. This expression vector allows expression and secretion of a plasma membrane associated protein of the invention by virtue of the strong *AOX1* promoter linked to the *Pichia pastoris* alkaline phosphatase (PHO) secretory signal peptide (i.e., leader) located upstream of a multiple cloning site.

Many other yeast vectors could be used in place of pPIC9K, such as, pYES2, pYDI, pTEF1/Zeo, pYES2/GS, pPICZ, pGAPZ, pGAPZalpha, pPIC9, pPIC3.5, pHIL-D2, pHIL-S1, pPIC3.5K, and PAO815, as one skilled in the art would readily appreciate, as long as the proposed expression construct provides appropriately located signals for transcription, translation, secretion (if desired), and the like, including an in-frame AUG as required.

In another embodiment, high-level expression of a heterologous coding sequence, such as, for example, a plasma membrane associated polynucleotide of the present invention, may be achieved by cloning the heterologous polynucleotide of the invention into an

expression vector such as, for example, pGAPZ or pGAPZalpha, and growing the yeast culture in the absence of methanol.

In addition to encompassing host cells containing the vector constructs discussed herein, the invention also encompasses host cells that have been engineered to delete or replace endogenous genetic material (e.g. coding sequences for the polypeptides of the present invention), and/or to include genetic material (e.g. heterologous polynucleotide sequences) that is operably associated with polynucleotides of the present invention, and which activates, alters, and/or amplifies endogenous polynucleotides of the present invention. For example, techniques known in the art may be used to operably associate heterologous control regions (e.g. promoter and/or enhancer) and endogenous polynucleotide sequences via homologous recombination (see, e.g. U.S. Patent No. 5,641,670, issued June 24, 1997; International Publication No. WO 96/29411, published September 26, 1996; International Publication No. WO 94/12650, published August 4, 1994; Koller et al., Proc. Natl. Acad. Sci. USA 86:8932-8935 (1989); and Zijlstra, et al., Nature 342:435-438 (1989), the disclosures of each of which are hereby incorporated by reference in their entireties).

In addition, polypeptides of the invention can be chemically synthesized using techniques known in the art (e.g., see Creighton, 1983, Proteins: Structures and Molecular Principles, W.H. Freeman & Co., N.Y., and Hunkapiller et al., Nature, 310:105-111 (1984)). For example, a polypeptide corresponding to a fragment of a *S. aureus* polypeptide can be synthesized by use of a peptide synthesizer. Furthermore, if desired, nonclassical amino acids or chemical amino acid analogs can be introduced as a substitution or addition into the polypeptide sequence. Non-classical amino acids include, but are not limited to, to the D-isomers of the common amino acids, 2,4-diaminobutyric acid, α -amino isobutyric acid, 4-aminobutyric acid, Abu, 2-amino butyric acid, γ -Abu, ϵ -Ahx, 6-amino hexanoic acid, Aib, 2-amino isobutyric acid, 3-amino propionic acid, ornithine, norleucine, norvaline, hydroxyproline, sarcosine, citrulline, homocitrulline, cysteic acid, t-butylglycine, t-butylalanine, phenylglycine, cyclohexylalanine, β -alanine, fluoro-amino acids, designer amino acids such as β -methyl amino acids, Ca-methyl amino acids, Na-methyl amino acids, and amino acid analogs in general. Furthermore, the amino acid can be D (dextrorotary) or L (levorotary).

Non-naturally occurring variants may be produced using art-known mutagenesis techniques, which include, but are not limited to oligonucleotide mediated mutagenesis, alanine scanning, PCR mutagenesis, site directed mutagenesis (see, e.g., Carter et al., Nucl.

Acids Res. 13:4331 (1986); and Zoller *et al.*, *Nucl. Acids Res.* 10:6487 (1982)), cassette mutagenesis (*see, e.g.*, Wells *et al.*, *Gene* 34:315 (1985)), restriction selection mutagenesis (*see, e.g.*, Wells *et al.*, *Philos. Trans. R. Soc. London SerA* 317:415 (1986)).

The invention additionally, encompasses polypeptides of the present invention which are differentially modified during or after translation, such as for example, by glycosylation, acetylation, phosphorylation, amidation, derivatization by known protecting/blocking groups, proteolytic cleavage, linkage to an antibody molecule or other cellular ligand, etc. Any of numerous chemical modifications may be carried out by known techniques, including but not limited to specific chemical cleavage by cyanogen bromide, trypsin, chymotrypsin, papain, V8 protease, NaBH₄; acetylation, formylation, oxidation, reduction; metabolic synthesis in the presence of tunicamycin; etc.

Additional post-translational modifications encompassed by the invention include, for example, N-linked or O-linked carbohydrate chains, processing of N-terminal or C-terminal ends, attachment of chemical moieties to the amino acid backbone, chemical modifications of N-linked or O-linked carbohydrate chains, and addition or deletion of an N-terminal methionine residue as a result of alternative host cell expression. The polypeptides may also be modified with a detectable label, such as an enzymatic, fluorescent, isotopic or affinity label to allow for detection and isolation of the protein.

Also provided by the invention are chemically modified derivatives of the polypeptides of the invention which may provide additional advantages such as increased solubility, stability and circulating time of the polypeptide, or decreased immunogenicity (*see* U.S. Patent No. 4,179,337). The chemical moieties for derivitization may be selected from water soluble polymers such as polyethylene glycol, ethylene glycol/propylene glycol copolymers, carboxymethylcellulose, dextran, polyvinyl alcohol and the like. The polypeptides may be modified at random positions within the molecule, or at predetermined positions within the molecule and may include one, two, three or more attached chemical moieties.

The polymer may be of any molecular weight, and may be branched or unbranched. For polyethylene glycol, the preferred molecular weight is between about 1 kDa and about 100 kDa (the term "about" indicating that in preparations of polyethylene glycol, some molecules will weigh more, some less, than the stated molecular weight) for ease in handling and manufacturing. Other sizes may be used, depending on the desired therapeutic profile,

which can include, for example, the duration of sustained release desired, the effects, if any on biological activity, the ease in handling, the degree or lack of antigenicity and other known effects of the polyethylene glycol to a therapeutic protein or analog. For example, the polyethylene glycol may have an average molecular weight of about 200, 500, 1000, 1500, 2000, 2500, 3000, 3500, 4000, 4500, 5000, 5500, 6000, 6500, 7000, 7500, 8000, 8500, 9000, 9500, 10,000, 10,500, 11,000, 11,500, 12,000, 12,500, 13,000, 13,500, 14,000, 14,500, 15,000, 15,500, 16,000, 16,500, 17,000, 17,500, 18,000, 18,500, 19,000, 19,500, 20,000, 25,000, 30,000, 35,000, 40,000, 50,000, 55,000, 60,000, 65,000, 70,000, 75,000, 80,000, 85,000, 90,000, 95,000, or 100,000 kDa.

As noted above, the polyethylene glycol may have a branched structure. Branched polyethylene glycols are described, for example, in U.S. Patent No. 5,643,575; Morpurgo *et al.*, *Appl. Biochem. Biotechnol.* 56:59-72 (1996); Vorobjev *et al.*, *Nucleosides Nucleotides* 18:2745-2750 (1999); and Caliceti *et al.*, *Bioconjug. Chem.* 10:638-646 (1999), the disclosures of each of which are incorporated herein by reference in their entireties.

The polyethylene glycol molecules (or other chemical moieties) should be attached to the protein with consideration of effects on functional or antigenic domains of the protein. There are a number of attachment methods available to those skilled in the art, e.g., EP 0 401 384, herein incorporated by reference (coupling PEG to G-CSF), see also Malik *et al.*, *Exp. Hematol.* 20:1028-1035 (1992) (reporting pegylation of GM-CSF using tresyl chloride). For example, polyethylene glycol may be covalently bound through amino acid residues via a reactive group, such as, a free amino or carboxyl group. Reactive groups are those to which an activated polyethylene glycol molecule may be bound. The amino acid residues having a free amino group may include lysine residues and the N-terminal amino acid residues; those having a free carboxyl group may include aspartic acid residues glutamic acid residues and the C-terminal amino acid residue. Sulfhydryl groups may also be used as a reactive group for attaching the polyethylene glycol molecules. Preferred for therapeutic purposes is attachment at an amino group, such as attachment at the N-terminus or lysine group.

As suggested above, polyethylene glycol may be attached to proteins via linkage to any of a number of amino acid residues. For example, polyethylene glycol can be linked to a proteins via covalent bonds to lysine, histidine, aspartic acid, glutamic acid, or cysteine residues. One or more reaction chemistries may be employed to attach polyethylene glycol to specific amino acid residues (e.g., lysine, histidine, aspartic acid, glutamic acid, or cysteine)

of the protein or to more than one type of amino acid residue (e.g., lysine, histidine, aspartic acid, glutamic acid, cysteine and combinations thereof) of the protein.

One may specifically desire proteins chemically modified at the N-terminus. Using polyethylene glycol as an illustration of the present composition, one may select from a variety of polyethylene glycol molecules (by molecular weight, branching, etc.), the proportion of polyethylene glycol molecules to protein (polypeptide) molecules in the reaction mix, the type of pegylation reaction to be performed, and the method of obtaining the selected N-terminally pegylated protein. The method of obtaining the N-terminally pegylated preparation (i.e., separating this moiety from other monopegylated moieties if necessary) may be by purification of the N-terminally pegylated material from a population of pegylated protein molecules. Selective proteins chemically modified at the N-terminus modification may be accomplished by reductive alkylation which exploits differential reactivity of different types of primary amino groups (lysine versus the N-terminal) available for derivatization in a particular protein. Under the appropriate reaction conditions, substantially selective derivatization of the protein at the N-terminus with a carbonyl group containing polymer is achieved.

As indicated above, pegylation of the proteins of the invention may be accomplished by any number of means. For example, polyethylene glycol may be attached to the protein either directly or by an intervening linker. Linkerless systems for attaching polyethylene glycol to proteins are described in Delgado *et al.*, *Crit. Rev. Thera. Drug Carrier Sys.* 9:249-304 (1992); Francis *et al.*, *Intern. J. of Hematol.* 68:1-18 (1998); U.S. Patent No. 4,002,531; U.S. Patent No. 5,349,052; WO 95/06058; and WO 98/32466, the disclosures of each of which are incorporated herein by reference.

One system for attaching polyethylene glycol directly to amino acid residues of proteins without an intervening linker employs tresylated MPEG, which is produced by the modification of monmethoxy polyethylene glycol (MPEG) using tresylchloride ($\text{ClSO}_2\text{CH}_2\text{CF}_3$). Upon reaction of protein with tresylated MPEG, polyethylene glycol is directly attached to amine groups of the protein. Thus, the invention includes protein-polyethylene glycol conjugates produced by reacting proteins of the invention with a polyethylene glycol molecule having a 2,2,2-trifluoroethane sulphonyl group.

Polyethylene glycol can also be attached to proteins using a number of different intervening linkers. For example, U.S. Patent No. 5,612,460, the entire disclosure of which is

incorporated herein by reference, discloses urethane linkers for connecting polyethylene glycol to proteins. Protein-polyethylene glycol conjugates wherein the polyethylene glycol is attached to the protein by a linker can also be produced by reaction of proteins with compounds such as MPEG-succinimidylsuccinate, MPEG activated with 1,1'-carbonyldiimidazole, MPEG-2,4,5-trichloropenylcarbonate, MPEG-p-nitrophenolcarbonate, and various MPEG-succinate derivatives. A number additional polyethylene glycol derivatives and reaction chemistries for attaching polyethylene glycol to proteins are described in WO 98/32466, the entire disclosure of which is incorporated herein by reference. Pegylated protein products produced using the reaction chemistries set out herein are included within the scope of the invention.

The number of polyethylene glycol moieties attached to each protein of the invention (*i.e.*, the degree of substitution) may also vary. For example, the pegylated proteins of the invention may be linked, on average, to 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 12, 15, 17, 20, or more polyethylene glycol molecules. Similarly, the average degree of substitution within ranges such as 1-3, 2-4, 3-5, 4-6, 5-7, 6-8, 7-9, 8-10, 9-11, 10-12, 11-13, 12-14, 13-15, 14-16, 15-17, 16-18, 17-19, or 18-20 polyethylene glycol moieties per protein molecule. Methods for determining the degree of substitution are discussed, for example, in Delgado *et al.*, *Crit. Rev. Thera. Drug Carrier Sys.* 9:249-304 (1992).

The polypeptides of the invention may be in monomers or multimers (*i.e.*, dimers, trimers, tetramers and higher multimers). Accordingly, the present invention relates to monomers and multimers of the polypeptides of the invention, their preparation, and compositions (preferably, Therapeutics) containing them. In specific embodiments, the polypeptides of the invention are monomers, dimers, trimers or tetramers. In additional embodiments, the multimers of the invention are at least dimers, at least trimers, or at least tetramers.

Multimers encompassed by the invention may be homomers or heteromers. As used herein, the term homomer, refers to a multimer containing polypeptides corresponding to only one of the amino acid sequences of Table 1 (including fragments, variants, splice variants, and fusion proteins, corresponding to these as described herein). These homomers may contain polypeptides having identical or different amino acid sequences. In a specific embodiment, a homomer of the invention is a multimer containing only polypeptides having an identical amino acid sequence. In another specific embodiment, a homomer of the

invention is a multimer containing polypeptides having different amino acid sequences. In specific embodiments, the multimer of the invention is a homodimer (e.g., containing polypeptides having identical or different amino acid sequences) or a homotrimer (e.g., containing polypeptides having identical and/or different amino acid sequences). In
5 additional embodiments, the homomeric multimer of the invention is at least a homodimer, at least a homotrimer, or at least a homotetramer.

As used herein, the term heteromer refers to a multimer containing one or more heterologous polypeptides (i.e., polypeptides of different proteins) in addition to the polypeptides of the invention. In a specific embodiment, the multimer of the invention is a
10 heterodimer, a heterotrimer, or a heterotetramer. In additional embodiments, the heteromeric multimer of the invention is at least a heterodimer, at least a heterotrimer, or at least a heterotetramer.

Multimers of the invention may be the result of hydrophobic, hydrophilic, ionic and/or covalent associations and/or may be indirectly linked, by for example, liposome
15 formation. Thus, in one embodiment, multimers of the invention, such as, for example, homodimers or homotrimers, are formed when polypeptides of the invention contact one another in solution. In another embodiment, heteromultimers of the invention, such as, for example, heterotrimers or heterotetramers, are formed when polypeptides of the invention contact antibodies to the polypeptides of the invention (including antibodies to the
20 heterologous polypeptide sequence in a fusion protein of the invention) in solution. In other embodiments, multimers of the invention are formed by covalent associations with and/or between the polypeptides of the invention. Such covalent associations may involve one or more amino acid residues contained in the polypeptide sequence (e.g., the polypeptide sequences shown in Table 1). In one instance, the covalent associations are cross-linking
25 between cysteine residues located within the polypeptide sequences which interact in the native (i.e., naturally occurring) polypeptide. In another instance, the covalent associations are the consequence of chemical or recombinant manipulation. Alternatively, such covalent associations may involve one or more amino acid residues contained in the heterologous polypeptide sequence in a fusion protein. In one example, covalent associations are between
30 the heterologous sequence contained in a fusion protein of the invention (see, e.g., US Patent Number 5,478,925). In a specific example, the covalent associations are between the heterologous sequence contained in a Fc fusion protein of the invention (as described herein).

In another specific example, covalent associations of fusion proteins of the invention are between heterologous polypeptide sequence from another protein that is capable of forming covalently associated multimers, such as for example, osteopontin (see, International Publication NO: WO 98/49305, the contents of which is incorporated herein incorporated by reference in its entirety). In another embodiment, two or more polypeptides of the invention are joined through peptide linkers. Examples include those peptide linkers described in U.S. Pat. No. 5,073,627 (incorporated herein by reference in its entirety). Proteins comprising multiple polypeptides of the invention separated by peptide linkers may be produced using conventional recombinant DNA technology.

Another method for preparing multimer polypeptides of the invention involves use of polypeptides of the invention fused to a leucine zipper or isoleucine zipper polypeptide sequence. Leucine zipper and isoleucine zipper domains are polypeptides that promote multimerization of the proteins in which they are found. Leucine zippers were originally identified in several DNA-binding proteins (Landschulz et al., Science 240:1759, (1988)), and have since been found in a variety of different proteins. Among the known leucine zippers are naturally occurring peptides and derivatives thereof that dimerize or trimerize. Examples of leucine zipper domains suitable for producing soluble multimeric proteins of the invention are those described in PCT application WO 94/10308, hereby incorporated by reference. Recombinant fusion proteins comprising a polypeptide of the invention fused to a polypeptide sequence that dimerizes or trimerizes in solution are expressed in suitable host cells, and the resulting soluble multimeric fusion protein is recovered from the culture supernatant using techniques known in the art.

Trimeric polypeptides of the invention may offer the advantage of enhanced biological activity. Preferred leucine zipper moieties and isoleucine moieties are those that preferentially form trimers. One example is a leucine zipper derived from lung surfactant protein D (SPD), as described in Hoppe et al. (FEBS Letters 344:191, (1994)) and in U.S. patent application Ser. No. 08/446,922, hereby incorporated by reference. Other peptides derived from naturally occurring trimeric proteins may be employed in preparing trimeric polypeptides of the invention.

In another example, proteins of the invention are associated by interactions between Flag® polypeptide sequence contained in fusion proteins of the invention containing Flag® polypeptide sequence. In a further embodiment, associations proteins of the invention are

associated by interactions between heterologous polypeptide sequence contained in Flag® fusion proteins of the invention and anti-Flag® antibody.

The multimers of the invention may be generated using chemical techniques known in the art. For example, polypeptides desired to be contained in the multimers of the invention may be chemically cross-linked using linker molecules and linker molecule length optimization techniques known in the art (see, e.g., US Patent Number 5,478,925, which is incorporated herein by reference in its entirety). Additionally, multimers of the invention may be generated using techniques known in the art to form one or more inter-molecule cross-links between the cysteine residues located within the sequence of the polypeptides desired to be contained in the multimer (see, e.g., US Patent Number 5,478,925, which is incorporated herein by reference in its entirety). Further, polypeptides of the invention may be routinely modified by the addition of cysteine or biotin to the C-terminus or N-terminus of the polypeptide and techniques known in the art may be applied to generate multimers containing one or more of these modified polypeptides (see, e.g., US Patent Number 5,478,925, which is incorporated herein by reference in its entirety). Additionally, techniques known in the art may be applied to generate liposomes containing the polypeptide components desired to be contained in the multimer of the invention (see, e.g., US Patent Number 5,478,925, which is incorporated herein by reference in its entirety).

Alternatively, multimers of the invention may be generated using genetic engineering techniques known in the art. In one embodiment, polypeptides contained in multimers of the invention are produced recombinantly using fusion protein technology described herein or otherwise known in the art (see, e.g., US Patent Number 5,478,925, which is incorporated herein by reference in its entirety). In a specific embodiment, polynucleotides coding for a homodimer of the invention are generated by ligating a polynucleotide sequence encoding a polypeptide of the invention to a sequence encoding a linker polypeptide and then further to a synthetic polynucleotide encoding the translated product of the polypeptide in the reverse orientation from the original C-terminus to the N-terminus (lacking the leader sequence) (see, e.g., US Patent Number 5,478,925, which is incorporated herein by reference in its entirety). In another embodiment, recombinant techniques described herein or otherwise known in the art are applied to generate recombinant polypeptides of the invention which contain a transmembrane domain (or hydrophobic or signal peptide) and which can be incorporated by

membrane reconstitution techniques into liposomes (see, e.g., US Patent Number 5,478,925, which is incorporated herein by reference in its entirety).

5 *Polypeptides and Fragments*

The invention further provides an isolated *S. aureus* polypeptide having an amino acid sequence in Table 1, or a peptide or polypeptide comprising a portion, fragment, variant or analog of the above polypeptides.

10 In the present invention, a "polypeptide fragment" refers to a short amino acid sequence contained in any one of the polypeptide sequences shown in Table 1 or encoded by the DNA contained in the deposit. Protein fragments may be "free-standing," or comprised within a larger polypeptide of which the fragment forms a part or region, most preferably as a single continuous region. Representative examples of polypeptide fragments of the invention, include, for example, fragments from about amino acid number 1-20, 21-40, 41-
15 60, 61-80, 81-100, 102-120, 121-140, 141-160, or 161 to the end of the coding region. Moreover, polypeptide fragments can be about 20, 30, 40, 50, 60, 70, 80, 90, 100, 110, 120, 130, 140, or 150 amino acids in length. In this context "about" includes the particularly recited ranges, larger or smaller by several (5, 4, 3, 2, or 1) amino acids, at either extreme or at both extremes.

20 Preferred polypeptide fragments include the mature form. Further preferred polypeptide fragments include the mature form having a continuous series of deleted residues from the amino or the carboxy terminus, or both. For example, any number of amino acids, ranging from 1-60, can be deleted from the amino terminus the mature form. Similarly, any number of amino acids, ranging from 1-30, can be deleted from the carboxy terminus of the
25 mature form. Furthermore, any combination of the above amino and carboxy terminus deletions are preferred. Similarly, polynucleotide fragments encoding these polypeptide fragments are also preferred.

Also preferred are polypeptide and polynucleotide fragments characterized by structural or functional domains, such as fragments that comprise alpha-helix and alpha-helix
30 forming regions, beta-sheet and beta-sheet-forming regions, turn and turn-forming regions, coil and coil-forming regions, hydrophilic regions, hydrophobic regions, alpha amphipathic regions, beta amphipathic regions, flexible regions, surface-forming regions, substrate

binding region, and high antigenic index regions. Polypeptide fragments of the sequences shown in Table 1 falling within conserved domains are specifically contemplated by the present invention. Moreover, polynucleotide fragments encoding these domains are also contemplated.

5 Other preferred fragments are biologically active fragments. Biologically active fragments are those exhibiting activity similar, but not necessarily identical, to an activity of the polypeptide of the present invention. The biological activity of the fragments may include an improved desired activity, or a decreased undesirable activity.

10 *Variant and Mutant Polypeptides*

To improve or alter the characteristics of *S. aureus* polypeptides of the present invention, protein engineering may be employed. Recombinant DNA technology known to those skilled in the art can be used to create novel mutant proteins or muteins including single or multiple amino acid substitutions, deletions, additions, or fusion proteins. Such modified polypeptides can show, e.g., increased/decreased activity or increased/decreased stability. In addition, they may be purified in higher yields and show better solubility than the corresponding natural polypeptide, at least under certain purification and storage conditions. Further, the polypeptides of the present invention may be produced as multimers including dimers, trimers and tetramers. Multimerization may be facilitated by linkers or recombinantly though fused heterologous polypeptides such as Fc regions.

N-Terminal and C-Terminal Deletion Mutants

25 It is known in the art that one or more amino acids may be deleted from the N-terminus or C-terminus without substantial loss of biological function. For instance, Ron et al. J. Biol. Chem., 268:2984-2988 (1993), reported modified KGF proteins that had heparin binding activity even if 3, 8, or 27 N-terminal amino acid residues were missing. Accordingly, the present invention provides polypeptides having one or more residues deleted from the amino terminus of the polypeptides shown in Table 1.

30 Similarly, many examples of biologically functional C-terminal deletion mutants are known. For instance, Interferon gamma shows up to ten times higher activities by deleting 8-10 amino acid residues from the carboxy terminus of the protein See, e.g., Dobeli, et al. (1988) J. Biotechnology 7:199-216. Accordingly, the present invention provides polypeptides having one or more residues from the carboxy terminus of the polypeptides shown in Table 1. The invention also provides polypeptides having one or more amino acids

deleted from both the amino and the carboxyl termini as described below.

The polypeptide fragments of the present invention can be immediately envisaged using the above description and are therefore not individually listed solely for the purpose of not unnecessarily lengthening the specification.

5 The present invention is further directed to polynucleotide encoding portions or fragments of the amino acid sequences described herein as well as to portions or fragments of the isolated amino acid sequences described herein. Fragments include portions of the amino acid sequences of Table 1, at least 7 contiguous amino acid in length, selected from any two integers, one of which representing a N-terminal position. The first codon of the
10 polypeptides of Table 1 is position 1. Every combination of a N-terminal and C-terminal position that a fragment at least 7 contiguous amino acid residues in length could occupy, on any given amino acid sequence of Table 1 is included in the invention. At least means a fragment may be 7 contiguous amino acid residues in length or any integer between 7 and the number of residues in a full length amino acid sequence minus 1. Therefore, included in the
15 invention are contiguous fragments specified by any N-terminal and C-terminal positions of amino acid sequence set forth in Table 1 wherein the contiguous fragment is any integer between 7 and the number of residues in a full length sequence minus 1.

Further, the invention includes polypeptides comprising fragments specified by size, in amino acid residues, rather than by N-terminal and C-terminal positions. The invention
20 includes any fragment size, in contiguous amino acid residues, selected from integers between 7 and the number of residues in a full length sequence minus 1. Preferred sizes of contiguous polypeptide fragments include about 7 amino acid residues, about 10 amino acid residues, about 20 amino acid residues, about 30 amino acid residues, about 40 amino acid residues, about 50 amino acid residues, about 100 amino acid residues, about 200 amino acid
25 residues, about 300 amino acid residues, and about 400 amino acid residues. The preferred sizes are, of course, meant to exemplify, not limit, the present invention as all size fragments representing any integer between 7 and the number of residues in a full length sequence minus 1 are included in the invention. The present invention also provides for the exclusion of any fragments specified by N-terminal and C-terminal positions or by size in amino acid
30 residues as described above. Any number of fragments specified by N-terminal and C-terminal positions or by size in amino acid residues as described above may be excluded.

Moreover, polypeptide fragments can be at least 10, 20, 30, 40, 50, 60, 70, 80, 90, 100, 110, 120, 130, 140, 150, 175 or 200 amino acids in length. Polynucleotides encoding these polypeptides are also encompassed by the invention. In this context "about" includes

the particularly recited ranges, larger or smaller by several (5, 4, 3, 2, or 1) amino acids, at either extreme or at both extremes.

The present invention further provides polypeptides having one or more residues deleted from the amino terminus of the amino acid sequence of a polypeptide disclosed herein (e.g., any polypeptide of Table 1). In particular, N-terminal deletions may be described by the general formula m-q, where q is a whole integer representing the total number of amino acid residues in a polypeptide of the invention (e.g., a polypeptide disclosed in Table 1), and m is defined as any integer ranging from 2 to q-6. Polynucleotides encoding these polypeptides are also encompassed by the invention.

The present invention further provides polypeptides having one or more residues from the carboxy-terminus of the amino acid sequence of a polypeptide disclosed herein (e.g., a polypeptide disclosed in Table 1). In particular, C-terminal deletions may be described by the general formula 1-n, where n is any whole integer ranging from 6 to q-1, and where n corresponds to the position of amino acid residue in a polypeptide of the invention.

Polynucleotides encoding these polypeptides are also encompassed by the invention.

In addition, any of the above described N- or C-terminal deletions can be combined to produce a N- and C-terminal deleted polypeptide. The invention also provides polypeptides having one or more amino acids deleted from both the amino and the carboxyl termini, which may be described generally as having residues m-n of a polypeptide encoded by a nucleotide sequence (e.g., including, but not limited to the preferred polypeptide disclosed in Table 1), or the cDNA contained in a deposited clone, and/or the complement thereof, where n and m are integers as described above. Polynucleotides encoding these polypeptides are also encompassed by the invention.

The polypeptide fragments of the present invention can be immediately envisaged using the above description and are therefore not individually listed solely for the purpose of not unnecessarily lengthening the specification.

The above fragments need not be active since they would be useful, for example, in immunoassays, in epitope mapping, epitope tagging, to generate antibodies to a particular portion of the polypeptide, as vaccines, and as molecular weight markers.

Other Mutants

In addition to N- and C-terminal deletion forms of the protein discussed above, it also

will be recognized by one of ordinary skill in the art that some amino acid sequences of the *S. aureus* polypeptides of the present invention can be varied without significant effect of the structure or function of the protein. If such differences in sequence are contemplated, it should be remembered that there will be critical areas on the protein which determine activity.

Thus, the invention further includes variations of the *S. aureus* polypeptides which show substantial *S. aureus* polypeptide activity or which include regions of *S. aureus* protein such as the protein portions discussed below. Such mutants include deletions, insertions, inversions, repeats, and substitutions selected according to general rules known in the art so as to have little effect on activity. For example, guidance concerning how to make phenotypically silent amino acid substitutions is provided. There are two main approaches for studying the tolerance of an amino acid sequence to change. See, Bowie, J. U. *et al.* (1990), Science 247:1306-1310. The first method relies on the process of evolution, in which mutations are either accepted or rejected by natural selection. The second approach uses genetic engineering to introduce amino acid changes at specific positions of a cloned gene and selections or screens to identify sequences that maintain functionality.

These studies have revealed that proteins are surprisingly tolerant of amino acid substitutions. The studies indicate which amino acid changes are likely to be permissive at a certain position of the protein. For example, most buried amino acid residues require nonpolar side chains, whereas few features of surface side chains are generally conserved. Other such phenotypically silent substitutions are described by Bowie *et al.* (*supra*) and the references cited therein. Typically seen as conservative substitutions are the replacements, one for another, among the aliphatic amino acids Ala, Val, Leu and Ile; interchange of the hydroxyl residues Ser and Thr, exchange of the acidic residues Asp and Glu, substitution between the amide residues Asn and Gln, exchange of the basic residues Lys and Arg and replacements among the aromatic residues Phe, Tyr.

Thus, the fragment, derivative, analog, or homolog of the polypeptide of Table I may be, for example: (i) one in which one or more of the amino acid residues are substituted with a conserved or non-conserved amino acid residue (preferably a conserved amino acid residue) and such substituted amino acid residue may or may not be one encoded by the genetic code; or (ii) one in which one or more of the amino acid residues includes a substituent group; or (iii) one in which the *S. aureus* polypeptide is fused with another compound, such as a compound to increase the half-life of the polypeptide (for example, polyethylene glycol); or (iv) one in which the additional amino acids are fused to the above form of the polypeptide, such as an Hexa-Histidine tag peptide or leader or secretory sequence or a sequence which is employed for purification of the above form of the polypeptide or a proprotein sequence.

Such fragments, derivatives and analogs are deemed to be within the scope of those skilled in the art from the teachings herein.

- Thus, the *S. aureus* polypeptides of the present invention may include one or more amino acid substitutions, deletions, or additions, either from natural mutations or human manipulation. As indicated, changes are preferably of a minor nature, such as conservative amino acid substitutions that do not significantly affect the folding or activity of the protein (see Table 3).

TABLE 3. Conservative Amino Acid Substitutions.

Aromatic	Phenylalanine Tryptophan Tyrosine
Hydrophobic	Leucine Isoleucine Valine
Polar	Glutamine Asparagine
Basic	Arginine Lysine Histidine
Acidic	Aspartic Acid Glutamic Acid
Small	Alanine Serine Threonine Methionine Glycine

Amino acids in the *S. aureus* proteins of the present invention that are essential for function can be identified by methods known in the art, such as site-directed mutagenesis or alanine-scanning mutagenesis. See, e.g., Cunningham et al. (1989) Science 244:1081-1085.

The latter procedure introduces single alanine mutations at every residue in the molecule. The resulting mutant molecules are then tested for biological activity using assays appropriate for measuring the function of the particular protein.

Of special interest are substitutions of charged amino acids with other charged or neutral amino acids which may produce proteins with highly desirable improved characteristics, such as less aggregation. Aggregation may not only reduce activity but also be problematic when preparing pharmaceutical formulations, because aggregates can be immunogenic. See, e.g., Pinckard et al., (1967) Clin. Exp. Immunol. 2:331-340; Robbins, et al., (1987) Diabetes 36:838-845; Cleland, et al., (1993) Crit. Rev. Therapeutic Drug Carrier Systems 10:307-377.

The polypeptides of the present invention are preferably provided in an isolated form, and may partially or substantially purified. A recombinantly produced version of the *S. aureus* polypeptide can be substantially purified by the one-step method described by Smith et al. (1988) Gene 67:31-40. Polypeptides of the invention also can be purified from natural or recombinant sources using antibodies directed against the polypeptides of the invention in methods which are well known in the art of protein purification. The purity of the polypeptide of the present invention may also specified in percent purity as relative to heterologous containing polypeptides. Preferred purities include at least 25%, 50%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5%, 99.75%, and 100% pure, as relative to heretologous containing polypeptides.

The invention provides for isolated *S. aureus* proteins comprising, or alternatively consisting of, polypeptides having an amino acid sequence selected from the group consisting of: (a) a full-length *S. aureus* polypeptide having the complete amino acid sequence shown in Table 1, (b) a full-length *S. aureus* polypeptide having the complete amino acid sequence shown in Table 1—excepting the N-terminal codon (e.g., including but not limited to, methionine, leucine, and/or valine), (c) an antigenic fragment of any of the polypeptides shown in Table 1, (d) a biologically active fragment of any of the polypeptides shown in Table 1, (e) a polypeptide encoded by any of the polynucleotide sequences shown in Table 1, and (f) a polypeptide shown in Table 1. The polypeptides of the present invention also include polypeptides having an amino acid sequence at least 80% identical, more preferably at least 90% identical, and still more preferably 95%, 96%, 97%, 98% or 99% identical to those described in (a), (b), (c), (d), (e) or (f) above. Further polypeptides of the present invention include polypeptides which have at least 90% similarity, more preferably at least

95% similarity, and still more preferably at least 96%, 97%, 98% or 99% similarity to those described above. Polynucleotides encoding these polypeptides are also encompassed by the invention.

5 A further embodiment of the invention relates to a polypeptide which comprises the amino acid sequence of a *S. aureus* polypeptide having an amino acid sequence which contains at least one conservative amino acid substitution, but not more than 50 conservative amino acid substitutions, not more than 40 conservative amino acid substitutions, not more than 30 conservative amino acid substitutions, and not more than 20 conservative amino acid substitutions. Also provided are polypeptides which comprise the amino acid sequence of a
10 *S. aureus* polypeptide, having at least one, but not more than 10, 9, 8, 7, 6, 5, 4, 3, 2 or 1 conservative amino acid substitutions.

By a polypeptide having an amino acid sequence at least, for example, 95% "identical" to a query amino acid sequence of the present invention, it is intended that the amino acid sequence of the subject polypeptide is identical to the query sequence except that
15 the subject polypeptide sequence may include up to five amino acid alterations per each 100 amino acids of the query amino acid sequence. In other words, to obtain a polypeptide having an amino acid sequence at least 95% identical to a query amino acid sequence, up to 5% (5 of 100) of the amino acid residues in the subject sequence may be inserted, deleted, (indels) or substituted with another amino acid. These alterations of the reference sequence
20 may occur at the amino or carboxy terminal positions of the reference amino acid sequence or anywhere between those terminal positions, interspersed either individually among residues in the reference sequence or in one or more contiguous groups within the reference sequence.

As a practical matter, whether any particular polypeptide is at least 80%, 85%, 90%, 95%, 96%, 97%, 98% or 99% identical to, for instance, the amino acid sequences shown in
25 Table 1, or a fragment thereof, can be determined conventionally using known computer programs. A preferred method for determining the best overall match between a query sequence (a sequence of the present invention) and a subject sequence, also referred to as a global sequence alignment, can be determined using the FASTDB computer program based on the algorithm of Brutlag et al., (1990) Comp. App. Biosci. 6:237-245. In a sequence
30 alignment the query and subject sequences are both amino acid sequences. The result of said global sequence alignment is in percent identity. Preferred parameters used in a FASTDB amino acid alignment are: Matrix=PAM 0, k-tuple=2, Mismatch Penalty=1, Joining Penalty=20, Randomization Group Length=0, Cutoff Score=1, Window Size=sequence length, Gap Penalty=5, Gap Size Penalty=0.05, Window Size=500 or the length of the subject
35 amino acid sequence, whichever is shorter.

If the subject sequence is shorter than the query sequence due to N- or C-terminal

deletions, not because of internal deletions, the results, in percent identity, must be manually corrected. This is because the FASTDB program does not account for N- and C-terminal truncations of the subject sequence when calculating global percent identity. For subject sequences truncated at the N- and C-termini, relative to the query sequence, the percent identity is corrected by calculating the number of residues of the query sequence that are N- and C-terminal of the subject sequence, which are not matched/aligned with a corresponding subject residue, as a percent of the total bases of the query sequence. Whether a residue is matched/aligned is determined by results of the FASTDB sequence alignment. This percentage is then subtracted from the percent identity, calculated by the above FASTDB program using the specified parameters, to arrive at a final percent identity score. This final percent identity score is what is used for the purposes of the present invention. Only residues to the N- and C-termini of the subject sequence, which are not matched/aligned with the query sequence, are considered for the purposes of manually adjusting the percent identity score. That is, only query amino acid residues outside the farthest N- and C-terminal residues of the subject sequence.

For example, a 90 amino acid residue subject sequence is aligned with a 100 residue query sequence to determine percent identity. The deletion occurs at the N-terminus of the subject sequence and therefore, the FASTDB alignment does not match/align with the first 10 residues at the N-terminus. The 10 unpaired residues represent 10% of the sequence (number of residues at the N- and C-termini not matched/total number of residues in the query sequence) so 10% is subtracted from the percent identity score calculated by the FASTDB program. If the remaining 90 residues were perfectly matched the final percent identity would be 90%. In another example, a 90 residue subject sequence is compared with a 100 residue query sequence. This time the deletions are internal so there are no residues at the N- or C-termini of the subject sequence which are not matched/aligned with the query. In this case the percent identity calculated by FASTDB is not manually corrected. Once again, only residue positions outside the N- and C-terminal ends of the subject sequence, as displayed in the FASTDB alignment, which are not matched/aligned with the query sequence are manually corrected. No other manual corrections are to be made for the purposes of the present invention.

The above polypeptide sequences are included irrespective of whether they have their normal biological activity. This is because even where a particular polypeptide molecule does not have biological activity, one of skill in the art would still know how to use the polypeptide, for instance, as a vaccine or to generate antibodies. Other uses of the polypeptides of the present invention that do not have *S. aureus* activity include, *inter alia*, as epitope tags, in epitope mapping, and as molecular weight markers on SDS-PAGE gels or on

molecular sieve gel filtration columns using methods known to those of skill in the art.

As described below, the polypeptides of the present invention can also be used to raise polyclonal and monoclonal antibodies, which are useful in assays for detecting *S. aureus* protein expression or as agonists and antagonists capable of enhancing or inhibiting *S. aureus* protein function. Further, such polypeptides can be used in the yeast two-hybrid system to "capture" *S. aureus* protein binding proteins which are also candidate agonists and antagonists according to the present invention. See, e.g., Fields et al. (1989) Nature 340:245-246.

10 Antibodies

Further polypeptides of the invention relate to antibodies and T-cell antigen receptors (TCR) which immunospecifically bind a polypeptide, polypeptide fragment, or variant of any one of the polypeptide sequences in Table 1, and/or an epitope, of the present invention (as determined by immunoassays well known in the art for assaying specific antibody-antigen binding). Antibodies of the invention include, but are not limited to, polyclonal, monoclonal, multispecific, human, humanized or chimeric antibodies, single chain antibodies, Fab fragments, F(ab') fragments, fragments produced by a Fab expression library, anti-idiotypic (anti-Id) antibodies (including, e.g., anti-Id antibodies to antibodies of the invention), and epitope-binding fragments of any of the above. The term "antibody," as used herein, refers to immunoglobulin molecules and immunologically active portions of immunoglobulin molecules, i.e., molecules that contain an antigen binding site that immunospecifically binds an antigen. The immunoglobulin molecules of the invention can be of any type (e.g., IgG, IgE, IgM, IgD, IgA and IgY), class (e.g., IgG1, IgG2, IgG3, IgG4, IgA1 and IgA2) or subclass of immunoglobulin molecule. In a specific embodiment, the immunoglobulin molecules of the invention are IgG1. In another specific embodiment, the immunoglobulin molecules of the invention are IgG4.

Most preferably the antibodies are human antigen-binding antibody fragments of the present invention and include, but are not limited to, Fab, Fab' and F(ab')₂, Fd, single-chain Fvs (scFv), single-chain antibodies, disulfide-linked Fvs (sdFv) and fragments comprising either a VL or VH domain. Antigen-binding antibody fragments, including single-chain antibodies, may comprise the variable region(s) alone or in combination with the entirety or a portion of the following: hinge region, CH1, CH2, and CH3 domains. Also included in the invention are antigen-binding fragments also comprising any combination of variable

region(s) with a hinge region, CH1, CH2, and CH3 domains. The antibodies of the invention may be from any animal origin including birds and mammals. Preferably, the antibodies are human, murine (e.g., mouse and rat), donkey, sheep, rabbit, goat, guinea pig, camel, horse, or chicken. As used herein, "human" antibodies include antibodies having the amino acid sequence of a human immunoglobulin and include antibodies isolated from human immunoglobulin libraries or from animals transgenic for one or more human immunoglobulin and that do not express endogenous immunoglobulins, as described infra and, for example in, U.S. Patent No. 5,939,598 by Kucherlapati et al.

The antibodies of the present invention may be monospecific, bispecific, trispecific or of greater multispecificity. Multispecific antibodies may be specific for different epitopes of a polypeptide of the present invention or may be specific for both a polypeptide of the present invention as well as for a heterologous epitope, such as a heterologous polypeptide or solid support material. See, e.g., PCT publications WO 93/17715; WO 92/08802; WO 91/00360; WO 92/05793; Tutt, et al., J. Immunol. 147:60-69 (1991); U.S. Patent Nos. 4,474,893; 4,714,681; 4,925,648; 5,573,920; 5,601,819; Kostelny et al., J. Immunol. 148:1547-1553 (1992).

Antibodies of the present invention may be described or specified in terms of the epitope(s) or portion(s) of a polypeptide of the present invention which they recognize or specifically bind. The epitope(s) or polypeptide portion(s) may be specified as described herein, e.g., by N-terminal and C-terminal positions, by size in contiguous amino acid residues, or listed in the Table 4 below. Preferred epitopes of the invention include the predicted antigenic epitopes shown in Table 4, below. It is pointed out that Table 4 only lists amino acid residues comprising epitopes predicted to have the highest degree of antigenicity by particular algorithm. The polypeptides not listed in Table 4 and portions of polypeptides not listed in Table 4 are not considered non-antigenic. This is because they may still be antigenic *in vivo* but merely not recognized as such by the particular algorithm used. Thus, Table 4 lists the amino acid residues comprising only preferred antigenic epitopes, not a complete list. In fact, all fragments of the polypeptide sequence of Table 1, at least 7 amino acids residues in length, are included in the present invention as being useful in epitope mapping and in making antibodies to particular portions of the polypeptides. Moreover, Table 4 lists only the critical residues of the epitopes determined by the Jameson-Wolf analysis. Thus, additional flanking residues on either the N-terminal, C-terminal, or both N-

and C-terminal ends may be added to the sequences of Table 4 to generate a epitope-bearing portion at least 7 residues in length. Amino acid residues comprising other antigenic epitopes may be determined by algorithms similar to the Jameson-Wolf analysis or by *in vivo* testing for an antigenic response using the methods described herein or those known in the art.

TABLE 4.

Residues Comprising Antigenic Epitopes

HGS010 MurC	from about Gly-137 to about Lys-139, from about Lys-236 to about Asp-239.
HGS027 Rfl	from about Asn-106 to about Lys-109, from about Glu-191 to about Gly-194; from about Arg-227 to about Ala-231.
HGS038 NusA	from about Lys-39 to about Asp-42, from about Pro-170 to about Lys-173, from about Thr-302 to about Gln-304.
HGS041 NadE	from about Lys-173 to about Asp-176, from about Lys-189 to about Gly-192, from about Lys-273 to about Arg-275.
HGS042 TrxB	from about Lys-192 to about Asp-194, from about Lys-210 to about Gly-212.
HGS043 FemD/GlmM	from about Arg-29 to about Gly-31, from about Pro-210 to about Gly-212, from about Asn-305 to about Thr-307.
HGS044 GlmU	from about Asp-261 to about Thr-263, from about Asp-390 to about Asn-393, from about Arg-452 to about Gly-454.
HGS045 CoADR	from about Thr-377 to about Asn-379.
HGS046 SVR	from about Tyr-89 to about Ser-92.
HGS050 MurF	from about Asp-258 to about Thr-262.
HGS053 Ribosomal Protein S15	from about Arg-53 to about Gly-55.
HGS057 Ribosomal Protein S9	from about Arg-7 to about Thr-9, from about Arg-11 to about Lys-13, from about Lys-58 to about Asn-60.
HGS059 Ribosomal Protein S14	from about Pro-40 to about Asp-42.
HGS060 Ribosomal Protein S19	from about Asp-53 to about Arg-55.
HGS064 YycF	from about Asp-34 to about Asn-36, from about Gly-58 to about Asp-60.
HGS063	from about Asp-27 to about Thr-31, from about Tyr-52 to about Gly-54, from about Glu-104 to about Gly-109, from about Gln-196 to about Asp-202.
HGS067	from about Pro-27 to about Asp-29, from about Pro-236 to about Lys-238.
HGS068	from about Pro-221 to about Lys-223.
HGS069	from about Pro-180 to about Asp-182.
HGS071 DdlA	from about Asn-45 to about Asp-48, from about Ser-82 to about Ser-84, from about Lys-249 to about Gly-255, from about Lys-350 to about Tyr-353.
HGS072 IspA	from about Asp-88 to about Asp-91, from about Arg-93 to about Gly-95, from about Asn-240 to about Ser-243.
HGS073 IspB	from about Lys-44 to about Gly-47.
HGS075 YycG	from about Tyr-140 to about Gly-143, from about Ser-221 to about Asn-224, from about Ser-506 to about Asp-509.
PbpI	from about Glu-64 to about Gly-66, from about Asp-70 to about Asn-72, from about Arg-140 to about Gly-142, from about Pro-172 to about Gly-174, from about Pro-234 to about Asp-238.

	from about Glu-292 to about Gly-294, from about Pro-312 to about Ser-314, from about Lys-337 to about Gly-339.
DeaD	from about Asn-380 to about Arg-382, from about Arg-462 to about Asn-466, from about Asn-474 to about Gly-480, from about Asp-485 to about Tyr-494, from about Lys-509 to about Gly-513.

These polypeptide fragments have been determined to bear antigenic epitopes of the *S. aureus* proteins shown in Table 1 by the analysis of the Jameson-Wolf antigenic index. Antibodies which specifically bind any epitope or polypeptide of the present invention may also be excluded. Therefore, the present invention includes antibodies that specifically bind polypeptides of the present invention, and allows for the exclusion of the same.

Antibodies of the present invention may also be described or specified in terms of their cross-reactivity. Antibodies that do not bind any other analog, ortholog, or homolog of a polypeptide of the present invention are included. Antibodies that bind polypeptides with at least 95%, at least 90%, at least 85%, at least 80%, at least 75%, at least 70%, at least 65%, at least 60%, at least 55%, and at least 50% identity (as calculated using methods known in the art and described herein) to a polypeptide of the present invention are also included in the present invention. In specific embodiments, antibodies of the present invention cross-react with murine, rat and/or rabbit homologs of human proteins and the corresponding epitopes thereof. Antibodies that do not bind polypeptides with less than 95%, less than 90%, less than 85%, less than 80%, less than 75%, less than 70%, less than 65%, less than 60%, less than 55%, and less than 50% identity (as calculated using methods known in the art and described herein) to a polypeptide of the present invention are also included in the present invention. In a specific embodiment, the above-described cross-reactivity is with respect to any single specific antigenic or immunogenic polypeptide, or combination(s) of 2, 3, 4, 5, or more of the specific antigenic and/or immunogenic polypeptides disclosed herein. Further included in the present invention are antibodies which bind polypeptides encoded by polynucleotides which hybridize to a polynucleotide of the present invention under stringent hybridization conditions (as described herein). Antibodies of the present invention may also be described or specified in terms of their binding affinity to a polypeptide of the invention. Preferred binding affinities include those with a dissociation constant or K_d less than 5×10^{-2} M, 10^{-2} M, 5×10^{-3} M, 10^{-3} M, 5×10^{-4} M, 10^{-4} M, 5×10^{-5} M, 10^{-5} M, 5×10^{-6} M, 10^{-6} M, 5×10^{-7} M, 10^{-7} M, 5×10^{-8} M, 10^{-8} M, 5×10^{-9} M, 10^{-9} M, 5×10^{-10} M, 10^{-10} M, 5×10^{-11} M, 10^{-11} M, 5×10^{-12} M, 10^{-12} M, 5×10^{-13} M, 10^{-13} M, 5×10^{-14} M, 10^{-14} M, 5×10^{-15} M, or 10^{-15} M.

The invention also provides antibodies that competitively inhibit binding of an antibody to an epitope of the invention as determined by any method known in the art for determining competitive binding, for example, the immunoassays described herein. In preferred embodiments, the antibody competitively inhibits binding to the epitope by at least 95%, at least 90%, at least 85 %, at least 80%, at least 75%, at least 70%, at least 60%, or at least 50%.

Antibodies of the present invention may act as agonists or antagonists of the polypeptides of the present invention. For example, the present invention includes antibodies which disrupt the receptor/ligand interactions with the polypeptides of the invention either partially or fully. Preferably, antibodies of the present invention bind an antigenic epitope disclosed herein, or a portion thereof. The invention features both receptor-specific antibodies and ligand-specific antibodies. The invention also features receptor-specific antibodies which do not prevent ligand binding but prevent receptor activation. Receptor activation (i.e., signaling) may be determined by techniques described herein or otherwise known in the art. For example, receptor activation can be determined by detecting the phosphorylation (e.g., tyrosine or serine/threonine) of the receptor or its substrate by immunoprecipitation followed by western blot analysis (for example, as described supra). In specific embodiments, antibodies are provided that inhibit ligand activity or receptor activity by at least 95%, at least 90%, at least 85%, at least 80%, at least 75%, at least 70%, at least 60%, or at least 50% of the activity in absence of the antibody.

The invention also features receptor-specific antibodies which both prevent ligand binding and receptor activation as well as antibodies that recognize the receptor-ligand complex, and, preferably, do not specifically recognize the unbound receptor or the unbound ligand. Likewise, included in the invention are neutralizing antibodies which bind the ligand and prevent binding of the ligand to the receptor, as well as antibodies which bind the ligand, thereby preventing receptor activation, but do not prevent the ligand from binding the receptor. Further included in the invention are antibodies which activate the receptor. These antibodies may act as receptor agonists, i.e., potentiate or activate either all or a subset of the biological activities of the ligand-mediated receptor activation, for example, by inducing dimerization of the receptor. The antibodies may be specified as agonists, antagonists or inverse agonists for biological activities comprising the specific biological activities of the peptides of the invention disclosed herein. The above antibody agonists can be made using

methods known in the art. See, e.g., PCT publication WO 96/40281; U.S. Patent No. 5,811,097; Deng et al., *Blood* 92(6):1981-1988 (1998); Chen et al., *Cancer Res.* 58(16):3668-3678 (1998); Harrop et al., *J. Immunol.* 161(4):1786-1794 (1998); Zhu et al., *Cancer Res.* 58(15):3209-3214 (1998); Yoon et al., *J. Immunol.* 160(7):3170-3179 (1998);
5 Prat et al., *J. Cell. Sci.* 111(Pt2):237-247 (1998); Pitard et al., *J. Immunol. Methods* 205(2):177-190 (1997); Liautard et al., *Cytokine* 9(4):233-241 (1997); Carlson et al., *J. Biol. Chem.* 272(17):11295-11301 (1997); Taryman et al., *Neuron* 14(4):755-762 (1995); Muller et al., *Structure* 6(9):1153-1167 (1998); Bartunek et al., *Cytokine* 8(1):14-20 (1996) (which are all incorporated by reference herein in their entireties).

10 Antibodies of the present invention may be used, for example, but not limited to, to purify, detect, and target the polypeptides of the present invention, including both in vitro and in vivo diagnostic and therapeutic methods. For example, the antibodies have use in immunoassays for qualitatively and quantitatively measuring levels of the polypeptides of the present invention in biological samples. See, e.g., Harlow et al., *Antibodies: A Laboratory*
15 *Manual*, (Cold Spring Harbor Laboratory Press, 2nd ed. 1988) (incorporated by reference herein in its entirety).

As discussed in more detail below, the antibodies of the present invention may be used either alone or in combination with other compositions. The antibodies may further be recombinantly fused to a heterologous polypeptide at the N- or C-terminus or chemically
20 conjugated (including covalently and non-covalently conjugations) to polypeptides or other compositions. For example, antibodies of the present invention may be recombinantly fused or conjugated to molecules useful as labels in detection assays and effector molecules such as heterologous polypeptides, drugs, radionuclides, or toxins. See, e.g., PCT publications WO 92/08495; WO 91/14438; WO 89/12624; U.S. Patent No. 5,314,995; and EP 396,387.

25 The antibodies of the invention include derivatives that are modified, i.e., by the covalent attachment of any type of molecule to the antibody such that covalent attachment does not prevent the antibody from generating an anti-idiotypic response. For example, but not by way of limitation, the antibody derivatives include antibodies that have been modified, e.g., by glycosylation, acetylation, pegylation, phosphorylation, amidation, derivatization by
30 known protecting/blocking groups, proteolytic cleavage, linkage to a cellular ligand or other protein, etc. Any of numerous chemical modifications may be carried out by known techniques, including, but not limited to specific chemical cleavage, acetylation, formylation,

metabolic synthesis of tunicamycin, etc. Additionally, the derivative may contain one or more non-classical amino acids.

The antibodies of the present invention may be generated by any suitable method known in the art. Polyclonal antibodies to an antigen-of-interest can be produced by various procedures well known in the art. For example, a polypeptide of the invention can be administered to various host animals including, but not limited to, rabbits, mice, rats, etc. to induce the production of sera containing polyclonal antibodies specific for the antigen. Various adjuvants may be used to increase the immunological response, depending on the host species, and include but are not limited to, Freund's (complete and incomplete), mineral gels such as aluminum hydroxide, surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, keyhole limpet hemocyanins, dinitrophenol, and potentially useful human adjuvants such as BCG (bacille Calmette-Guerin) and corynebacterium parvum. Such adjuvants are also well known in the art.

Monoclonal antibodies can be prepared using a wide variety of techniques known in the art including the use of hybridoma, recombinant, and phage display technologies, or a combination thereof. For example, monoclonal antibodies can be produced using hybridoma techniques including those known in the art and taught, for example, in Harlow et al., *Antibodies: A Laboratory Manual*, (Cold Spring Harbor Laboratory Press, 2nd ed. 1988); Hammerling, et al., in: *Monoclonal Antibodies and T-Cell Hybridomas* 563-681 (Elsevier, N.Y., 1981) (said references incorporated by reference in their entireties). The term "monoclonal antibody" as used herein is not limited to antibodies produced through hybridoma technology. The term "monoclonal antibody" refers to an antibody that is derived from a single clone, including any eukaryotic, prokaryotic, or phage clone, and not the method by which it is produced.

Methods for producing and screening for specific antibodies using hybridoma technology are routine and well known in the art and are discussed in detail in the Examples. In a non-limiting example, mice can be immunized with a polypeptide of the invention or a cell expressing such peptide. Once an immune response is detected, e.g., antibodies specific for the antigen are detected in the mouse serum, the mouse spleen is harvested and splenocytes isolated. The splenocytes are then fused by well known techniques to any suitable myeloma cells, for example cells from cell line SP20 available from the ATCC. Hybridomas are selected and cloned by limited dilution. The hybridoma clones are then

assayed by methods known in the art for cells that secrete antibodies capable of binding a polypeptide of the invention. Ascites fluid, which generally contains high levels of antibodies, can be generated by immunizing mice with positive hybridoma clones.

Accordingly, the present invention provides methods of generating monoclonal antibodies as well as antibodies produced by the method comprising culturing a hybridoma cell secreting an antibody of the invention wherein, preferably, the hybridoma is generated by fusing splenocytes isolated from a mouse immunized with an antigen of the invention with myeloma cells and then screening the hybridomas resulting from the fusion for hybridoma clones that secrete an antibody able to bind a polypeptide of the invention.

Antibody fragments which recognize specific epitopes may be generated by known techniques. For example, Fab and F(ab')₂ fragments of the invention may be produced by proteolytic cleavage of immunoglobulin molecules, using enzymes such as papain (to produce Fab fragments) or pepsin (to produce F(ab')₂ fragments). F(ab')₂ fragments contain the variable region, the light chain constant region and the CH1 domain of the heavy chain.

For example, the antibodies of the present invention can also be generated using various phage display methods known in the art. In phage display methods, functional antibody domains are displayed on the surface of phage particles which carry the polynucleotide sequences encoding them. In a particular embodiment, such phage can be utilized to display antigen binding domains expressed from a repertoire or combinatorial antibody library (e.g., human or murine). Phage expressing an antigen binding domain that binds the antigen of interest can be selected or identified with antigen, e.g., using labeled antigen or antigen bound or captured to a solid surface or bead. Phage used in these methods are typically filamentous phage including fd and M13 binding domains expressed from phage with Fab, Fv or disulfide stabilized Fv antibody domains recombinantly fused to either the phage gene III or gene VIII protein. Examples of phage display methods that can be used to make the antibodies of the present invention include those disclosed in Brinkman et al., J. Immunol. Methods 182:41-50 (1995); Ames et al., J. Immunol. Methods 184:177-186 (1995); Kettleborough et al., Eur. J. Immunol. 24:952-958 (1994); Persic et al., Gene 187 9-18 (1997); Burton et al., Advances in Immunology 57:191-280 (1994); PCT application No. PCT/GB91/01134; PCT publications WO 90/02809; WO 91/10737; WO 92/01047; WO 92/18619; WO 93/11236; WO 95/15982; WO 95/20401; and U.S. Patent Nos. 5,698,426;

5,223,409; 5,403,484; 5,580,717; 5,427,908; 5,750,753; 5,821,047; 5,571,698; 5,427,908; 5,516,637; 5,780,225; 5,658,727; 5,733,743 and 5,969,108; each of which is incorporated herein by reference in its entirety.

As described in the above references, after phage selection, the antibody coding regions from the phage can be isolated and used to generate whole antibodies, including human antibodies, or any other desired antigen binding fragment, and expressed in any desired host, including mammalian cells, insect cells, plant cells, yeast, and bacteria, e.g., as described in detail below. For example, techniques to recombinantly produce Fab, Fab' and F(ab')₂ fragments can also be employed using methods known in the art such as those disclosed in PCT publication WO 92/22324; Mullinax et al., *BioTechniques* 12(6):864-869 (1992); and Sawai et al., *AJRI* 34:26-34 (1995); and Better et al., *Science* 240:1041-1043 (1988) (said references incorporated by reference in their entireties).

Examples of techniques which can be used to produce single-chain Fvs and antibodies include those described in U.S. Patents 4,946,778 and 5,258,498; Huston et al., *Methods in Enzymology* 203:46-88 (1991); Shu et al., *PNAS* 90:7995-7999 (1993); and Skerra et al., *Science* 240:1038-1040 (1988). For some uses, including in vivo use of antibodies in humans and in vitro detection assays, it may be preferable to use chimeric, humanized, or human antibodies. A chimeric antibody is a molecule in which different portions of the antibody are derived from different animal species, such as antibodies having a variable region derived from a murine monoclonal antibody and a human immunoglobulin constant region. Methods for producing chimeric antibodies are known in the art. See e.g., Morrison, *Science* 229:1202 (1985); Oi et al., *BioTechniques* 4:214 (1986); Gillies et al., (1989) *J. Immunol. Methods* 125:191-202; U.S. Patent Nos. 5,807,715; 4,816,567; and 4,816,397, which are incorporated herein by reference in their entirety. Humanized antibodies are antibody molecules from non-human species antibody that binds the desired antigen having one or more complementarity determining regions (CDRs) from the non-human species and a framework regions from a human immunoglobulin molecule. Often, framework residues in the human framework regions will be substituted with the corresponding residue from the CDR donor antibody to alter, preferably improve, antigen binding. These framework substitutions are identified by methods well known in the art, e.g., by modeling of the interactions of the CDR and framework residues to identify framework residues important for antigen binding and sequence comparison to identify unusual framework residues at

particular positions. (See, e.g., Queen et al., U.S. Patent No. 5,585,089; Riechmann et al., Nature 332:323 (1988), which are incorporated herein by reference in their entireties.) Antibodies can be humanized using a variety of techniques known in the art including, for example, CDR-grafting (EP 239,400; PCT publication WO 91/09967; U.S. Patent Nos. 5,225,539; 5,530,101; and 5,585,089), veneering or resurfacing (EP 592,106; EP 519,596; Padlan, Molecular Immunology 28(4/5):489-498 (1991); Studnicka et al., Protein Engineering 7(6):805-814 (1994); Roguska et al., PNAS 91:969-973 (1994)), and chain shuffling (U.S. Patent No. 5,565,332).

Completely human antibodies are particularly desirable for therapeutic treatment of human patients. Human antibodies can be made by a variety of methods known in the art including phage display methods described above using antibody libraries derived from human immunoglobulin sequences. See also, U.S. Patent Nos. 4,444,887 and 4,716,111; and PCT publications WO 98/46645, WO 98/50433, WO 98/24893, WO 98/16654, WO 96/34096, WO 96/33735, and WO 91/10741; each of which is incorporated herein by reference in its entirety.

Human antibodies can also be produced using transgenic mice which are incapable of expressing functional endogenous immunoglobulins, but which can express human immunoglobulin genes. For example, the human heavy and light chain immunoglobulin gene complexes may be introduced randomly or by homologous recombination into mouse embryonic stem cells. Alternatively, the human variable region, constant region, and diversity region may be introduced into mouse embryonic stem cells in addition to the human heavy and light chain genes. The mouse heavy and light chain immunoglobulin genes may be rendered non-functional separately or simultaneously with the introduction of human immunoglobulin loci by homologous recombination. In particular, homozygous deletion of the JH region prevents endogenous antibody production. The modified embryonic stem cells are expanded and microinjected into blastocysts to produce chimeric mice. The chimeric mice are then bred to produce homozygous offspring which express human antibodies. The transgenic mice are immunized in the normal fashion with a selected antigen, e.g., all or a portion of a polypeptide of the invention. Monoclonal antibodies directed against the antigen can be obtained from the immunized, transgenic mice using conventional hybridoma technology. The human immunoglobulin transgenes harbored by the transgenic mice rearrange during B cell differentiation, and subsequently undergo class switching and

somatic mutation. Thus, using such a technique, it is possible to produce therapeutically useful IgG, IgA, IgM and IgE antibodies. For an overview of this technology for producing human antibodies, see Lonberg and Huszar, *Int. Rev. Immunol.* 13:65-93 (1995). For a detailed discussion of this technology for producing human antibodies and human monoclonal antibodies and protocols for producing such antibodies, see, e.g., PCT publications WO 98/24893; WO 92/01047; WO 96/34096; WO 96/33735; European Patent No. 0 598 877; U.S. Patent Nos. 5,413,923; 5,625,126; 5,633,425; 5,569,825; 5,661,016; 5,545,806; 5,814,318; 5,885,793; 5,916,771; and 5,939,598, which are incorporated by reference herein in their entirety. In addition, companies such as Abgenix, Inc. (Freemont, CA) and Genpharm (San Jose, CA) can be engaged to provide human antibodies directed against a selected antigen using technology similar to that described above.

Completely human antibodies which recognize a selected epitope can be generated using a technique referred to as "guided selection." In this approach a selected non-human monoclonal antibody, e.g., a mouse antibody, is used to guide the selection of a completely human antibody recognizing the same epitope. (Jespers et al., *Bio/technology* 12:899-903 (1988)).

Further, antibodies to the polypeptides of the invention can, in turn, be utilized to generate anti-idiotypic antibodies that "mimic" polypeptides of the invention using techniques well known to those skilled in the art. (See, e.g., Greenspan & Bona, *FASEB J.* 7(5):437-444; (1989) and Nissinoff, J. *Immunol.* 147(8):2429-2438 (1991)). For example, antibodies which bind to and competitively inhibit polypeptide multimerization and/or binding of a polypeptide of the invention to a ligand can be used to generate anti-idiotypes that "mimic" the polypeptide multimerization and/or binding domain and, as a consequence, bind to and neutralize polypeptide and/or its ligand. Such neutralizing anti-idiotypes or Fab fragments of such anti-idiotypes can be used in therapeutic regimens to neutralize polypeptide ligand. For example, such anti-idiotypic antibodies can be used to bind a polypeptide of the invention and/or to bind its ligands/receptors, and thereby block its biological activity.

Polynucleotides Encoding Antibodies

The invention further provides polynucleotides comprising a nucleotide sequence encoding an antibody of the invention and fragments thereof. The invention also encompasses polynucleotides that hybridize under stringent or lower stringency hybridization

conditions, e.g., as defined supra, to polynucleotides that encode an antibody, preferably, that specifically binds to a polypeptide of the invention, preferably, an antibody that binds to a polypeptide having any of the amino acid sequences shown in Table 1.

5 The polynucleotides may be obtained, and the nucleotide sequence of the polynucleotides determined, by any method known in the art. For example, if the nucleotide sequence of the antibody is known, a polynucleotide encoding the antibody may be assembled from chemically synthesized oligonucleotides (e.g., as described in Kutmeier et al., *BioTechniques* 17:242 (1994)), which, briefly, involves the synthesis of overlapping oligonucleotides containing portions of the sequence encoding the antibody, annealing and
10 ligating of those oligonucleotides, and then amplification of the ligated oligonucleotides by PCR.

Alternatively, a polynucleotide encoding an antibody may be generated from nucleic acid from a suitable source. If a clone containing a nucleic acid encoding a particular antibody is not available, but the sequence of the antibody molecule is known, a nucleic acid
15 encoding the immunoglobulin may be chemically synthesized or obtained from a suitable source (e.g., an antibody cDNA library, or a cDNA library generated from, or nucleic acid, preferably poly A+ RNA, isolated from, any tissue or cells expressing the antibody, such as hybridoma cells selected to express an antibody of the invention) by PCR amplification using synthetic primers hybridizable to the 3' and 5' ends of the sequence or by cloning using
20 an oligonucleotide probe specific for the particular gene sequence to identify, e.g., a cDNA clone from a cDNA library that encodes the antibody. Amplified nucleic acids generated by PCR may then be cloned into replicable cloning vectors using any method well known in the art.

Once the nucleotide sequence and corresponding amino acid sequence of the antibody
25 is determined, the nucleotide sequence of the antibody may be manipulated using methods well known in the art for the manipulation of nucleotide sequences, e.g., recombinant DNA techniques, site directed mutagenesis, PCR, etc. (see, for example, the techniques described in Sambrook et al., 1990, *Molecular Cloning, A Laboratory Manual*, 2d Ed., Cold Spring Harbor Laboratory, Cold Spring Harbor, NY and Ausubel et al., eds., 1998, *Current Protocols in Molecular Biology*, John Wiley & Sons, NY, which are both incorporated by reference
30 herein in their entireties), to generate antibodies having a different amino acid sequence, for example to create amino acid substitutions, deletions, and/or insertions.

In a specific embodiment, the amino acid sequence of the heavy and/or light chain variable domains may be inspected to identify the sequences of the complementarity determining regions (CDRs) by methods that are well known in the art, e.g., by comparison to known amino acid sequences of other heavy and light chain variable regions to determine the regions of sequence hypervariability. Using routine recombinant DNA techniques, one or more of the CDRs may be inserted within framework regions, e.g., into human framework regions to humanize a non-human antibody, as described supra. The framework regions may be naturally occurring or consensus framework regions, and preferably human framework regions (see, e.g., Chothia et al., J. Mol. Biol. 278: 457-479 (1998) for a listing of human framework regions). Preferably, the polynucleotide generated by the combination of the framework regions and CDRs encodes an antibody that specifically binds a polypeptide of the invention. Preferably, as discussed supra, one or more amino acid substitutions may be made within the framework regions, and, preferably, the amino acid substitutions improve binding of the antibody to its antigen. Additionally, such methods may be used to make amino acid substitutions or deletions of one or more variable region cysteine residues participating in an intrachain disulfide bond to generate antibody molecules lacking one or more intrachain disulfide bonds. Other alterations to the polynucleotide are encompassed by the present invention and within the skill of the art.

In addition, techniques developed for the production of "chimeric antibodies" (Morrison et al., Proc. Natl. Acad. Sci. 81:851-855 (1984); Neuberger et al., Nature 312:604-608 (1984); Takeda et al., Nature 314:452-454 (1985)) by splicing genes from a mouse antibody molecule of appropriate antigen specificity together with genes from a human antibody molecule of appropriate biological activity can be used. As described supra, a chimeric antibody is a molecule in which different portions are derived from different animal species, such as those having a variable region derived from a murine mAb and a human immunoglobulin constant region, e.g., humanized antibodies.

Alternatively, techniques described for the production of single chain antibodies (U.S. Patent No. 4,946,778; Bird, Science 242:423-42 (1988); Huston et al., Proc. Natl. Acad. Sci. USA 85:5879-5883 (1988); and Ward et al., Nature 334:544-54 (1989)) can be adapted to produce single chain antibodies. Single chain antibodies are formed by linking the heavy and light chain fragments of the Fv region via an amino acid bridge, resulting in a single

chain polypeptide. Techniques for the assembly of functional Fv fragments in *E. coli* may also be used (Skerra et al., Science 242:1038- 1041 (1988)).

Methods of Producing Antibodies

5 The antibodies of the invention can be produced by any method known in the art for the synthesis of antibodies, in particular, by chemical synthesis or preferably, by recombinant expression techniques.

Recombinant expression of an antibody of the invention, or fragment, derivative or analog thereof, (e.g., a heavy or light chain of an antibody of the invention or a single chain
10 antibody of the invention), requires construction of an expression vector containing a polynucleotide that encodes the antibody. Once a polynucleotide encoding an antibody molecule or a heavy or light chain of an antibody, or portion thereof (preferably containing the heavy or light chain variable domain), of the invention has been obtained, the vector for the production of the antibody molecule may be produced by recombinant DNA technology
15 using techniques well known in the art. Thus, methods for preparing a protein by expressing a polynucleotide containing an antibody encoding nucleotide sequence are described herein. Methods which are well known to those skilled in the art can be used to construct expression vectors containing antibody coding sequences and appropriate transcriptional and translational control signals. These methods include, for example, in vitro recombinant DNA
20 techniques, synthetic techniques, and in vivo genetic recombination. The invention, thus, provides replicable vectors comprising a nucleotide sequence encoding an antibody molecule of the invention, or a heavy or light chain thereof, or a heavy or light chain variable domain, operably linked to a promoter. Such vectors may include the nucleotide sequence encoding the constant region of the antibody molecule (see, e.g., PCT Publication WO 86/05807; PCT
25 Publication WO 89/01036; and U.S. Patent No. 5,122,464) and the variable domain of the antibody may be cloned into such a vector for expression of the entire heavy or light chain.

The expression vector is transferred to a host cell by conventional techniques and the transfected cells are then cultured by conventional techniques to produce an antibody of the invention. Thus, the invention includes host cells containing a polynucleotide encoding an
30 antibody of the invention, or a heavy or light chain thereof, or a single chain antibody of the invention, operably linked to a heterologous promoter. In preferred embodiments for the expression of double-chained antibodies, vectors encoding both the heavy and light chains

may be co-expressed in the host cell for expression of the entire immunoglobulin molecule, as detailed below.

A variety of host-expression vector systems may be utilized to express the antibody molecules of the invention. Such host-expression systems represent vehicles by which the coding sequences of interest may be produced and subsequently purified, but also represent cells which may, when transformed or transfected with the appropriate nucleotide coding sequences, express an antibody molecule of the invention in situ. These include but are not limited to microorganisms such as bacteria (e.g., *E. coli*, *B. subtilis*) transformed with recombinant bacteriophage DNA, plasmid DNA or cosmid DNA expression vectors containing antibody coding sequences; yeast (e.g., *Saccharomyces*, *Pichia*) transformed with recombinant yeast expression vectors containing antibody coding sequences; insect cell systems infected with recombinant virus expression vectors (e.g., baculovirus) containing antibody coding sequences; plant cell systems infected with recombinant virus expression vectors (e.g., cauliflower mosaic virus, CaMV; tobacco mosaic virus, TMV) or transformed with recombinant plasmid expression vectors (e.g., Ti plasmid) containing antibody coding sequences; or mammalian cell systems (e.g., COS, CHO, BHK, 293, 3T3 cells) harboring recombinant expression constructs containing promoters derived from the genome of mammalian cells (e.g., metallothionein promoter) or from mammalian viruses (e.g., the adenovirus late promoter; the vaccinia virus 7.5K promoter). Preferably, bacterial cells such as *Escherichia coli*, and more preferably, eukaryotic cells, especially for the expression of whole recombinant antibody molecule, are used for the expression of a recombinant antibody molecule. For example, mammalian cells such as Chinese hamster ovary cells (CHO), in conjunction with a vector such as the major intermediate early gene promoter element from human cytomegalovirus is an effective expression system for antibodies (Foecking et al., Gene 45:101 (1986); Cockett et al., Bio/Technology 8:2 (1990)).

In bacterial systems, a number of expression vectors may be advantageously selected depending upon the use intended for the antibody molecule being expressed. For example, when a large quantity of such a protein is to be produced, for the generation of pharmaceutical compositions of an antibody molecule, vectors which direct the expression of high levels of fusion protein products that are readily purified may be desirable. Such vectors include, but are not limited, to the *E. coli* expression vector pUR278 (Ruther et al., EMBO J. 2:1791 (1983)), in which the antibody coding sequence may be ligated individually into the

vector in frame with the lac Z coding region so that a fusion protein is produced; pIN vectors (Inouye & Inouye, Nucleic Acids Res. 13:3101-3109 (1985); Van Heeke & Schuster, J. Biol. Chem. 24:5503-5509 (1989)); and the like. pGEX vectors may also be used to express foreign polypeptides as fusion proteins with glutathione S-transferase (GST). In general, such fusion proteins are soluble and can easily be purified from lysed cells by adsorption and binding to matrix glutathione-agarose beads followed by elution in the presence of free glutathione. The pGEX vectors are designed to include thrombin or factor Xa protease cleavage sites so that the cloned target gene product can be released from the GST moiety.

In an insect system, Autographa californica nuclear polyhedrosis virus (AcNPV) is used as a vector to express foreign genes. The virus grows in *Spodoptera frugiperda* cells. The antibody coding sequence may be cloned individually into non-essential regions (for example the polyhedrin gene) of the virus and placed under control of an AcNPV promoter (for example the polyhedrin promoter).

In mammalian host cells, a number of viral-based expression systems may be utilized. In cases where an adenovirus is used as an expression vector, the antibody coding sequence of interest may be ligated to an adenovirus transcription/translation control complex, e.g., the late promoter and tripartite leader sequence. This chimeric gene may then be inserted in the adenovirus genome by in vitro or in vivo recombination. Insertion in a non-essential region of the viral genome (e.g., region E1 or E3) will result in a recombinant virus that is viable and capable of expressing the antibody molecule in infected hosts. (e.g., see Logan & Shenk, Proc. Natl. Acad. Sci. USA 81:355-359 (1984)). Specific initiation signals may also be required for efficient translation of inserted antibody coding sequences. These signals include the ATG initiation codon and adjacent sequences. Furthermore, the initiation codon must be in phase with the reading frame of the desired coding sequence to ensure translation of the entire insert. These exogenous translational control signals and initiation codons can be of a variety of origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of appropriate transcription enhancer elements, transcription terminators, etc. (see Bittner et al., Methods in Enzymol. 153:51-544 (1987)).

In addition, a host cell strain may be chosen which modulates the expression of the inserted sequences, or modifies and processes the gene product in the specific fashion desired. Such modifications (e.g., glycosylation) and processing (e.g., cleavage) of protein products may be important for the function of the protein. Different host cells have

characteristic and specific mechanisms for the post-translational processing and modification of proteins and gene products. Appropriate cell lines or host systems can be chosen to ensure the correct modification and processing of the foreign protein expressed. To this end, eukaryotic host cells which possess the cellular machinery for proper processing of the primary transcript, glycosylation, and phosphorylation of the gene product may be used. Such mammalian host cells include but are not limited to CHO, VERY, BHK, Hela, COS, MDCK, 293, 3T3, WI38, and in particular, breast cancer cell lines such as, for example, BT483, Hs578T, HTB2, BT20 and T47D, and normal mammary gland cell line such as, for example, CRL7030 and Hs578Bst.

For long-term, high-yield production of recombinant proteins, stable expression is preferred. For example, cell lines which stably express the antibody molecule may be engineered. Rather than using expression vectors which contain viral origins of replication, host cells can be transformed with DNA controlled by appropriate expression control elements (e.g., promoter, enhancer, sequences, transcription terminators, polyadenylation sites, etc.), and a selectable marker. Following the introduction of the foreign DNA, engineered cells may be allowed to grow for 1-2 days in an enriched media, and then are switched to a selective media. The selectable marker in the recombinant plasmid confers resistance to the selection and allows cells to stably integrate the plasmid into their chromosomes and grow to form foci which in turn can be cloned and expanded into cell lines. This method may advantageously be used to engineer cell lines which express the antibody molecule. Such engineered cell lines may be particularly useful in screening and evaluation of compounds that interact directly or indirectly with the antibody molecule.

A number of selection systems may be used, including but not limited to the herpes simplex virus thymidine kinase (Wigler et al., Cell 11:223 (1977)), hypoxanthine-guanine phosphoribosyltransferase (Szybalska & Szybalski, Proc. Natl. Acad. Sci. USA 48:202 (1992)), and adenine phosphoribosyltransferase (Lowy et al., Cell 22:817 (1980)) genes can be employed in tk-, hgp^rt- or ap^rt- cells, respectively. Also, antimetabolite resistance can be used as the basis of selection for the following genes: dhfr, which confers resistance to methotrexate (Wigler et al., Natl. Acad. Sci. USA 77:357 (1980); O'Hare et al., Proc. Natl. Acad. Sci. USA 78:1527 (1981)); gpt, which confers resistance to mycophenolic acid (Mulligan & Berg, Proc. Natl. Acad. Sci. USA 78:2072 (1981)); neo, which confers resistance to the aminoglycoside G-418 Clinical Pharmacy 12:488-505; Wu and Wu,

Biotherapy 3:87-95 (1991); Tolstoshev, *Ann. Rev. Pharmacol. Toxicol.* 32:573-596 (1993); Mulligan, *Science* 260:926-932 (1993); and Morgan and Anderson, *Ann. Rev. Biochem.* 62:191-217 (1993); May, 1993, *TIB TECH* 11(5):155-215; and hygromycin, which confers resistance to hygromycin (Santerre et al., *Gene* 30:147 (1984)). Methods commonly known in the art of recombinant DNA technology may be routinely applied to select the desired recombinant clone, and such methods are described, for example, in Ausubel et al. (eds.), *Current Protocols in Molecular Biology*, John Wiley & Sons, NY (1993); Kriegler, *Gene Transfer and Expression, A Laboratory Manual*, Stockton Press, NY (1990); and in Chapters 12 and 13, Dracopoli et al. (eds), *Current Protocols in Human Genetics*, John Wiley & Sons, NY (1994); Colberre-Garapin et al., *J. Mol. Biol.* 150:1 (1981), which are incorporated by reference herein in their entireties.

The expression levels of an antibody molecule can be increased by vector amplification (for a review, see Bebbington and Hentschel, *The use of vectors based on gene amplification for the expression of cloned genes in mammalian cells in DNA cloning*, Vol.3. (Academic Press, New York, 1987)). When a marker in the vector system expressing antibody is amplifiable, increase in the level of inhibitor present in culture of host cell will increase the number of copies of the marker gene. Since the amplified region is associated with the antibody gene, production of the antibody will also increase (Crouse et al., *Mol. Cell. Biol.* 3:257 (1983)).

The host cell may be co-transfected with two expression vectors of the invention, the first vector encoding a heavy chain derived polypeptide and the second vector encoding a light chain derived polypeptide. The two vectors may contain identical selectable markers which enable equal expression of heavy and light chain polypeptides. Alternatively, a single vector may be used which encodes, and is capable of expressing, both heavy and light chain polypeptides. In such situations, the light chain should be placed before the heavy chain to avoid an excess of toxic free heavy chain (Proudfoot, *Nature* 322:52 (1986); Kohler, *Proc. Natl. Acad. Sci. USA* 77:2197 (1980)). The coding sequences for the heavy and light chains may comprise cDNA or genomic DNA.

Once an antibody molecule of the invention has been produced by an animal, chemically synthesized, or recombinantly expressed, it may be purified by any method known in the art for purification of an immunoglobulin molecule, for example, by chromatography (e.g., ion exchange, affinity, particularly by affinity for the specific antigen

after Protein A, and sizing column chromatography), centrifugation, differential solubility, or by any other standard technique for the purification of proteins. In addition, the antibodies of the present invention or fragments thereof can be fused to heterologous polypeptide sequences described herein or otherwise known in the art, to facilitate purification.

5 The present invention encompasses antibodies recombinantly fused or chemically conjugated (including both covalently and non-covalently conjugations) to a polypeptide (or portion thereof, preferably at least 10, 20, 30, 40, 50, 60, 70, 80, 90 or 100 amino acids of the polypeptide) of the present invention to generate fusion proteins. The fusion does not necessarily need to be direct, but may occur through linker sequences. The antibodies may
10 be specific for antigens other than polypeptides (or portion thereof, preferably at least 10, 20, 30, 40, 50, 60, 70, 80, 90 or 100 amino acids of the polypeptide) of the present invention. For example, antibodies may be used to target the polypeptides of the present invention to particular cell types, either in vitro or in vivo, by fusing or conjugating the polypeptides of the present invention to antibodies specific for particular cell surface receptors. Antibodies
15 fused or conjugated to the polypeptides of the present invention may also be used in in vitro immunoassays and purification methods using methods known in the art. See e.g., Harbor et al., supra, and PCT publication WO 93/21232; EP 439,095; Naramura et al., Immunol. Lett. 39:91-99 (1994); U.S. Patent 5,474,981; Gillies et al., PNAS 89:1428-1432 (1992); Fell et al., J. Immunol. 146:2446-2452(1991), which are incorporated by reference in their entireties.

20 The present invention further includes compositions comprising the polypeptides of the present invention fused or conjugated to antibody domains other than the variable regions. For example, the polypeptides of the present invention may be fused or conjugated to an antibody Fc region, or portion thereof. The antibody portion fused to a polypeptide of the present invention may comprise the constant region, hinge region, CH1 domain, CH2
25 domain, and CH3 domain or any combination of whole domains or portions thereof. The polypeptides may also be fused or conjugated to the above antibody portions to form multimers. For example, Fc portions fused to the polypeptides of the present invention can form dimers through disulfide bonding between the Fc portions. Higher multimeric forms can be made by fusing the polypeptides to portions of IgA and IgM. Methods for fusing or
30 conjugating the polypeptides of the present invention to antibody portions are known in the art. See, e.g., U.S. Patent Nos. 5,336,603; 5,622,929; 5,359,046; 5,349,053; 5,447,851; 5,112,946; EP 307,434; EP 367,166; PCT publications WO 96/04388; WO 91/06570;

Ashkenazi et al., Proc. Natl. Acad. Sci. USA 88:10535-10539 (1991); Zheng et al., J. Immunol. 154:5590-5600 (1995); and Vil et al., Proc. Natl. Acad. Sci. USA 89:11337-11341(1992)-(said references incorporated by reference in their entireties).

As discussed, supra, the polypeptides corresponding to a polypeptide, polypeptide
5 fragment, or a variant of any one of the amino acid sequences shown in Table 1 may be fused
or conjugated to the above antibody portions to increase the in vivo half life of the
polypeptides or for use in immunoassays using methods known in the art. Further, the
polypeptides corresponding to *S. aureus* proteins shown in Table 1 may be fused or
conjugated to the above antibody portions to facilitate purification. One reported example
10 describes chimeric proteins consisting of the first two domains of the human CD4-
polypeptide and various domains of the constant regions of the heavy or light chains of
mammalian immunoglobulins. (EP 394,827; Traunecker et al., Nature 331:84-86 (1988).
The polypeptides of the present invention fused or conjugated to an antibody having
disulfide-linked dimeric structures (due to the IgG) may also be more efficient in binding
15 and neutralizing other molecules, than the monomeric secreted protein or protein fragment
alone. (Fountoulakis et al., J. Biochem. 270:3958-3964 (1995)). In many cases, the Fc part
in a fusion protein is beneficial in therapy and diagnosis, and thus can result in, for example,
improved pharmacokinetic properties. (EP A 232,262). Alternatively, deleting the Fc part
after the fusion protein has been expressed, detected, and purified, would be desired. For
20 example, the Fc portion may hinder therapy and diagnosis if the fusion protein is used as an
antigen for immunizations. In drug discovery, for example, human proteins, such as hIL-5,
have been fused with Fc portions for the purpose of high-throughput screening assays to
identify antagonists of hIL-5. (See, Bennett et al., J. Molecular Recognition 8:52-58 (1995);
Johanson et al., J. Biol. Chem. 270:9459-9471 (1995).

25 Moreover, the antibodies or fragments thereof of the present invention can be fused to
marker sequences, such as a peptide to facilitate purification. In preferred embodiments, the
marker amino acid sequence is a hexa-histidine peptide, such as the tag provided in a pQE
vector (QIAGEN, Inc., 9259 Eton Avenue, Chatsworth, CA, 91311), among others, many of
which are commercially available. As described in Gentz et al., Proc. Natl. Acad. Sci. USA
30 86:821-824 (1989), for instance, hexa-histidine provides for convenient purification of the
fusion protein. Other peptide tags useful for purification include, but are not limited to, the

"HA" tag, which corresponds to an epitope derived from the influenza hemagglutinin protein (Wilson et al., Cell 37:767 (1984)) and the "flag" tag.

The present invention further encompasses antibodies or fragments thereof conjugated to a diagnostic or therapeutic agent. The antibodies can be used diagnostically to, for example, monitor the development or progression of a tumor as part of a clinical testing procedure to, e.g., determine the efficacy of a given treatment regimen. Detection can be facilitated by coupling the antibody to a detectable substance. Examples of detectable substances include various enzymes, prosthetic groups, fluorescent materials, luminescent materials, bioluminescent materials, radioactive materials, positron emitting metals using various positron emission tomographies, and nonradioactive paramagnetic metal ions. The detectable substance may be coupled or conjugated either directly to the antibody (or fragment thereof) or indirectly, through an intermediate (such as, for example, a linker known in the art) using techniques known in the art. See, for example, U.S. Patent No. 4,741,900 for metal ions which can be conjugated to antibodies for use as diagnostics according to the present invention. Examples of suitable enzymes include horseradish peroxidase, alkaline phosphatase, beta-galactosidase, or acetylcholinesterase; examples of suitable prosthetic group complexes include streptavidin/biotin and avidin/biotin; examples of suitable fluorescent materials include umbelliferone, fluorescein, fluorescein isothiocyanate, rhodamine, dichlorotriazinylamine fluorescein, dansyl chloride or phycoerythrin; an example of a luminescent material includes luminol; examples of bioluminescent materials include luciferase, luciferin, and aequorin; and examples of suitable radioactive material include ^{125}I , ^{131}I , ^{111}In or ^{99}Tc .

Further, an antibody or fragment thereof may be conjugated to a therapeutic moiety such as a cytotoxin, e.g., a cytostatic or cytocidal agent, a therapeutic agent or a radioactive metal ion, e.g., alpha-emitters such as, for example, ^{213}Bi . A cytotoxin or cytotoxic agent includes any agent that is detrimental to cells. Examples include paclitaxol, cytochalasin B, gramicidin D, ethidium bromide, emetine, mitomycin, etoposide, tenoposide, vincristine, vinblastine, colchicin, doxorubicin, daunorubicin, dihydroxy anthracin dione, mitoxantrone, mithramycin, actinomycin D, 1-dehydrotestosterone, glucocorticoids, procaine, tetracaine, lidocaine, propranolol, and puromycin and analogs or homologs thereof. Therapeutic agents include, but are not limited to, antimetabolites (e.g., methotrexate, 6-mercaptopurine, 6-thioguanine, cytarabine, 5-fluorouracil decarbazine), alkylating agents (e.g.,

mechlorethamine, thioepa chlorambucil, melphalan, carmustine (BSNU) and lomustine (CCNU), cyclophosphamide, busulfan, dibromomannitol, streptozotocin, mitomycin C, and cis-dichlorodiamine platinum (II) (DDP) cisplatin, anthracyclines (e.g., daunorubicin (formerly daunomycin) and doxorubicin), antibiotics (e.g., dactinomycin (formerly actinomycin), bleomycin, mithramycin, and anthramycin (AMC)), and anti-mitotic agents (e.g., vincristine and vinblastine).

The conjugates of the invention can be used for modifying a given biological response, the therapeutic agent or drug moiety is not to be construed as limited to classical chemical therapeutic agents. For example, the drug moiety may be a protein or polypeptide possessing a desired biological activity. Such proteins may include, for example, a toxin such as abrin, ricin A, pseudomonas exotoxin, or diphtheria toxin; a protein such as tumor necrosis factor, α -interferon, β -interferon, nerve growth factor, platelet derived growth factor, tissue plasminogen activator, an apoptotic agent, e.g., TNF- α , TNF- β , AIM I (See, International Publication No. WO 97/33899), AIM II (See, International Publication No. WO 97/34911), Fas-Ligand (Takahashi *et al.*, *Int. Immunol.*, 6:1567-1574 (1994)), VEGI (See, International Publication No. WO 99/23105), a thrombotic agent or an anti-angiogenic agent, e.g., angiostatin or endostatin; or, biological response modifiers such as, for example, lymphokines, interleukin-1 ("IL-1"), interleukin-2 ("IL-2"), interleukin-6 ("IL-6"), granulocyte macrophage colony stimulating factor ("GM-CSF"), granulocyte colony stimulating factor ("G-CSF"), or other growth factors.

Antibodies may also be attached to solid supports, which are particularly useful for immunoassays or purification of the target antigen. Such solid supports include, but are not limited to, glass, cellulose, polyacrylamide, nylon, polystyrene, polyvinyl chloride or polypropylene.

Techniques for conjugating such therapeutic moiety to antibodies are well known, see, e.g., Arnon *et al.*, "Monoclonal Antibodies For Immunotargeting Of Drugs In Cancer Therapy", in *Monoclonal Antibodies And Cancer Therapy*, Reisfeld *et al.* (eds.), pp. 243-56 (Alan R. Liss, Inc. 1985); Hellstrom *et al.*, "Antibodies For Drug Delivery", in *Controlled Drug Delivery* (2nd Ed.), Robinson *et al.* (eds.), pp. 623-53 (Marcel Dekker, Inc. 1987); Thorpe, "Antibody Carriers Of Cytotoxic Agents In Cancer Therapy: A Review", in *Monoclonal Antibodies '84: Biological And Clinical Applications*, Pinchera *et al.* (eds.), pp. 475-506 (1985); "Analysis, Results, And Future Prospective Of The Therapeutic Use Of

Radiolabeled Antibody In Cancer Therapy", in *Monoclonal Antibodies For Cancer Detection And Therapy*, Baldwin et al. (eds.), pp. 303-16 (Academic Press 1985), and Thorpe et al., "The Preparation And Cytotoxic Properties Of Antibody-Toxin Conjugates", *Immunol. Rev.* 62:119-58 (1982).

5 Alternatively, an antibody can be conjugated to a second antibody to form an antibody heteroconjugate as described by Segal in U.S. Patent No. 4,676,980, which is incorporated herein by reference in its entirety.

An antibody, with or without a therapeutic moiety conjugated to it, administered alone or in combination with cytotoxic factor(s) and/or cytokine(s) can be used as a therapeutic.

10 *Immunophenotyping*

The antibodies of the invention may be utilized for immunophenotyping of cell lines and biological samples. The translation product of the gene of the present invention may be useful as a cell specific marker, or more specifically as a cellular marker that is differentially
15 expressed at various stages of differentiation and/or maturation of particular cell types. Monoclonal antibodies directed against a specific epitope, or combination of epitopes, will allow for the screening of cellular populations expressing the marker. Various techniques can be utilized using monoclonal antibodies to screen for cellular populations expressing the marker(s), and include magnetic separation using antibody-coated magnetic beads, "panning"
20 with antibody attached to a solid matrix (i.e., plate), and flow cytometry (See, e.g., U.S. Patent 5,985,660; and Morrison *et al.*, *Cell*, 96:737-49 (1999)).

These techniques allow for the screening of particular populations of cells, such as might be found with hematological malignancies (i.e. minimal residual disease (MRD) in acute leukemic patients) and "non-self" cells in transplantations to prevent Graft-versus-Host
25 Disease (GVHD). Alternatively, these techniques allow for the screening of hematopoietic stem and progenitor cells capable of undergoing proliferation and/or differentiation; as might be found in human umbilical cord blood.

Assays For Antibody Binding

30 The antibodies of the invention may be assayed for immunospecific binding by any method known in the art. The immunoassays which can be used include but are not limited to competitive and non-competitive assay systems using techniques such as western blots,

radioimmunoassays, ELISA (enzyme linked immunosorbent assay), "sandwich" immunoassays, immunoprecipitation assays, precipitin reactions, gel diffusion precipitin reactions, immunodiffusion assays, agglutination assays, complement-fixation assays, immunoradiometric assays, fluorescent immunoassays, protein A immunoassays, to name but a few. Such assays are routine and well known in the art (see, e.g., Ausubel et al, eds, 1994, Current Protocols in Molecular Biology, Vol. 1, John Wiley & Sons, Inc., New York, which is incorporated by reference herein in its entirety). Exemplary immunoassays are described briefly below (but are not intended by way of limitation).

Immunoprecipitation protocols generally comprise lysing a population of cells in a lysis buffer such as RIPA buffer (1% NP-40 or Triton X- 100, 1% sodium deoxycholate, 0.1% SDS, 0.15 M NaCl, 0.01 M sodium phosphate at pH 7.2, 1% Trasylol) supplemented with protein phosphatase and/or protease inhibitors (e.g., EDTA, PMSF, aprotinin, sodium vanadate), adding the antibody of interest to the cell lysate, incubating for a period of time (e.g., 1-4 hours) at 4° C, adding protein A and/or protein G sepharose beads to the cell lysate, incubating for about an hour or more at 4° C, washing the beads in lysis buffer and resuspending the beads in SDS/sample buffer. The ability of the antibody of interest to immunoprecipitate a particular antigen can be assessed by, e.g., western blot analysis. One of skill in the art would be knowledgeable as to the parameters that can be modified to increase the binding of the antibody to an antigen and decrease the background (e.g., pre-clearing the cell lysate with sepharose beads). For further discussion regarding immunoprecipitation protocols see, e.g., Ausubel et al, eds, 1994, Current Protocols in Molecular Biology, Vol. 1, John Wiley & Sons, Inc., New York at 10.16.1.

Western blot analysis generally comprises preparing protein samples, electrophoresis of the protein samples in a polyacrylamide gel (e.g., 8%- 20% SDS-PAGE depending on the molecular weight of the antigen), transferring the protein sample from the polyacrylamide gel to a membrane such as nitrocellulose, PVDF or nylon, blocking the membrane in blocking solution (e.g., PBS with 3% BSA or non-fat milk), washing the membrane in washing buffer (e.g., PBS-Tween 20), blocking the membrane with primary antibody (the antibody of interest) diluted in blocking buffer, washing the membrane in washing buffer, blocking the membrane with a secondary antibody (which recognizes the primary antibody, e.g., an anti-human antibody) conjugated to an enzymatic substrate (e.g., horseradish peroxidase or alkaline phosphatase) or radioactive molecule (e.g., 32P or 125I) diluted in blocking buffer,

washing the membrane in wash buffer, and detecting the presence of the antigen. One of skill in the art would be knowledgeable as to the parameters that can be modified to increase the signal detected and to reduce the background noise. For further discussion regarding western blot protocols see, e.g., Ausubel et al, eds, 1994, Current Protocols in Molecular Biology, Vol. 1, John Wiley & Sons, Inc., New York at 10.8.1.

ELISAs comprise preparing antigen, coating the well of a 96 well microtiter plate with the antigen, adding the antibody of interest conjugated to a detectable compound such as an enzymatic substrate (e.g., horseradish peroxidase or alkaline phosphatase) to the well and incubating for a period of time, and detecting the presence of the antigen. In ELISAs the antibody of interest does not have to be conjugated to a detectable compound; instead, a second antibody (which recognizes the antibody of interest) conjugated to a detectable compound may be added to the well. Further, instead of coating the well with the antigen, the antibody may be coated to the well. In this case, a second antibody conjugated to a detectable compound may be added following the addition of the antigen of interest to the coated well. One of skill in the art would be knowledgeable as to the parameters that can be modified to increase the signal detected as well as other variations of ELISAs known in the art. For further discussion regarding ELISAs see, e.g., Ausubel et al, eds, 1994, Current Protocols in Molecular Biology, Vol. 1, John Wiley & Sons, Inc., New York at 11.2.1.

The binding affinity of an antibody to an antigen and the off-rate of an antibody-antigen interaction can be determined by competitive binding assays. One example of a competitive binding assay is a radioimmunoassay comprising the incubation of labeled antigen (e.g., ^3H or ^{125}I) with the antibody of interest in the presence of increasing amounts of unlabeled antigen, and the detection of the antibody bound to the labeled antigen. The affinity of the antibody of interest for a particular antigen and the binding off-rates can be determined from the data by scatchard plot analysis. Competition with a second antibody can also be determined using radioimmunoassays. In this case, the antigen is incubated with antibody of interest conjugated to a labeled compound (e.g., ^3H or ^{125}I) in the presence of increasing amounts of an unlabeled second antibody.

Therapeutic Uses

The present invention is further directed to antibody-based therapies which involve administering antibodies of the invention to an animal, preferably a mammal, and most

preferably a human, patient for treating one or more of the disclosed diseases, disorders, or conditions. Therapeutic compounds of the invention include, but are not limited to, antibodies of the invention (including fragments, analogs and derivatives thereof as described herein) and nucleic acids encoding antibodies of the invention (including fragments, analogs and derivatives thereof and anti-idiotypic antibodies as described herein). The antibodies of the invention can be used to treat, inhibit or prevent diseases, disorders or conditions associated with aberrant expression and/or activity of a polypeptide of the invention, including, but not limited to, any one or more of the diseases, disorders, or conditions described herein. The treatment and/or prevention of diseases, disorders, or conditions associated with aberrant expression and/or activity of a polypeptide of the invention includes, but is not limited to, alleviating symptoms associated with those diseases, disorders or conditions. Antibodies of the invention may be provided in pharmaceutically acceptable compositions as known in the art or as described herein.

A summary of the ways in which the antibodies of the present invention may be used therapeutically includes binding polynucleotides or polypeptides of the present invention locally or systemically in the body or by direct cytotoxicity of the antibody, e.g. as mediated by complement (CDC) or by effector cells (ADCC). Some of these approaches are described in more detail below. Armed with the teachings provided herein, one of ordinary skill in the art will know how to use the antibodies of the present invention for diagnostic, monitoring or therapeutic purposes without undue experimentation.

The antibodies of this invention may be advantageously utilized in combination with other monoclonal or chimeric antibodies, or with lymphokines or hematopoietic growth factors (such as, e.g., IL-2, IL-3 and IL-7), for example, which serve to increase the number or activity of effector cells which interact with the antibodies.

The antibodies of the invention may be administered alone or in combination with other types of treatments (e.g., radiation therapy, chemotherapy, hormonal therapy, immunotherapy and anti-tumor agents). Generally, administration of products of a species origin or species reactivity (in the case of antibodies) that is the same species as that of the patient is preferred. Thus, in a preferred embodiment, human antibodies, fragments derivatives, analogs, or nucleic acids, are administered to a human patient for therapy or prophylaxis.

It is preferred to use high affinity and/or potent in vivo inhibiting and/or neutralizing antibodies against polypeptides or polynucleotides of the present invention, fragments or regions thereof, for both immunoassays directed to and therapy of disorders related to polynucleotides or polypeptides, including fragments thereof, of the present invention. Such antibodies, fragments, or regions, will preferably have an affinity for polynucleotides or polypeptides of the invention, including fragments thereof. Preferred binding affinities include those with a dissociation constant or K_d less than 5×10^{-2} M, 10^{-2} M, 5×10^{-3} M, 10^{-3} M, 5×10^{-4} M, 10^{-4} M, 5×10^{-5} M, 10^{-5} M, 5×10^{-6} M, 10^{-6} M, 5×10^{-7} M, 10^{-7} M, 5×10^{-8} M, 10^{-8} M, 5×10^{-9} M, 10^{-9} M, 5×10^{-10} M, 10^{-10} M, 5×10^{-11} M, 10^{-11} M, 5×10^{-12} M, 10^{-12} M, 5×10^{-13} M, 10^{-13} M, 5×10^{-14} M, 10^{-14} M, 5×10^{-15} M, and 10^{-15} M.

Gene Therapy

In a specific embodiment, nucleic acids comprising sequences encoding antibodies or functional derivatives thereof, are administered to treat, inhibit or prevent a disease or disorder associated with aberrant expression and/or activity of a polypeptide of the invention, by way of gene therapy. Gene therapy refers to therapy performed by the administration to a subject of an expressed or expressible nucleic acid. In this embodiment of the invention, the nucleic acids produce their encoded protein that mediates a therapeutic effect.

Any of the methods for gene therapy available in the art can be used according to the present invention. Exemplary methods are described below.

For general reviews of the methods of gene therapy, see Goldspiel et al., *Clinical Pharmacy* 12:488-505 (1993); Wu and Wu, *Biotherapy* 3:87-95 (1991); Tolstoshev, *Ann. Rev. Pharmacol. Toxicol.* 32:573-596 (1993); Mulligan, *Science* 260:926-932 (1993); and Morgan and Anderson, *Ann. Rev. Biochem.* 62:191-217 (1993); May, *TIBTECH* 11(5):155-215 (1993). Methods commonly known in the art of recombinant DNA technology which can be used are described in Ausubel et al. (eds.), *Current Protocols in Molecular Biology*, John Wiley & Sons, NY (1993); and Kriegler, *Gene Transfer and Expression, A Laboratory Manual*, Stockton Press, NY (1990).

In a preferred aspect, the compound comprises nucleic acid sequences encoding an antibody, said nucleic acid sequences being part of expression vectors that express the antibody or fragments or chimeric proteins or heavy or light chains thereof in a suitable host. In particular, such nucleic acid sequences have promoters operably linked to the antibody

coding region, said promoter being inducible or constitutive, and, optionally, tissue- specific. In another particular embodiment, nucleic acid molecules are used in which the antibody coding sequences and any other desired sequences are flanked by regions that promote homologous recombination at a desired site in the genome, thus providing for intrachromosomal expression of the antibody encoding nucleic acids (Koller and Smithies, 5 Proc. Natl. Acad. Sci. USA 86:8932-8935 (1989); Zijlstra et al., Nature 342:435-438 (1989). In specific embodiments, the expressed antibody molecule is a single chain antibody; alternatively, the nucleic acid sequences include sequences encoding both the heavy and light chains, or fragments thereof, of the antibody.

10 Delivery of the nucleic acids into a patient may be either direct, in which case the patient is directly exposed to the nucleic acid or nucleic acid- carrying vectors, or indirect, in which case, cells are first transformed with the nucleic acids in vitro, then transplanted into the patient. These two approaches are known, respectively, as in vivo or ex vivo gene therapy.

15 In a specific embodiment, the nucleic acid sequences are directly administered in vivo, where it is expressed to produce the encoded product. This can be accomplished by any of numerous methods known in the art, e.g., by constructing them as part of an appropriate nucleic acid expression vector and administering it so that they become intracellular, e.g., by infection using defective or attenuated retrovirals or other viral vectors 20 (see U.S. Patent No. 4,980,286), or by direct injection of naked DNA, or by use of microparticle bombardment (e.g., a gene gun; Biolistic, Dupont), or coating with lipids or cell-surface receptors or transfecting agents, encapsulation in liposomes, microparticles, or microcapsules, or by administering them in linkage to a peptide which is known to enter the nucleus, by administering it in linkage to a ligand subject to receptor-mediated endocytosis 25 (see, e.g., Wu and Wu, J. Biol. Chem. 262:4429-4432 (1987)) (which can be used to target cell types specifically expressing the receptors), etc. In another embodiment, nucleic acid-ligand complexes can be formed in which the ligand comprises a fusogenic viral peptide to disrupt endosomes, allowing the nucleic acid to avoid lysosomal degradation. In yet another embodiment, the nucleic acid can be targeted in vivo for cell specific uptake and expression, 30 by targeting a specific receptor (see, e.g., PCT Publications WO 92/06180; WO 92/22635; WO92/20316; WO93/14188, WO 93/20221). Alternatively, the nucleic acid can be introduced intracellularly and incorporated within host cell DNA for expression, by

homologous recombination (Koller and Smithies, *Proc. Natl. Acad. Sci. USA* 86:8932-8935 (1989); Zijlstra et al., *Nature* 342:435-438 (1989)).

In a specific embodiment, viral vectors that contains nucleic acid sequences encoding an antibody of the invention are used. For example, a retroviral vector can be used (see 5 Miller et al., *Meth. Enzymol.* 217:581-599 (1993)). These retroviral vectors contain the components necessary for the correct packaging of the viral genome and integration into the host cell DNA. The nucleic acid sequences encoding the antibody to be used in gene therapy are cloned into one or more vectors, which facilitates delivery of the gene into a patient. More detail about retroviral vectors can be found in Boesen et al., *Biotherapy* 6:291-302 10 (1994), which describes the use of a retroviral vector to deliver the *mdr1* gene to hematopoietic stem cells in order to make the stem cells more resistant to chemotherapy. Other references illustrating the use of retroviral vectors in gene therapy are: Clowes et al., *J. Clin. Invest.* 93:644-651 (1994); Kiem et al., *Blood* 83:1467-1473 (1994); Salmons and Gunzberg, *Human Gene Therapy* 4:129-141 (1993); and Grossman and Wilson, *Curr. Opin.* 15 in *Genetics and Devel.* 3:110-114 (1993).

Adenoviruses are other viral vectors that can be used in gene therapy. Adenoviruses are especially attractive vehicles for delivering genes to respiratory epithelia. Adenoviruses naturally infect respiratory epithelia where they cause a mild disease. Other targets for adenovirus-based delivery systems are liver, the central nervous system, endothelial cells, 20 and muscle. Adenoviruses have the advantage of being capable of infecting non-dividing cells. Kozarsky and Wilson, *Current Opinion in Genetics and Development* 3:499-503 (1993) present a review of adenovirus-based gene therapy. Bout et al., *Human Gene Therapy* 5:3-10 (1994) demonstrated the use of adenovirus vectors to transfer genes to the respiratory epithelia of rhesus monkeys. Other instances of the use of adenoviruses in gene 25 therapy can be found in Rosenfeld et al., *Science* 252:431-434 (1991); Rosenfeld et al., *Cell* 68:143-155 (1992); Mastrangeli et al., *J. Clin. Invest.* 91:225-234 (1993); PCT Publication WO94/12649; and Wang, et al., *Gene Therapy* 2:775-783 (1995). In a preferred embodiment, adenovirus vectors are used.

Adeno-associated virus (AAV) has also been proposed for use in gene therapy (Walsh 30 et al., *Proc. Soc. Exp. Biol. Med.* 204:289-300 (1993); U.S. Patent No. 5,436,146).

Another approach to gene therapy involves transferring a gene to cells in tissue culture by such methods as electroporation, lipofection, calcium phosphate mediated

transfection, or viral infection. Usually, the method of transfer includes the transfer of a selectable marker to the cells. The cells are then placed under selection to isolate those cells that have taken up and are expressing the transferred gene. Those cells are then delivered to a patient.

5 In this embodiment, the nucleic acid is introduced into a cell prior to administration in vivo of the resulting recombinant cell. Such introduction can be carried out by any method known in the art, including but not limited to transfection, electroporation, microinjection, infection with a viral or bacteriophage vector containing the nucleic acid sequences, cell fusion, chromosome-mediated gene transfer, microcell-mediated gene transfer, spheroplast
10 fusion, etc. Numerous techniques are known in the art for the introduction of foreign genes into cells (see, e.g., Loeffler and Behr, Meth. Enzymol. 217:599-618 (1993); Cohen et al., Meth. Enzymol. 217:618-644 (1993); Cline, Pharmac. Ther. 29:69-92m (1985) and may be used in accordance with the present invention, provided that the necessary developmental and physiological functions of the recipient cells are not disrupted. The technique should
15 provide for the stable transfer of the nucleic acid to the cell, so that the nucleic acid is expressible by the cell and preferably heritable and expressible by its cell progeny.

The resulting recombinant cells can be delivered to a patient by various methods known in the art. Recombinant blood cells (e.g., hematopoietic stem or progenitor cells) are preferably administered intravenously. The amount of cells envisioned for use depends on
20 the desired effect, patient state, etc., and can be determined by one skilled in the art.

Cells into which a nucleic acid can be introduced for purposes of gene therapy encompass any desired, available cell type, and include but are not limited to epithelial cells, endothelial cells, keratinocytes, fibroblasts, muscle cells, hepatocytes; blood cells such as Tlymphocytes, Blymphocytes, monocytes, macrophages, neutrophils, eosinophils,
25 megakaryocytes, granulocytes; various stem or progenitor cells, in particular hematopoietic stem or progenitor cells, e.g., as obtained from bone marrow, umbilical cord blood, peripheral blood, fetal liver, etc.

In a preferred embodiment, the cell used for gene therapy is autologous to the patient.

In an embodiment in which recombinant cells are used in gene therapy, nucleic acid
30 sequences encoding an antibody are introduced into the cells such that they are expressible by the cells or their progeny, and the recombinant cells are then administered in vivo for therapeutic effect. In a specific embodiment, stem or progenitor cells are used. Any stem

and/or progenitor cells which can be isolated and maintained in vitro can potentially be used in accordance with this embodiment of the present invention (see e.g. PCT Publication WO 94/08598; Stemple and Anderson, Cell 71:973-985 (1992); Rheinwald, Meth. Cell Bio. 21A:229 (1980); and Pittelkow and Scott, Mayo Clinic Proc. 61:771 (1986)).

5 In a specific embodiment, the nucleic acid to be introduced for purposes of gene therapy comprises an inducible promoter operably linked to the coding region, such that expression of the nucleic acid is controllable by controlling the presence or absence of the appropriate inducer of transcription. *Demonstration of Therapeutic or Prophylactic Activity*

The compounds or pharmaceutical compositions of the invention are preferably tested
10 in vitro, and then in vivo for the desired therapeutic or prophylactic activity, prior to use in humans. For example, in vitro assays to demonstrate the therapeutic or prophylactic utility of a compound or pharmaceutical composition include, the effect of a compound on a cell line or a patient tissue sample. The effect of the compound or composition on the cell line and/or tissue sample can be determined utilizing techniques known to those of skill in the art
15 including, but not limited to, rosette formation assays and cell lysis assays. In accordance with the invention, in vitro assays which can be used to determine whether administration of a specific compound is indicated, include in vitro cell culture assays in which a patient tissue sample is grown in culture, and exposed to or otherwise administered a compound, and the effect of such compound upon the tissue sample is observed.

Therapeutic/Prophylactic Administration and Composition

20 The invention provides methods of treatment, inhibition and prophylaxis by administration to a subject of an effective amount of a compound or pharmaceutical composition of the invention, preferably an antibody of the invention. In a preferred aspect,
25 the compound is substantially purified (e.g., substantially free from substances that limit its effect or produce undesired side-effects). The subject is preferably an animal, including but not limited to animals such as cows, pigs, horses, chickens, cats, dogs, etc., and is preferably a mammal, and most preferably human.

30 Formulations and methods of administration that can be employed when the compound comprises a nucleic acid or an immunoglobulin are described above; additional appropriate formulations and routes of administration can be selected from among those described herein below.

Various delivery systems are known and can be used to administer a compound of the invention, e.g., encapsulation in liposomes, microparticles, microcapsules, recombinant cells capable of expressing the compound, receptor-mediated endocytosis (see, e.g., Wu and Wu, J. Biol. Chem. 262:4429-4432 (1987)), construction of a nucleic acid as part of a retroviral or other vector, etc. Methods of introduction include but are not limited to intradermal, intramuscular, intraperitoneal, intravenous, subcutaneous, intranasal, epidural, and oral routes. The compounds or compositions may be administered by any convenient route, for example by infusion or bolus injection, by absorption through epithelial or mucocutaneous linings (e.g., oral mucosa, rectal and intestinal mucosa, etc.) and may be administered together with other biologically active agents. Administration can be systemic or local. In addition, it may be desirable to introduce the pharmaceutical compounds or compositions of the invention into the central nervous system by any suitable route, including intraventricular and intrathecal injection; intraventricular injection may be facilitated by an intraventricular catheter, for example, attached to a reservoir, such as an Ommaya reservoir. Pulmonary administration can also be employed, e.g., by use of an inhaler or nebulizer, and formulation with an aerosolizing agent.

In a specific embodiment, it may be desirable to administer the pharmaceutical compounds or compositions of the invention locally to the area in need of treatment; this may be achieved by, for example, and not by way of limitation, local infusion during surgery, topical application, e.g., in conjunction with a wound dressing after surgery, by injection, by means of a catheter, by means of a suppository, or by means of an implant, said implant being of a porous, non-porous, or gelatinous material, including membranes, such as sialastic membranes, or fibers. Preferably, when administering a protein, including an antibody, of the invention, care must be taken to use materials to which the protein does not absorb.

In another embodiment, the compound or composition can be delivered in a vesicle, in particular a liposome (see Langer, Science 249:1527-1533 (1990); Treat et al., in Liposomes in the Therapy of Infectious Disease and Cancer, Lopez-Berestein and Fidler (eds.), Liss, New York, pp. 353- 365 (1989); Lopez-Berestein, *ibid.*, pp. 317-327; see generally *ibid.*)

In yet another embodiment, the compound or composition can be delivered in a controlled release system. In one embodiment, a pump may be used (see Langer, *supra*; Sefton, CRC Crit. Ref. Biomed. Eng. 14:201 (1987); Buchwald et al., Surgery 88:507 (1980);

Saudek et al., N. Engl. J. Med. 321:574 (1989)). In another embodiment, polymeric materials can be used (see Medical Applications of Controlled Release, Langer and Wise (eds.), CRC Pres., Boca Raton, Florida (1974); Controlled Drug Bioavailability, Drug Product Design and Performance, Smolen and Ball (eds.), Wiley, New York (1984); Ranger and Peppas, J., Macromol. Sci. Rev. Macromol. Chem. 23:61 (1983); see also Levy et al., Science 228:190 (1985); During et al., Ann. Neurol. 25:351 (1989); Howard et al., J.Neurosurg. 71:105 (1989)). In yet another embodiment, a controlled release system can be placed in proximity of the therapeutic target, i.e., the brain, thus requiring only a fraction of the systemic dose (see, e.g., Goodson, in Medical Applications of Controlled Release, supra, vol. 2, pp. 115-138 (1984)).

Other controlled release systems are discussed in the review by Langer (Science 249:1527-1533 (1990)).

In a specific embodiment where the compound of the invention is a nucleic acid encoding a protein, the nucleic acid can be administered in vivo to promote expression of its encoded protein, by constructing it as part of an appropriate nucleic acid expression vector and administering it so that it becomes intracellular, e.g., by use of a retroviral vector (see U.S. Patent No. 4,980,286), or by direct injection, or by use of microparticle bombardment (e.g., a gene gun; Biolistic, Dupont), or coating with lipids or cell-surface receptors or transfecting agents, or by administering it in linkage to a homeobox-like peptide which is known to enter the nucleus (see e.g., Joliot et al., Proc. Natl. Acad. Sci. USA 88:1864-1868 (1991)), etc. Alternatively, a nucleic acid can be introduced intracellularly and incorporated within host cell DNA for expression, by homologous recombination.

The present invention also provides pharmaceutical compositions. Such compositions comprise a therapeutically effective amount of a compound, and a pharmaceutically acceptable carrier. In a specific embodiment, the term "pharmaceutically acceptable" means approved by a regulatory agency of the Federal or a state government or listed in the U.S. Pharmacopeia or other generally recognized pharmacopeia for use in animals, and more particularly in humans. The term "carrier" refers to a diluent, adjuvant, excipient, or vehicle with which the therapeutic is administered. Such pharmaceutical carriers can be sterile liquids, such as water and oils, including those of petroleum, animal, vegetable or synthetic origin, such as peanut oil, soybean oil, mineral oil, sesame oil and the like. Water is a preferred carrier when the pharmaceutical composition is administered intravenously. Saline

solutions and aqueous dextrose and glycerol solutions can also be employed as liquid carriers, particularly for injectable solutions. Suitable pharmaceutical excipients include starch, glucose, lactose, sucrose, gelatin, malt, rice, flour, chalk, silica gel, sodium stearate, glycerol monostearate, talc, sodium chloride, dried skim milk, glycerol, propylene, glycol, water, ethanol and the like. The composition, if desired, can also contain minor amounts of wetting or emulsifying agents, or pH buffering agents. These compositions can take the form of solutions, suspensions, emulsion, tablets, pills, capsules, powders, sustained-release formulations and the like. The composition can be formulated as a suppository, with traditional binders and carriers such as triglycerides. Oral formulation can include standard carriers such as pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharine, cellulose, magnesium carbonate, etc. Examples of suitable pharmaceutical carriers are described in "Remington's Pharmaceutical Sciences" by E.W. Martin. Such compositions will contain a therapeutically effective amount of the compound, preferably in purified form, together with a suitable amount of carrier so as to provide the form for proper administration to the patient. The formulation should suit the mode of administration.

In a preferred embodiment, the composition is formulated in accordance with routine procedures as a pharmaceutical composition adapted for intravenous administration to human beings. Typically, compositions for intravenous administration are solutions in sterile isotonic aqueous buffer. Where necessary, the composition may also include a solubilizing agent and a local anesthetic such as lignocaine to ease pain at the site of the injection. Generally, the ingredients are supplied either separately or mixed together in unit dosage form, for example, as a dry lyophilized powder or water free concentrate in a hermetically sealed container such as an ampoule or sachette indicating the quantity of active agent. Where the composition is to be administered by infusion, it can be dispensed with an infusion bottle containing sterile pharmaceutical grade water or saline. Where the composition is administered by injection, an ampoule of sterile water for injection or saline can be provided so that the ingredients may be mixed prior to administration.

The compounds of the invention can be formulated as neutral or salt forms. Pharmaceutically acceptable salts include those formed with anions such as those derived from hydrochloric, phosphoric, acetic, oxalic, tartaric acids, etc., and those formed with

cations such as those derived from sodium, potassium, ammonium, calcium, ferric hydroxides, isopropylamine, triethylamine, 2-ethylamino ethanol, histidine, procaine, etc.

5 The amount of the compound of the invention which will be effective in the treatment, inhibition and prevention of a disease or disorder associated with aberrant expression and/or activity of a polypeptide of the invention can be determined by standard clinical techniques. In addition, in vitro assays may optionally be employed to help identify optimal dosage ranges. The precise dose to be employed in the formulation will also depend on the route of administration, and the seriousness of the disease or disorder, and should be decided according to the judgment of the practitioner and each patient's circumstances.

10 Effective doses may be extrapolated from dose-response curves derived from in vitro or animal model test systems.

For antibodies, the dosage administered to a patient is typically 0.1 mg/kg to 100 mg/kg of the patient's body weight. Preferably, the dosage administered to a patient is between 0.1 mg/kg and 20 mg/kg of the patient's body weight, more preferably 1 mg/kg to 10 mg/kg of the patient's body weight. Generally, human antibodies have a longer half-life within the human body than antibodies from other species due to the immune response to the foreign polypeptides. Thus, lower dosages of human antibodies and less frequent administration is often possible. Further, the dosage and frequency of administration of antibodies of the invention may be reduced by enhancing uptake and tissue penetration (e.g.,

15 into the brain) of the antibodies by modifications such as, for example, lipidation.

20

The invention also provides a pharmaceutical pack or kit comprising one or more containers filled with one or more of the ingredients of the pharmaceutical compositions of the invention. Optionally associated with such container(s) can be a notice in the form prescribed by a governmental agency regulating the manufacture, use or sale of pharmaceuticals or biological products, which notice reflects approval by the agency of manufacture, use or sale for human administration.

25

Diagnosis and Imaging

Labeled antibodies, and derivatives and analogs thereof, which specifically bind to a polypeptide of interest can be used for diagnostic purposes to detect, diagnose, or monitor diseases, disorders, and/or conditions associated with the aberrant expression and/or activity of a polypeptide of the invention. The invention provides for the detection of aberrant

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expression of a polypeptide of interest, comprising (a) assaying the expression of the polypeptide of interest in cells or body fluid of an individual using one or more antibodies specific to the polypeptide interest and (b) comparing the level of gene expression with a standard gene expression level, whereby an increase or decrease in the assayed polypeptide gene expression level compared to the standard expression level is indicative of aberrant expression.

The invention provides a diagnostic assay for diagnosing a disorder, comprising (a) assaying the expression of the polypeptide of interest in cells or body fluid of an individual using one or more antibodies specific to the polypeptide interest and (b) comparing the level of gene expression with a standard gene expression level, whereby an increase or decrease in the assayed polypeptide gene expression level compared to the standard expression level is indicative of a particular disorder. With respect to cancer, the presence of a relatively high amount of transcript in biopsied tissue from an individual may indicate a predisposition for the development of the disease, or may provide a means for detecting the disease prior to the appearance of actual clinical symptoms. A more definitive diagnosis of this type may allow health professionals to employ preventative measures or aggressive treatment earlier thereby preventing the development or further progression of the cancer.

Antibodies of the invention can be used to assay protein levels in a biological sample using classical immunohistological methods known to those of skill in the art (e.g., see Jalkanen, et al., J. Cell. Biol. 101:976-985 (1985); Jalkanen, et al., J. Cell. Biol. 105:3087-3096 (1987)). Other antibody-based methods useful for detecting protein gene expression include immunoassays, such as the enzyme linked immunosorbent assay (ELISA) and the radioimmunoassay (RIA). Suitable antibody assay labels are known in the art and include enzyme labels, such as, glucose oxidase; radioisotopes, such as iodine (125I, 121I), carbon (14C), sulfur (35S), tritium (3H), indium (112In), and technetium (99Tc); luminescent labels, such as luminol; and fluorescent labels, such as fluorescein and rhodamine, and biotin.

One aspect of the invention is the detection and diagnosis of a disease or disorder associated with aberrant expression of a polypeptide of interest in an animal, preferably a mammal and most preferably a human. In one embodiment, diagnosis comprises: a) administering (for example, parenterally, subcutaneously, or intraperitoneally) to a subject an effective amount of a labeled molecule which specifically binds to the polypeptide of interest; b) waiting for a time interval following the administering for permitting the labeled

molecule to preferentially concentrate at sites in the subject where the polypeptide is expressed (and for unbound labeled molecule to be cleared to background level); c) determining background level; and d) detecting the labeled molecule in the subject, such that detection of labeled molecule above the background level indicates that the subject has a particular disease or disorder associated with aberrant expression of the polypeptide of interest. Background level can be determined by various methods including, comparing the amount of labeled molecule detected to a standard value previously determined for a particular system.

It will be understood in the art that the size of the subject and the imaging system used will determine the quantity of imaging moiety needed to produce diagnostic images. In the case of a radioisotope moiety, for a human subject, the quantity of radioactivity injected will normally range from about 5 to 20 millicuries of ^{99m}Tc. The labeled antibody or antibody fragment will then preferentially accumulate at the location of cells which contain the specific protein. In vivo tumor imaging is described in S.W. Burchiel et al., "Immunopharmacokinetics of Radiolabeled Antibodies and Their Fragments." (Chapter 13 in Tumor Imaging: The Radiochemical Detection of Cancer, S.W. Burchiel and B. A. Rhodes, eds., Masson Publishing Inc. (1982).

Depending on several variables, including the type of label used and the mode of administration, the time interval following the administration for permitting the labeled molecule to preferentially concentrate at sites in the subject and for unbound labeled molecule to be cleared to background level is 6 to 48 hours or 6 to 24 hours or 6 to 12 hours. In another embodiment the time interval following administration is 5 to 20 days or 5 to 10 days.

In an embodiment, monitoring of the disease or disorder is carried out by repeating the method for diagnosing the disease or disease, for example, one month after initial diagnosis, six months after initial diagnosis, one year after initial diagnosis, etc.

Presence of the labeled molecule can be detected in the patient using methods known in the art for in vivo scanning. These methods depend upon the type of label used. Skilled artisans will be able to determine the appropriate method for detecting a particular label. Methods and devices that may be used in the diagnostic methods of the invention include, but are not limited to, computed tomography (CT), whole body scan such as position emission tomography (PET), magnetic resonance imaging (MRI), and sonography.

In a specific embodiment, the molecule is labeled with a radioisotope and is detected in the patient using a radiation responsive surgical instrument (Thurston et al., U.S. Patent No. 5,441,050). In another embodiment, the molecule is labeled with a fluorescent compound and is detected in the patient using a fluorescence responsive scanning instrument.

5 In another embodiment, the molecule is labeled with a positron emitting metal and is detected in the patient using positron emission-tomography. In yet another embodiment, the molecule is labeled with a paramagnetic label and is detected in a patient using magnetic resonance imaging (MRI).

10 *Kits*

The present invention provides kits that can be used in the above methods. In one embodiment, a kit comprises an antibody of the invention, preferably a purified antibody, in one or more containers. In a specific embodiment, the kits of the present invention contain a substantially isolated polypeptide comprising an epitope which is specifically

15 immunoreactive with an antibody included in the kit. Preferably, the kits of the present invention further comprise a control antibody which does not react with the polypeptide of interest. In another specific embodiment, the kits of the present invention contain a means for detecting the binding of an antibody to a polypeptide of interest (e.g., the antibody may be conjugated to a detectable substrate such as a fluorescent compound, an enzymatic substrate,

20 a radioactive compound or a luminescent compound, or a second antibody which recognizes the first antibody may be conjugated to a detectable substrate).

In another specific embodiment of the present invention, the kit is a diagnostic kit for use in screening serum containing antibodies specific against proliferative and/or cancerous polynucleotides and polypeptides. Such a kit may include a control antibody that does not

25 react with the polypeptide of interest. Such a kit may include a substantially isolated polypeptide antigen comprising an epitope which is specifically immunoreactive with at least one anti-polypeptide antigen antibody. Further, such a kit includes means for detecting the binding of said antibody to the antigen (e.g., the antibody may be conjugated to a fluorescent compound such as fluorescein or rhodamine which can be detected by flow cytometry). In

30 specific embodiments, the kit may include a recombinantly produced or chemically synthesized polypeptide antigen. The polypeptide antigen of the kit may also be attached to a solid support.

In a more specific embodiment the detecting means of the above-described kit includes a solid support to which said polypeptide antigen is attached. Such a kit may also include a non-attached reporter-labeled anti-human antibody. In this embodiment, binding of the antibody to the polypeptide antigen can be detected by binding of the said reporter-labeled antibody.

In an additional embodiment, the invention includes a diagnostic kit for use in screening serum containing antigens of the polypeptide of the invention. The diagnostic kit includes a substantially isolated antibody specifically immunoreactive with polypeptide or polynucleotide antigens, and means for detecting the binding of the polynucleotide or polypeptide antigen to the antibody. In one embodiment, the antibody is attached to a solid support. In a specific embodiment, the antibody may be a monoclonal antibody. The detecting means of the kit may include a second, labeled monoclonal antibody. Alternatively, or in addition, the detecting means may include a labeled, competing antigen.

In one diagnostic configuration, test serum is reacted with a solid phase reagent having a surface-bound antigen obtained by the methods of the present invention. After binding with specific antigen antibody to the reagent and removing unbound serum components by washing, the reagent is reacted with reporter-labeled anti-human antibody to bind reporter to the reagent in proportion to the amount of bound anti-antigen antibody on the solid support. The reagent is again washed to remove unbound labeled antibody, and the amount of reporter associated with the reagent is determined. Typically, the reporter is an enzyme which is detected by incubating the solid phase in the presence of a suitable fluorometric, luminescent or colorimetric substrate (Sigma, St. Louis, MO).

The solid surface reagent in the above assay is prepared by known techniques for attaching protein material to solid support material, such as polymeric beads, dip sticks, 96-well plate or filter material. These attachment methods generally include non-specific adsorption of the protein to the support or covalent attachment of the protein, typically through a free amine group, to a chemically reactive group on the solid support, such as an activated carboxyl, hydroxyl, or aldehyde group. Alternatively, streptavidin coated plates can be used in conjunction with biotinylated antigen(s).

Thus, the invention provides an assay system or kit for carrying out this diagnostic method. The kit generally includes a support with surface-bound recombinant antigens, and a reporter-labeled anti-human antibody for detecting surface-bound anti-antigen antibody.

Diagnostic Assays

The present invention further relates to methods for assaying staphylococcal infection in an animal by detecting the expression of genes encoding staphylococcal polypeptides of the present invention. The methods comprise analyzing tissue or body fluid from the animal for *Staphylococcus*-specific antibodies, nucleic acids, or proteins. Analysis of nucleic acid specific to *Staphylococcus* is assayed by PCR or hybridization techniques using nucleic acid sequences of the present invention as either hybridization probes or primers. See, e.g., Sambrook et al. Molecular cloning: A Laboratory Manual (Cold Spring Harbor Laboratory Press, 2nd ed., 1989, page 54 reference); Ereemeeva et al. (1994) J. Clin. Microbiol. 32:803-810 (describing differentiation among spotted fever group *Rickettsiae* species by analysis of restriction fragment length polymorphism of PCR-amplified DNA) and Chen et al. 1994 J. Clin. Microbiol. 32:589-595 (detecting bacterial nucleic acids via PCR).

Where diagnosis of a disease state related to infection with *Staphylococcus* has already been made, the present invention is useful for monitoring progression or regression of the disease state by measuring the amount of *Staphylococcus* cells present in a patient or whereby patients exhibiting enhanced *Staphylococcus* gene expression will experience a worse clinical outcome relative to patients expressing these gene(s) at a lower level.

By "biological sample" is intended any biological sample obtained from an animal, cell line, tissue culture, or other source which contains *Staphylococcus* polypeptide, mRNA, or DNA. Biological samples include body fluids (such as saliva, blood, plasma, urine, mucus, synovial fluid, etc.) tissues (such as muscle, skin, and cartilage) and any other biological source suspected of containing *Staphylococcus* polypeptides or nucleic acids. Methods for obtaining biological samples such as tissue are well known in the art.

The present invention is useful for detecting diseases related to *Staphylococcus* infections in animals. Preferred animals include monkeys, apes, cats, dogs, birds, cows, pigs, mice, horses, rabbits and humans. Particularly preferred are humans.

Total RNA can be isolated from a biological sample using any suitable technique such as the single-step guanidinium-thiocyanate-phenol-chloroform method described in Chomczynski et al. (1987) Anal. Biochem. 162:156-159. mRNA encoding *Staphylococcus* polypeptides having sufficient homology to the nucleic acid sequences identified in Table 1 to allow for hybridization between complementary sequences are then assayed using any

appropriate method. These include Northern blot analysis, S1 nuclease mapping, the polymerase chain reaction (PCR), reverse transcription in combination with the polymerase chain reaction (RT-PCR), and reverse transcription in combination with the ligase chain reaction (RT-LCR).

5 Northern blot analysis can be performed as described in Harada et al. (1990) Cell 63:303-312. Briefly, total RNA is prepared from a biological sample as described above. For the Northern blot, the RNA is denatured in an appropriate buffer (such as glyoxal/dimethyl sulfoxide/sodium phosphate buffer), subjected to agarose gel electrophoresis, and transferred onto a nitrocellulose filter. After the RNAs have been linked
10 to the filter by a UV linker, the filter is prehybridized in a solution containing formamide, SSC, Denhardt's solution, denatured salmon sperm, SDS, and sodium phosphate buffer. A *S. aureus* polynucleotide sequence shown in Table 1 labeled according to any appropriate method (such as the ³²P-multiprimed DNA labeling system (Amersham)) is used as probe. After hybridization overnight, the filter is washed and exposed to x-ray film. DNA for use as
15 probe according to the present invention is described in the sections above and will preferably at least 15 nucleotides in length.

S1 mapping can be performed as described in Fujita et al. (1987) Cell 49:357-367. To prepare probe DNA for use in S1 mapping, the sense strand of an above-described *S. aureus* DNA sequence of the present invention is used as a template to synthesize labeled antisense
20 DNA. The antisense DNA can then be digested using an appropriate restriction endonuclease to generate further DNA probes of a desired length. Such antisense probes are useful for visualizing protected bands corresponding to the target mRNA (i.e., mRNA encoding polypeptides of the present invention).

Levels of mRNA encoding *Staphylococcus* polypeptides are assayed, for e.g., using
25 the RT-PCR method described in Makino et al. (1990) Technique 2:295-301. By this method, the radioactivities of the "amplicons" in the polyacrylamide gel bands are linearly related to the initial concentration of the target mRNA. Briefly, this method involves adding total RNA isolated from a biological sample in a reaction mixture containing a RT primer and appropriate buffer. After incubating for primer annealing, the mixture can be supplemented
30 with a RT buffer, dNTPs, DTT, RNase inhibitor and reverse transcriptase. After incubation to achieve reverse transcription of the RNA, the RT products are then subject to PCR using labeled primers. Alternatively, rather than labeling the primers, a labeled dNTP can be

included in the PCR reaction mixture. PCR amplification can be performed in a DNA thermal cyclor according to conventional techniques. After a suitable number of rounds to achieve amplification, the PCR reaction mixture is electrophoresed on a polyacrylamide gel. After drying the gel, the radioactivity of the appropriate bands (corresponding to the mRNA encoding the *Staphylococcus* polypeptides of the present invention) are quantified using an
5 imaging analyzer. RT and PCR reaction ingredients and conditions, reagent and gel concentrations, and labeling methods are well known in the art. Variations on the RT-PCR method will be apparent to the skilled artisan. Other PCR methods that can detect the nucleic acid of the present invention can be found in PCR PRIMER: A LABORATORY MANUAL
10 (C.W. Dieffenbach et al. eds., Cold Spring Harbor Lab Press, 1995).

The polynucleotides of the present invention, including both DNA and RNA, may be used to detect polynucleotides of the present invention or *Staphylococcus* species including *S. aureus* using bio chip technology. The present invention includes both high density chip arrays (>1000 oligonucleotides per cm^2) and low density chip arrays (<1000 oligonucleotides
15 per cm^2). Bio chips comprising arrays of polynucleotides of the present invention may be used to detect *Staphylococcus* species, including *S. aureus*, in biological and environmental samples and to diagnose an animal, including humans, with an *S. aureus* or other *Staphylococcus* infection. The bio chips of the present invention may comprise polynucleotide sequences of other pathogens including bacteria, viral, parasitic, and fungal
20 polynucleotide sequences, in addition to the polynucleotide sequences of the present invention, for use in rapid differential pathogenic detection and diagnosis. The bio chips can also be used to monitor an *S. aureus* or other *Staphylococcus* infections and to monitor the genetic changes (deletions, insertions, mismatches, etc.) in response to drug therapy in the clinic and drug development in the laboratory. The bio chip technology comprising arrays of
25 polynucleotides of the present invention may also be used to simultaneously monitor the expression of a multiplicity of genes, including those of the present invention. The polynucleotides used to comprise a selected array may be specified in the same manner as for the fragments, i.e., by their 5' and 3' positions or length in contiguous base pairs and include from. Methods and particular uses of the polynucleotides of the present invention to detect
30 *Staphylococcus* species, including *S. aureus*, using bio chip technology include those known in the art and those of: U.S. Patent Nos. 5510270, 5545531, 5445934, 5677195, 5532128, 5556752, 5527681, 5451683, 5424186, 5607646, 5658732 and World Patent Nos.

WO/9710365, WO/9511995, WO/9743447, WO/9535505, each incorporated herein in their entireties.

Biosensors using the polynucleotides of the present invention may also be used to detect, diagnose, and monitor *S. aureus* or other *Staphylococcus* species and infections thereof. Biosensors using the polynucleotides of the present invention may also be used to detect particular polynucleotides of the present invention. Biosensors using the polynucleotides of the present invention may also be used to monitor the genetic changes (deletions, insertions, mismatches, etc.) in response to drug therapy in the clinic and drug development in the laboratory. Methods and particular uses of the polynucleotides of the present invention to detect *Staphylococcus* species, including *S. aureus*, using biosensors include those known in the art and those of: U.S. Patent Nos 5721102, 5658732, 5631170, and World Patent Nos. WO97/35011, WO/9720203, each incorporated herein in their entireties.

Thus, the present invention includes both bio chips and biosensors comprising polynucleotides of the present invention and methods of their use.

A preferred composition of matter comprises isolated nucleic acid molecules wherein the nucleotide sequences of said nucleic acid molecules comprise a bio chip or biosensor of at least 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25, 30, 40, 50, 100, 150, 200, 250, 300, 500, 1000, 2000, 3000 or 4000 nucleotide sequences, wherein at least one sequence in said DNA bio chip or biosensor is at least 95% identical to a sequence of at least 50 contiguous nucleotides in a *S. aureus* polynucleotide shown in Table 1. The nucleic acid molecules can comprise DNA molecules or RNA molecules.

Assaying *Staphylococcus* polypeptide levels in a biological sample can occur using any art-known method, such as antibody-based techniques. For example, *Staphylococcus* polypeptide expression in tissues can be studied with classical immunohistological methods. In these, the specific recognition is provided by the primary antibody (polyclonal or monoclonal) but the secondary detection system can utilize fluorescent, enzyme, or other conjugated secondary antibodies. As a result, an immunohistological staining of tissue section for pathological examination is obtained. Tissues can also be extracted, e.g., with urea and neutral detergent, for the liberation of *Staphylococcus* polypeptides for Western-blot or dot/slot assay. See, e.g., Jalkanen, M. et al. (1985) J. Cell. Biol. 101:976-985; Jalkanen, M. et al. (1987) J. Cell. Biol. 105:3087-3096. In this technique, which is based on the use of

cationic solid phases, quantitation of a *Staphylococcus* polypeptide can be accomplished using an isolated *Staphylococcus* polypeptide as a standard. This technique can also be applied to body fluids.

Other antibody-based methods useful for detecting *Staphylococcus* polypeptide gene expression include immunoassays, such as the ELISA and the radioimmunoassay (RIA). For example, a *Staphylococcus* polypeptide-specific monoclonal antibodies can be used both as an immunoabsorbent and as an enzyme-labeled probe to detect and quantify a *Staphylococcus* polypeptide. The amount of a *Staphylococcus* polypeptide present in the sample can be calculated by reference to the amount present in a standard preparation using a linear regression computer algorithm. Such an ELISA is described in Iacobelli et al. (1988) Breast Cancer Research and Treatment 11:19-30. In another ELISA assay, two distinct specific monoclonal antibodies can be used to detect *Staphylococcus* polypeptides in a body fluid. In this assay, one of the antibodies is used as the immunoabsorbent and the other as the enzyme-labeled probe.

The above techniques may be conducted essentially as a "one-step" or "two-step" assay. The "one-step" assay involves contacting the *Staphylococcus* polypeptide with immobilized antibody and, without washing, contacting the mixture with the labeled antibody. The "two-step" assay involves washing before contacting the mixture with the labeled antibody. Other conventional methods may also be employed as suitable. It is usually desirable to immobilize one component of the assay system on a support, thereby allowing other components of the system to be brought into contact with the component and readily removed from the sample. Variations of the above and other immunological methods included in the present invention can also be found in Harlow et al., ANTIBODIES: A LABORATORY MANUAL, (Cold Spring Harbor Laboratory Press, 2nd ed. 1988).

Suitable enzyme labels include, for example, those from the oxidase group, which catalyze the production of hydrogen peroxide by reacting with substrate. Glucose oxidase is particularly preferred as it has good stability and its substrate (glucose) is readily available. Activity of an oxidase label may be assayed by measuring the concentration of hydrogen peroxide formed by the enzyme-labeled antibody/substrate reaction. Besides enzymes, other suitable labels include radioisotopes, such as iodine (^{125}I , ^{121}I), carbon (^{14}C), sulphur (^{35}S), tritium (^3H), indium (^{112}In), and technetium ($^{99\text{m}}\text{Tc}$), and fluorescent labels, such as fluorescein and rhodamine, and biotin.

Further suitable labels for the *Staphylococcus* polypeptide-specific antibodies of the present invention are provided below. Examples of suitable enzyme labels include malate dehydrogenase, *Staphylococcus* nuclease, delta-5-steroid isomerase, yeast-alcohol dehydrogenase, alpha-glycerol phosphate dehydrogenase, triose phosphate isomerase, 5 peroxidase, alkaline phosphatase, asparaginase, glucose oxidase, beta-galactosidase, ribonuclease, urease, catalase, glucose-6-phosphate dehydrogenase, glucoamylase, and acetylcholine esterase.

Examples of suitable radioisotopic labels include ^3H , ^{111}In , ^{125}I , ^{131}I , ^{32}P , ^{35}S , ^{14}C , ^{51}Cr , ^{57}To , ^{58}Co , ^{59}Fe , ^{75}Se , ^{152}Eu , ^{90}Y , ^{67}Cu , ^{217}Ci , ^{211}At , ^{212}Pb , ^{47}Sc , ^{109}Pd , etc. ^{111}In is a 10 preferred isotope where *in vivo* imaging is used since it avoids the problem of dehalogenation of the ^{125}I or ^{131}I -labeled monoclonal antibody by the liver. In addition, this radionucleotide has a more favorable gamma emission energy for imaging. See, e.g., Perkins et al. (1985) Eur. J. Nucl. Med. 10:296-301; Carasquillo et al. (1987) J. Nucl. Med. 28:281-287. For example, ^{111}In coupled to monoclonal antibodies with 15 1-(P-isothiocyanatobenzyl)-DPTA has shown little uptake in non-tumors tissues, particularly the liver, and therefore enhances specificity of tumor localization. See, Esteban et al. (1987) J. Nucl. Med. 28:861-870.

Examples of suitable non-radioactive isotopic labels include ^{157}Gd , ^{55}Mn , ^{162}Dy , ^{52}Tr , and ^{56}Fe .

20 Examples of suitable fluorescent labels include an ^{152}Eu label, a fluorescein label, an isothiocyanate label, a rhodamine label, a phycoerythrin label, a phycocyanin label, an allophycocyanin label, an o-phthaldehyde label, and a fluorescamine label.

Examples of suitable toxin labels include, *Pseudomonas* toxin, diphtheria toxin, ricin, and cholera toxin.

25 Examples of chemiluminescent labels include a luminal label, an isoluminal label, an aromatic acridinium ester label, an imidazole label, an acridinium salt label, an oxalate ester label, a luciferin label, a luciferase label, and an aequorin label.

Examples of nuclear magnetic resonance contrasting agents include heavy metal nuclei such as Gd, Mn, and iron.

30 Typical techniques for binding the above-described labels to antibodies are provided by Kennedy et al. (1976) Clin. Chim. Acta 70:1-31, and Schurs et al. (1977) Clin. Chim. Acta 81:1-40. Coupling techniques mentioned in the latter are the glutaraldehyde method, the

periodate method, the dimaleimide method, the m-maleimidobenzyl-N-hydroxy-succinimide ester method, all of which methods are incorporated by reference herein.

In a related aspect, the invention includes a diagnostic kit for use in screening serum containing antibodies specific against *S. aureus* infection. Such a kit may include an isolated *S. aureus* antigen comprising an epitope which is specifically immunoreactive with at least one anti-*S. aureus* antibody. Such a kit also includes means for detecting the binding of said antibody to the antigen. In specific embodiments, the kit may include a recombinantly produced or chemically synthesized peptide or polypeptide antigen. The peptide or polypeptide antigen may be attached to a solid support.

In a more specific embodiment, the detecting means of the above-described kit includes a solid support to which said peptide or polypeptide antigen is attached. Such a kit may also include a non-attached reporter-labeled anti-human antibody. In this embodiment, binding of the antibody to the *S. aureus* antigen can be detected by binding of the reporter labeled antibody to the anti-*S. aureus* polypeptide antibody.

In a related aspect, the invention includes a method of detecting *S. aureus* infection in a subject. This detection method includes reacting a body fluid, preferably serum, from the subject with an isolated *S. aureus* antigen, and examining the antigen for the presence of bound antibody. In a specific embodiment, the method includes a polypeptide antigen attached to a solid support, and serum is reacted with the support. Subsequently, the support is reacted with a reporter-labeled anti-human antibody. The support is then examined for the presence of reporter-labeled antibody.

The solid surface reagent employed in the above assays and kits is prepared by known techniques for attaching protein material to solid support material, such as polymeric beads, dip sticks, 96-well plates or filter material. These attachment methods generally include non-specific adsorption of the protein to the support or covalent attachment of the protein, typically through a free amine group, to a chemically reactive group on the solid support, such as an activated carboxyl, hydroxyl, or aldehyde group. Alternatively, streptavidin coated plates can be used in conjunction with biotinylated antigen(s).

The polypeptides and antibodies of the present invention, including fragments thereof, may be used to detect *Staphylococcus* species including *S. aureus* using bio chip and biosensor technology. Bio chip and biosensors of the present invention may comprise the polypeptides of the present invention to detect antibodies, which specifically recognize

Staphylococcus species, including *S. aureus*. Bio chip and biosensors of the present invention may also comprise antibodies which specifically recognize the polypeptides of the present invention to detect Staphylococcus species, including *S. aureus* or specific polypeptides of the present invention. Bio chips or biosensors comprising polypeptides or antibodies of the present invention may be used to detect Staphylococcus species, including *S. aureus*, in biological and environmental samples and to diagnose an animal, including humans, with an *S. aureus* or other Staphylococcus infection. Thus, the present invention includes both bio chips and biosensors comprising polypeptides or antibodies of the present invention and methods of their use.

The bio chips of the present invention may further comprise polypeptide sequences of other pathogens including bacteria, viral, parasitic, and fungal polypeptide sequences, in addition to the polypeptide sequences of the present invention, for use in rapid differential pathogenic detection and diagnosis. The bio chips of the present invention may further comprise antibodies or fragments thereof specific for other pathogens including bacteria, viral, parasitic, and fungal polypeptide sequences, in addition to the antibodies or fragments thereof of the present invention, for use in rapid differential pathogenic detection and diagnosis. The bio chips and biosensors of the present invention may also be used to monitor an *S. aureus* or other Staphylococcus infection and to monitor the genetic changes (amino acid deletions, insertions, substitutions, etc.) in response to drug therapy in the clinic and drug development in the laboratory. The bio chip and biosensors comprising polypeptides or antibodies of the present invention may also be used to simultaneously monitor the expression of a multiplicity of polypeptides, including those of the present invention. The polypeptides used to comprise a bio chip or biosensor of the present invention may be specified in the same manner as for the fragments, i.e, by their N-terminal and C-terminal positions or length in contiguous amino acid residue. Methods and particular uses of the polypeptides and antibodies of the present invention to detect Staphylococcus species, including *S. aureus*, or specific polypeptides using bio chip and biosensor technology include those known in the art, those of the U.S. Patent Nos. and World Patent Nos. listed above for bio chips and biosensors using polynucleotides of the present invention, and those of: U.S. Patent Nos. 5658732, 5135852, 5567301, 5677196, 5690894 and World Patent Nos. WO9729366, WO9612957, each incorporated herein in their entireties.

Treatment

Agonists and Antagonists - Assays and Molecules

The invention also provides a method of screening compounds to identify those which enhance or block the biological activity of the *S. aureus* polypeptides of the present invention. The present invention further provides where the compounds kill or slow the growth of *S. aureus*. The ability of *S. aureus* antagonists, including *S. aureus* ligands, to prophylactically or therapeutically block antibiotic resistance may be easily tested by the skilled artisan. See, e.g., Straden et al. (1997) J Bacteriol. 179(1):9-16.

An agonist is a compound which increases the natural biological function or which functions in a manner similar to the polypeptides of the present invention, while antagonists decrease or eliminate such functions. Potential antagonists include small organic molecules, peptides, polypeptides, and antibodies that bind to a polypeptide of the invention and thereby inhibit or extinguish its activity.

The antagonists may be employed for instance to inhibit peptidoglycan cross bridge formation. Antibodies against *S. aureus* may be employed to bind to and inhibit *S. aureus* activity to treat antibiotic resistance. Any of the above antagonists may be employed in a composition with a pharmaceutically acceptable carrier.

Polypeptides and polynucleotides of the invention may also be used to assess the binding of small molecule substrates and ligands in, for example, cell-free preparations, chemical libraries, and natural product mixtures. These substrates and ligands may be natural substrates and ligands or may be structural or functional mimetics. See, e.g., Coligan et al., Current Protocols in Immunology 1(2): Chapter 5 (1991).

Polypeptides and polynucleotides of the present invention are responsible for many biological functions, including many disease states, in particular the Diseases hereinbefore mentioned. It is therefor desirable to devise screening methods to identify compounds which stimulate or which inhibit the function of the polypeptide or polynucleotide. Accordingly, in a further aspect, the present invention provides for a method of screening compounds to identify those which stimulate or which inhibit the function of a polypeptide or a polynucleotide of the invention, as well as related polypeptides and polynucleotides. In general, agonists or antagonists may be employed for therapeutic and prophylactic purposes for such Diseases as hereinbefore mentioned. Compounds may be identified from a variety of sources, for example, cells, cell-free preparations, chemical libraries, and natural product

mixtures. Such agonists, antagonists or inhibitors so-identified may be natural or modified substrates, ligands, receptors, enzymes, etc., as the case may be, of *S. aureus* polypeptides and polynucleotides of the invention; or may be structural or functional mimetics thereof (see Coligan et al., *supra*).

5 The screening methods may simply measure the binding of a candidate compound to the polypeptide or polynucleotide, or to cells or membranes bearing the polypeptide or polynucleotide, or a fusion protein of the polypeptide by means of a label directly or indirectly associated with the candidate compound. Alternatively, the screening method may involve competition with a labeled competitor. Further, these screening methods may test
10 whether the candidate compound results in a signal generated by activation or inhibition of the polypeptide or polynucleotide, using detection systems appropriate to the cells comprising the polypeptide or polynucleotide. Inhibitors of activation are generally assayed in the presence of known agonists and the effect on activation by the agonist by the presence of the candidate compound is observed. Constitutively active polypeptide and/or constitutively
15 expressed polypeptides and polynucleotides may be employed in screening methods for inverse agonists or inhibitors, in the absence of an agonist or inhibitor, by testing whether the candidate compound results in inhibition of activation of the polypeptide or polynucleotide, as the case may be. Further the screening methods may simply comprise the steps of mixing a candidate compound with a solution containing a polypeptide or polynucleotide of the present
20 invention, to form a mixture, measuring *S. aureus* polypeptide and/or polynucleotide activity in the mixture, and comparing the *S. aureus* polypeptide and/or polynucleotide activity of the mixture to a standard. Fusion proteins, such as those made from His tag and *S. aureus* polypeptides of the invention, as described herein, can also be used for high-throughput screening assays to identify antagonists of the polypeptide of the present invention, as well as
25 of phylogenetically and/or functionally related polypeptides (see, e.g., Bennett et al., J. Mol. Recognition 8:52-58 (1995); and Johanson et al., J. Biol. Chem. 270(16):9459-71 (1995)).

30 The polynucleotides, polypeptides and antibodies that bind to and/or interact with a polypeptide of the present invention may also be used to configure screening methods for detecting the effect of added compounds on the production of mRNA and/or polypeptide in cells. For example, an ELISA assay may be constructed for measuring secreted or cell associated levels of polypeptide using monoclonal and polyclonal antibodies by standard methods known in the art. This can be used to discover agents which may inhibit or enhance

the production of polypeptide (also called antagonist or agonists, respectively) from suitably manipulated cells or tissues.

The invention also provides a method of screening compounds to identify those which enhance (agonist) or block (antagonist) the action of *S. aureus* polypeptide or polynucleotide of the invention, particularly those compounds that are bacteristatic and/or bactericidal. The method of screening may involve high-throughput techniques. For example, to screen for agonists or antagonists, a synthetic reaction mix, a cellular compartment, such as a membrane, cell envelope or cell wall, or a preparation of any thereof, comprising a *S. aureus* polypeptide of the invention and a labeled substrate or ligand of such polypeptide is incubated in the absence or the presence of a candidate molecule that may be an agonist or antagonist of a *S. aureus* polypeptide of the invention. The ability of the candidate molecule to agonize or antagonize the *S. aureus* polypeptide is reflected in decreased binding of the labeled ligand or decreased production of product from such substrate. Molecules that bind gratuitously, i.e., without inducing the effects of *S. aureus* polypeptides are most likely to be good antagonists. Molecules that bind well and, as the case may be, increase the rate of product production from substrate, increase signal transduction, or increase chemical channel activity are agonists. Using a reporter system may enhance the detection of the rate or level of, for example, the production of product from substrate, signal transduction, or chemical channel activity. Reporter systems that may be useful in this regard include but are not limited to colorimetric, labeled substrate converted into product, a reporter gene that is responsive to changes in a *S. aureus* polynucleotide or polypeptide activity, and binding assays known in the art.

S. aureus polypeptides of the invention may be used to identify membrane bound or soluble receptors, if any, for such polypeptide, through standard receptor binding techniques known in the art. These techniques include, but are not limited to, ligand binding and crosslinking assays in which the polypeptide is labeled with a radioactive isotope (for instance, ¹²⁵I), chemically modified (for instance, biotinylated), or fused to a peptide sequence suitable for detection or purification (for instance, a His tag), and incubated with a source of the putative receptor (*S. aureus* or human cells, cell membranes, cell supernatants, tissue extracts, bodily materials). Other methods include biophysical techniques such as surface plasmon resonance and spectroscopy. These screening methods may also be used to identify agonists and antagonists of the polypeptide which compete with the binding of the

polypeptide to its receptor(s), if any. Standard methods for conducting such assays are well understood in the art.

The fluorescence polarization value for a fluorescently-tagged molecule depends on the rotational correlation time or tumbling rate. Protein complexes, such as formed by one *S. aureus* polypeptide of the invention associating with itself or another *S. aureus* polypeptide of the invention, labeled to comprise a fluorescently-labeled molecule will have higher polarization values than a fluorescently labeled monomeric protein. It is preferred that this method be used to characterize small molecules that disrupt polypeptide complexes.

Fluorescence energy transfer may also be used to characterize small molecules that interfere with the formation of *S. aureus* polypeptide dimers, trimers, tetramers, or higher order structures, or structures formed by one *S. aureus* polypeptide bound to another polypeptide. *S. aureus* polypeptides can be labeled with both a donor and acceptor fluorophore. Upon mixing of the two labeled species and excitation of the donor fluorophore, fluorescence energy transfer can be detected by observing fluorescence of the acceptor. Compounds that block dimerization will inhibit fluorescence energy transfer.

Surface plasmon resonance can be used to monitor the effect of small molecules on *S. aureus* polypeptide self-association as well as an association of *S. aureus* polypeptide and another polypeptide or small molecule. *S. aureus* polypeptide can be coupled to a sensor chip at low site density such that covalently bound molecules will be monomeric. Solution protein can then be passed over the *S. aureus* polypeptide -coated surface and specific binding can be detected in real-time by monitoring the change in resonance angle caused by a change in local refractive index. This technique can be used to characterize the effect of small molecules on kinetic rates and equilibrium binding constants for *S. aureus* polypeptide self-association as well as an association of *S. aureus* polypeptides with another polypeptide or small molecule.

A scintillation proximity assay may be used to characterize the interaction between an association of *S. aureus* polypeptide with another *S. aureus* polypeptide or a different polypeptide. *S. aureus* polypeptide can be coupled to a scintillation-filled bead. Addition of radio-labeled *S. aureus* polypeptide results in binding where the radioactive source molecule is in close proximity to the scintillation fluid. Thus, signal is emitted upon *S. aureus* polypeptide binding and compounds that prevent *S. aureus* polypeptide self-association or an association of *S. aureus* polypeptide and another polypeptide or small molecule will diminish

signal.

ICS biosensors have been described by AMBRI (Australian Membrane Biotechnology Research Institute). They couple the self-association of macromolecules to the closing of gramicidin-facilitated ion channels in suspended membrane bilayers and hence to a measurable change in the admittance (similar to impedance) of the biosensor. This approach is linear over six decades of admittance change and is ideally suited for large scale, high through-put screening of small molecule combinatorial libraries.

In other embodiments of the invention there are provided methods for identifying compounds which bind to or otherwise interact with and inhibit or activate an activity or expression of a polypeptide and/or polynucleotide of the invention comprising: contacting a polypeptide and/or polynucleotide of the invention with a compound to be screened under conditions to permit binding to or other interaction between the compound and the polypeptide and/or polynucleotide to assess the binding to or other interaction with the compound, such as binding or interaction preferably being associated with a second component capable of providing a detectable signal in response to the binding or interaction of the polypeptide and/or polynucleotide with the compound; and determining whether the compound binds to or otherwise interacts with and activates or inhibits an activity or expression of the polypeptide and/or polynucleotide by detecting the presence or absence of a signal generated from the binding or interaction of the compound with the polypeptide and/or polynucleotide.

Another example of an assay for *S. aureus* polypeptide agonists is a competitive assay that combines a *S. aureus* polypeptide and a potential agonists with *S. aureus* polypeptide-binding molecules, recombinant *S. aureus* polypeptide-binding molecules, natural substrates or ligands, or substrate or ligand mimetics, under appropriate conditions for a competitive inhibition assay. *S. aureus* polypeptide can be labeled, such as by radioactivity or a colorimetric compound, such that the number of *S. aureus* polypeptide molecules bound to a binding molecule or converted to product can be determined accurately to assess the effectiveness of the potential antagonist.

Potential antagonists include, among others, small organic molecules, peptides, polypeptides and antibodies that bind to a polynucleotide and/or polypeptide of the invention and thereby inhibit or extinguish its activity or expression. Potential antagonists also may be small organic molecules, a peptide, a polypeptide such as a closely related protein or antibody.

that binds the same sites on a binding molecule, such as a binding molecule, without inducing *S. aureus* polypeptide induced activities, thereby preventing the action or expression of *S. aureus* polypeptides and/or polynucleotides by excluding *S. aureus* polypeptides and/or polynucleotides from binding.

5 Potential antagonists include a small molecule that binds to and occupies the binding site of the polypeptide thereby preventing binding to cellular binding molecules, such that normal biological activity is prevented. Examples of small molecules include but are not limited to small organic molecules, peptides or peptide-like molecules. Other potential antagonists include antisense molecules (see, e.g., Okano, J. Neurochem. 56:560 (1991);
10 OLIGODEOXYNUCLEOTIDES AS ANTISENSE INHIBITORS OF GENE EXPRESSION, CRC Press, Boca Raton, FL (1998)), for a description of these molecules). Preferred potential antagonists include compounds related to and variants of *S. aureus* polypeptides of the invention.

Other examples of potential *S. aureus* polypeptide antagonists include antibodies or,
15 in some cases, oligonucleotides or proteins which are closely related to the ligands, substrates, receptors, enzymes, etc., as the case may be, of the polypeptide, e.g., a fragment of the ligands, substrates, receptors, enzymes, etc.; or small molecules which bind to the polypeptide of the present invention but do not elicit a response, so that the activity of the polypeptide is prevented.

20 The invention further comprises biomimetics, or functional mimetics of the natural *S. aureus* polypeptides of the invention. These functional mimetics may be used for, among other things, antagonizing the activity of *S. aureus* polypeptide or as an antigen or immunogen in a manner described elsewhere herein. Functional mimetics of the polypeptides of the invention include but are not limited to truncated polypeptides. For example, preferred
25 functional mimetics include, a polypeptide comprising a polypeptide sequence set forth in Table I lacking 20, 30, 40, 50, 60, 70, or 80 amino- or carboxy-terminal amino acid residues, including fusion proteins comprising one or more of these truncated sequences. Polynucleotides encoding each of these functional mimetics may be used as expression cassettes to express each mimetic polypeptide. It is preferred that these cassettes comprise 5'
30 and 3' restriction sites to allow for a convenient means to ligate the cassettes together when desired. It is further preferred that these cassettes comprise gene expression signals known in the art or described elsewhere herein.

Thus, in another aspect, the present invention relates to a screening kit for identifying agonists, antagonists, ligands, receptors, substrates, enzymes, etc. for a polypeptide and/or polynucleotide of the present invention; or compounds which decrease or enhance the production of such polypeptides and/or polynucleotides, which comprises: (a) a polypeptide and/or a polynucleotide of the present invention; (b) a recombinant cell expressing a polypeptide and/or polynucleotide of the present invention; (c) a cell membrane expressing a polypeptide and/or a polynucleotide of the present invention; or (d) antibody to a polypeptide and/or polynucleotide of the present invention; which polypeptide is preferably one of the *S. aureus* polypeptides shown in Table 1, and which polynucleotide is preferably one of the *S. aureus* polynucleotides shown in Table 1.

It will be appreciated that in any such kit, (a), (b), (c), or (d) may comprise a substantial component.

It will be readily appreciated by the skilled artisan that a polypeptide and/or polynucleotide of the present invention may also be used in a method for the structure-based design of an agonist, antagonist or inhibitor of the polypeptide and/or polynucleotide, by: (a) determining in the first instance the three-dimensional structure of the polypeptide and/or polynucleotide, or complexes thereof; (b) deducing the three-dimensional structure for the likely reactive site(s), binding site(s) or motif(s) of an agonist, antagonist or inhibitor; (c) synthesizing candidate compounds that are predicted to bind to or react with the deduced binding site(s), reactive(s), and/or motif(s); and (d) testing whether the candidate compounds are indeed agonists, antagonists or inhibitors. It will be further appreciated that this will normally be an iterative process, and this iterative process may be performed using automated and computer-controlled steps.

Each of the polynucleotide sequences provided herein may be used in the discovery and development of antibacterial compounds. The encoded protein, upon expression, can be used as a target for the screening of antibacterial drugs. Additionally, the polynucleotide sequences encoding the amino terminal regions of the encoded protein or Shine-Delgarno or other translation facilitating sequences of the respective mRNA can be used to construct antisense sequences to control the expression of the coding sequence of interest.

The invention further encompasses the use of polypeptides, polynucleotides, agonists and/or antagonists of the invention to interfere with the initial physical interaction between a pathogen or pathogens and a eukaryotic, preferably mammalian, host responsible for sequelae

of infection. In particular, the molecules of the invention may be used: in the prevention of adhesion of bacteria, in particular gram positive and/or gram negative bacteria, to eukaryotic, preferably mammalian, extracellular matrix proteins on in-dwelling devices or to extracellular matrix proteins in wounds; to block bacterial adhesion between eukaryotic, preferably mammalian, extracellular matrix proteins and bacterial *S. aureus* proteins that mediate tissue damage and/or; to block the normal progression of pathogenesis in infections initiated other than by the implantation of in-dwelling devices or by other surgical techniques.

In a specific embodiment, the invention provides *S. aureus* polypeptide agonists and antagonists, preferably bacteristatic or bactericidal agonists and antagonists.

The antagonists and agonists of the invention may be employed, for example, to prevent, inhibit and/or treat diseases.

Vaccines

The present invention also provides vaccines comprising one or more polypeptides of the present invention. Heterogeneity in the composition of a vaccine may be provided by combining *S. aureus* polypeptides of the present invention. Multi-component vaccines of this type are desirable because they are likely to be more effective in eliciting protective immune responses against multiple species and strains of the *Staphylococcus* genus than single polypeptide vaccines.

Multi-component vaccines are known in the art to elicit antibody production to numerous immunogenic components. See, e.g., Decker et al. (1996) J. Infect. Dis. 174:S270-275. In addition, a hepatitis B, diphtheria, tetanus, pertussis tetravalent vaccine has recently been demonstrated to elicit protective levels of antibodies in human infants against all four pathogenic agents. See, e.g., Aristegui, J. et al. (1997) Vaccine 15:7-9.

The present invention in addition to single-component vaccines includes multi-component vaccines. These vaccines comprise more than one polypeptide, immunogen or antigen. Thus, a multi-component vaccine would be a vaccine comprising more than one of the *S. aureus* polypeptides of the present invention.

Further within the scope of the invention are whole cell and whole viral vaccines. Such vaccines may be produced recombinantly and involve the expression of one or more of the *S. aureus* polypeptides described in Table 1. For example, the *S. aureus* polypeptides of

the present invention may be either secreted or localized intracellularly, on the cell surface, or in the periplasmic space. Further, when a recombinant virus is used, the *S. aureus* polypeptides of the present invention may, for example, be localized in the viral envelope, on the surface of the capsid, or internally within the capsid. Whole cells vaccines which employ cells expressing heterologous proteins are known in the art. See, e.g., Robinson, K. et al. (1997) Nature Biotech. 15:653-657; Sirard, J. et al. (1997) Infect. Immun. 65:2029-2033; Chabalgoity, J. et al. (1997) Infect. Immun. 65:2402-2412. These cells may be administered live or may be killed prior to administration. Chabalgoity, J. et al., *supra*, for example, report the successful use in mice of a live attenuated *Salmonella* vaccine strain which expresses a portion of a platyhelminth fatty acid-binding protein as a fusion protein on its cells surface.

A multi-component vaccine can also be prepared using techniques known in the art by combining one or more *S. aureus* polypeptides of the present invention, or fragments thereof, with additional non-staphylococcal components (e.g., diphtheria toxin or tetanus toxin, and/or other compounds known to elicit an immune response). Such vaccines are useful for eliciting protective immune responses to both members of the *Staphylococcus* genus and non-staphylococcal pathogenic agents.

The vaccines of the present invention also include DNA vaccines. DNA vaccines are currently being developed for a number of infectious diseases. See, et al., Boyer, et al. (1997) Nat. Med. 3:526-532; reviewed in Spier, R. (1996) Vaccine 14:1285-1288. Such DNA vaccines contain a nucleotide sequence encoding one or more *S. aureus* polypeptides of the present invention oriented in a manner that allows for expression of the subject polypeptide. For example, the direct administration of plasmid DNA encoding *B. burgdorferi* OspA has been shown to elicit protective immunity in mice against borrelial challenge. See, Luke et al. (1997) J. Infect. Dis. 175:91-97.

The present invention also relates to the administration of a vaccine which is co-administered with a molecule capable of modulating immune responses. Kim et al. (1997) Nature Biotech. 15:641-646, for example, report the enhancement of immune responses produced by DNA immunizations when DNA sequences encoding molecules which stimulate the immune response are co-administered. In a similar fashion, the vaccines of the present invention may be co-administered with either nucleic acids encoding immune modulators or the immune modulators themselves. These immune modulators include granulocyte macrophage colony stimulating factor (GM-CSF) and CD86.

The vaccines of the present invention may be used to confer resistance to staphylococcal infection by either passive or active immunization. When the vaccines of the present invention are used to confer resistance to staphylococcal infection through active immunization, a vaccine of the present invention is administered to an animal to elicit a protective immune response which either prevents or attenuates a staphylococcal infection. When the vaccines of the present invention are used to confer resistance to staphylococcal infection through passive immunization, the vaccine is provided to a host animal (e.g., human, dog, or mouse), and the antisera elicited by this antisera is recovered and directly provided to a recipient suspected of having an infection caused by a member of the *Staphylococcus* genus.

The ability to label antibodies, or fragments of antibodies, with toxin molecules provides an additional method for treating staphylococcal infections when passive immunization is conducted. In this embodiment, antibodies, or fragments of antibodies, capable of recognizing the *S. aureus* polypeptides disclosed herein, or fragments thereof, as well as other *Staphylococcus* proteins, are labeled with toxin molecules prior to their administration to the patient. When such toxin derivatized antibodies bind to *Staphylococcus* cells, toxin moieties will be localized to these cells and will cause their death.

The present invention thus concerns and provides a means for preventing or attenuating a staphylococcal infection resulting from organisms which have antigens that are recognized and bound by antisera produced in response to the polypeptides of the present invention. As used herein, a vaccine is said to prevent or attenuate a disease if its administration to an animal results either in the total or partial attenuation (i.e., suppression) of a symptom or condition of the disease, or in the total or partial immunity of the animal to the disease.

The administration of the vaccine (or the antisera which it elicits) may be for either a "prophylactic" or "therapeutic" purpose. When provided prophylactically, the compound(s) are provided in advance of any symptoms of staphylococcal infection. The prophylactic administration of the compound(s) serves to prevent or attenuate any subsequent infection. When provided therapeutically, the compound(s) is provided upon or after the detection of symptoms which indicate that an animal may be infected with a member of the *Staphylococcus* genus. The therapeutic administration of the compound(s) serves to attenuate any actual infection. Thus, the *S. aureus* polypeptides, and fragments thereof, of the present

invention may be provided either prior to the onset of infection (so as to prevent or attenuate an anticipated infection) or after the initiation of an actual infection.

The polypeptides of the invention, whether encoding a portion of a native protein or a functional derivative thereof, may be administered in pure form or may be coupled to a macromolecular carrier. Example of such carriers are proteins and carbohydrates. Suitable proteins which may act as macromolecular carrier for enhancing the immunogenicity of the polypeptides of the present invention include keyhole limpet hemacyanin (KLH) tetanus toxoid, pertussis toxin, bovine serum albumin, and ovalbumin. Methods for coupling the polypeptides of the present invention to such macromolecular carriers are disclosed in Harlow et al., ANTIBODIES: A LABORATORY MANUAL, (Cold Spring Harbor Laboratory Press, 2nd ed. 1988).

A composition is said to be "pharmacologically or physiologically acceptable" if its administration can be tolerated by a recipient animal and is otherwise suitable for administration to that animal. Such an agent is said to be administered in a "therapeutically effective amount" if the amount administered is physiologically significant. An agent is physiologically significant if its presence results in a detectable change in the physiology of a recipient patient.

While in all instances the vaccine of the present invention is administered as a pharmacologically acceptable compound, one skilled in the art would recognize that the composition of a pharmacologically acceptable compound varies with the animal to which it is administered. For example, a vaccine intended for human use will generally not be co-administered with Freund's adjuvant. Further, the level of purity of the *S. aureus* polypeptides of the present invention will normally be higher when administered to a human than when administered to a non-human animal.

As would be understood by one of ordinary skill in the art, when the vaccine of the present invention is provided to an animal, it may be in a composition which may contain salts, buffers, adjuvants, or other substances which are desirable for improving the efficacy of the composition. Adjuvants are substances that can be used to specifically augment a specific immune response. These substances generally perform two functions: (1) they protect the antigen(s) from being rapidly catabolized after administration and (2) they nonspecifically stimulate immune responses.

Normally, the adjuvant and the composition are mixed prior to presentation to the

immune system, or presented separately, but into the same site of the animal being immunized. Adjuvants can be loosely divided into several groups based upon their composition. These groups include oil adjuvants (for example, Freund's complete and incomplete), mineral salts (for example, $\text{AlK}(\text{SO}_4)_2$, $\text{AlNa}(\text{SO}_4)_2$, $\text{AlNH}_4(\text{SO}_4)$, silica, kaolin, and carbon), polynucleotides (for example, poly IC and poly AU acids), and certain natural substances (for example, wax D from *Mycobacterium tuberculosis*, as well as substances found in *Corynebacterium parvum*, or *Bordetella pertussis*, and members of the genus *Brucella*. Other substances useful as adjuvants are the saponins such as, for example, Quil A. (Superfos A/S, Denmark). Preferred adjuvants for use in the present invention include aluminum salts, such as $\text{AlK}(\text{SO}_4)_2$, $\text{AlNa}(\text{SO}_4)_2$, and $\text{AlNH}_4(\text{SO}_4)$. Examples of materials suitable for use in vaccine compositions are provided in REMINGTON'S PHARMACEUTICAL SCIENCES 1324-1341 (A. Osol, ed, Mack Publishing Co, Easton, PA, (1980) (incorporated herein by reference).

The therapeutic compositions of the present invention can be administered parenterally by injection, rapid infusion, nasopharyngeal absorption (intranasopharyngeally), dermoabsorption, or orally. The compositions may alternatively be administered intramuscularly, or intravenously. Compositions for parenteral administration include sterile aqueous or non-aqueous solutions, suspensions, and emulsions. Examples of non-aqueous solvents are propylene glycol, polyethylene glycol, vegetable oils such as olive oil, and injectable organic esters such as ethyl oleate. Carriers or occlusive dressings can be used to increase skin permeability and enhance antigen absorption. Liquid dosage forms for oral administration may generally comprise a liposome solution containing the liquid dosage form. Suitable forms for suspending liposomes include emulsions, suspensions, solutions, syrups, and elixirs containing inert diluents commonly used in the art, such as purified water. Besides the inert diluents, such compositions can also include adjuvants, wetting agents, emulsifying and suspending agents, or sweetening, flavoring, or perfuming agents.

Therapeutic compositions of the present invention can also be administered in encapsulated form. For example, intranasal immunization using vaccines encapsulated in biodegradable microsphere composed of poly(DL-lactide-co-glycolide). See, Shahin, R. et al. (1995) Infect. Immun. 63:1195-1200. Similarly, orally administered encapsulated *Salmonella typhimurium* antigens can also be used. Allaoui-Attarki, K. et al. (1997) Infect. Immun. 65:853-857. Encapsulated vaccines of the present invention can be administered by

a variety of routes including those involving contacting the vaccine with mucous membranes (e.g., intranasally, intracolonicly, intraduodenally).

Many different techniques exist for the timing of the immunizations when a multiple administration regimen is utilized. It is possible to use the compositions of the invention more than once to increase the levels and diversities of expression of the immunoglobulin repertoire expressed by the immunized animal. Typically, if multiple immunizations are given, they will be given one to two months apart.

According to the present invention, an "effective amount" of a therapeutic composition is one which is sufficient to achieve a desired biological effect. Generally, the dosage needed to provide an effective amount of the composition will vary depending upon such factors as the animal's or human's age, condition, sex, and extent of disease, if any, and other variables which can be adjusted by one of ordinary skill in the art.

The antigenic preparations of the invention can be administered by either single or multiple dosages of an effective amount. Effective amounts of the compositions of the invention can vary from 0.01-1,000 $\mu\text{g/ml}$ per dose, more preferably 0.1-500 $\mu\text{g/ml}$ per dose, and most preferably 10-300 $\mu\text{g/ml}$ per dose.

TR16 Binding Peptides and Other Molecules

The invention also encompasses screening methods for identifying polypeptides and nonpolypeptides that bind the *S. aureus* polypeptides of the invention, and the *S. aureus* polypeptides binding molecules identified thereby. These binding molecules are useful, for example, as agonists and antagonists of the *S. aureus* polypeptides of the invention. Such agonists and antagonists can be used, in accordance with the invention, in the therapeutic embodiments described in detail, below.

This method comprises the steps of:

- a. contacting a *S. aureus* polypeptide with a plurality of molecules; and
- b. identifying a molecule that binds the *S. aureus* polypeptide.

The step of contacting the *S. aureus* polypeptide with the plurality of molecules may be effected in a number of ways. For example, one may contemplate immobilizing the *S. aureus* polypeptide on a solid support and bringing a solution of the plurality of molecules in contact with the immobilized *S. aureus* polypeptide. Such a procedure would be akin to an

affinity chromatographic process, with the affinity matrix being comprised of the immobilized *S. aureus* polypeptide. The molecules having a selective affinity for the *S. aureus* polypeptide can then be purified by affinity selection. The nature of the solid support, process for attachment of the *S. aureus* polypeptide to the solid support, solvent, and conditions of the affinity isolation or selection are largely conventional and well known to those of ordinary skill in the art.

Alternatively, one may also separate a plurality of polypeptides into substantially separate fractions comprising a subset of or individual polypeptides. For instance, one can separate the plurality of polypeptides by gel electrophoresis, column chromatography, or like method known to those of ordinary skill for the separation of polypeptides. The individual polypeptides can also be produced by a transformed host cell in such a way as to be expressed on or about its outer surface (e.g., a recombinant phage). Individual isolates can then be "probed" by the *S. aureus* polypeptide, optionally in the presence of an inducer should one be required for expression, to determine if any selective affinity interaction takes place between the *S. aureus* polypeptide and the individual clone. Prior to contacting the *S. aureus* polypeptide with each fraction comprising individual polypeptides, the polypeptides could first be transferred to a solid support for additional convenience. Such a solid support may simply be a piece of filter membrane, such as one made of nitrocellulose or nylon. In this manner, positive clones could be identified from a collection of transformed host cells of an expression library, which harbor a DNA construct encoding a polypeptide having a selective affinity for *S. aureus* polypeptide. Furthermore, the amino acid sequence of the polypeptide having a selective affinity for any one of the *S. aureus* polypeptides of the invention can be determined directly by conventional means or the coding sequence of the DNA encoding the polypeptide can frequently be determined more conveniently. The primary sequence can then be deduced from the corresponding DNA sequence. If the amino acid sequence is to be determined from the polypeptide itself, one may use microsequencing techniques. The sequencing technique may include mass spectroscopy.

In certain situations, it may be desirable to wash away any unbound *S. aureus* polypeptide, or alternatively, unbound polypeptides, from a mixture of the *S. aureus* polypeptide and the plurality of polypeptides prior to attempting to determine or to detect the presence of a selective affinity interaction. Such a wash step may be particularly desirable when the *S. aureus* polypeptide or the plurality of polypeptides is bound to a solid support.

The plurality of molecules provided according to this method may be provided by way of diversity libraries, such as random or combinatorial peptide or nonpeptide libraries which can be screened for molecules that specifically bind to a *S. aureus* polypeptide. Many libraries are known in the art that can be used, e.g., chemically synthesized libraries, recombinant (e.g., phage display libraries), and in vitro translation-based libraries. Examples of chemically synthesized libraries are described in Fodor et al., 1991, Science 251:767-773; Houghten et al., 1991, Nature 354:84-86; Lam et al., 1991, Nature 354:82-84; Medynski, 1994, Bio/Technology 12:709-710; Gallop et al., 1994, J. Medicinal Chemistry 37(9):1233-1251; Ohlmeyer et al., 1993, Proc. Natl. Acad. Sci. USA 90:10922-10926; Erb et al., 1994, Proc. Natl. Acad. Sci. USA 91:11422-11426; Houghten et al., 1992, Biotechniques 13:412; Jayawickreme et al., 1994, Proc. Natl. Acad. Sci. USA 91:1614-1618; Salmon et al., 1993, Proc. Natl. Acad. Sci. USA 90:11708-11712; PCT Publication No. WO 93/20242; and Brenner and Lerner, 1992, Proc. Natl. Acad. Sci. USA 89:5381-5383.

Examples of phage display libraries are described in Scott and Smith, 1990, Science 249:386-390; Devlin et al., 1990, Science, 249:404-406; Christian, R. B., et al., 1992, J. Mol. Biol. 227:711-718; Lenstra, 1992, J. Immunol. Meth. 152:149-157; Kay et al., 1993, Gene 128:59-65; and PCT Publication No. WO 94/18318 dated Aug. 18, 1994.

In vitro translation-based libraries include but are not limited to those described in PCT Publication No. WO 91/05058 dated Apr. 18, 1991; and Mattheakis et al., 1994, Proc. Natl. Acad. Sci. USA 91:9022-9026.

By way of examples of nonpeptide libraries, a benzodiazepine library (see e.g., Bunin et al., 1994, Proc. Natl. Acad. Sci. USA 91:4708-4712) can be adapted for use. Peptoid libraries (Simon et al., 1992, Proc. Natl. Acad. Sci. USA 89:9367-9371) can also be used. Another example of a library that can be used, in which the amide functionalities in peptides have been permethylated to generate a chemically transformed combinatorial library, is described by Ostresh et al. (1994, Proc. Natl. Acad. Sci. USA 91:11138-11142).

The variety of non-peptide libraries that are useful in the present invention is great. For example, Ecker and Crooke, 1995, Bio/Technology 13:351-360 list benzodiazepines, hydantoins, piperazinediones, biphenyls, sugar analogs, beta-mercaptoketones, arylacetic acids, acylpiperidines, benzopyrans, cubanes, xanthines, aminimides, and oxazolones as among the chemical species that form the basis of various libraries.

Non-peptide libraries can be classified broadly into two types: decorated monomers

and oligomers. Decorated monomer libraries employ a relatively simple scaffold structure upon which a variety functional groups is added. Often the scaffold will be a molecule with a known useful pharmacological activity. For example, the scaffold might be the benzodiazepine structure.

5 Non-peptide oligomer libraries utilize a large number of monomers that are assembled together in ways that create new shapes that depend on the order of the monomers. Among the monomer units that have been used are carbamates, pyrrolinones, and morpholinos. Peptoids, peptide-like oligomers in which the side chain is attached to the alpha amino group rather than the alpha carbon, form the basis of another version of non-peptide oligomer
10 libraries. The first non-peptide oligomer libraries utilized a single type of monomer and thus contained a repeating backbone. Recent libraries have utilized more than one monomer, giving the libraries added flexibility.

Screening the libraries can be accomplished by any of a variety of commonly known methods. See, e.g., the following references, which disclose screening of peptide libraries:
15 Parmley and Smith, 1989, Adv. Exp. Med. Biol. 251:215-218; Scott and Smith, 1990, Science 249:386-390; Fowlkes et al., 1992, BioTechniques 13:422-427; Oldenburg et al., 1992, Proc. Natl. Acad. Sci. USA 89:5393-5397; Yu et al., 1994, Cell 76:933-945; Staudt et al., 1988, Science 241:577-580; Bock et al., 1992, Nature 355:564-566; Tuerk et al., 1992, Proc. Natl. Acad. Sci. USA 89:6988-6992; Ellington et al., 1992, Nature 355:850-852; U.S.
20 Pat. No. 5,096,815, U.S. Pat. No. 5,223,409, and U.S. Pat. No. 5,198,346, all to Ladner et al.; Rebar and Pabo, 1993, Science 263:671-673; and CT Publication No. WO 94/18318.

In a specific embodiment, screening to identify a molecule that binds a *S. aureus* polypeptide can be carried out by contacting the library members with a *S. aureus* polypeptide immobilized on a solid phase and harvesting those library members that bind to
25 the *S. aureus* polypeptide. Examples of such screening methods, termed "panning" techniques are described by way of example in Parmley and Smith, 1988, Gene 73:305-318; Fowlkes et al., 1992, BioTechniques 13:422-427; PCT Publication No. WO 94/18318; and in references cited herein.

In another embodiment, the two-hybrid system for selecting interacting proteins in
30 yeast (Fields and Song, 1989, Nature 340:245-246; Chien et al., 1991, Proc. Natl. Acad. Sci. USA 88:9578-9582) can be used to identify molecules that specifically bind to any one of the *S. aureus* polypeptides shown in Table 1.

Where the *S. aureus* polypeptide binding molecule is a polypeptide, the polypeptide can be conveniently selected from any peptide library, including random peptide libraries, combinatorial peptide libraries, or biased peptide libraries. The term "biased" is used herein to mean that the method of generating the library is manipulated so as to restrict one or more parameters that govern the diversity of the resulting collection of molecules, in this case peptides.

Thus, a truly random peptide library would generate a collection of peptides in which the probability of finding a particular amino acid at a given position of the peptide is the same for all 20 amino acids. A bias can be introduced into the library, however, by specifying, for example, that a lysine occur every fifth amino acid or that positions 4, 8, and 9 of a decapeptide library be fixed to include only arginine. Clearly, many types of biases can be contemplated, and the present invention is not restricted to any particular bias. Furthermore, the present invention contemplates specific types of peptide libraries, such as phage displayed peptide libraries and those that utilize a DNA construct comprising a lambda phage vector with a DNA insert.

As mentioned above, in the case of a *S. aureus* polypeptide binding molecule that is a polypeptide, the polypeptide may have about 6 to less than about 60 amino acid residues, preferably about 6 to about 10 amino acid residues, and most preferably, about 6 to about 22 amino acids. In another embodiment, a *S. aureus* polypeptide binding polypeptide has in the range of 15-100 amino acids, or 20-50 amino acids.

The selected *S. aureus* polypeptide binding polypeptide can be obtained by chemical synthesis or recombinant expression.

Examples

Example 1: Isolation of a Selected DNA Clone From the Deposited Sample

Three approaches can be used to isolate a *S. aureus* clone comprising a polynucleotide of the present invention from any *S. aureus* genomic DNA library. The *S. aureus* strain ISP3 has been deposited as a convenient source for obtaining a *S. aureus* strain although a wide variety of strains *S. aureus* strains can be used which are known in the art.

S. aureus genomic DNA is prepared using the following method. A 20ml overnight bacterial culture grown in a rich medium (e.g., Trypticase Soy Broth, Brain Heart Infusion broth or Super broth), pelleted, washed two times with TES (30mM Tris-pH 8.0, 25mM EDTA, 50mM NaCl), and resuspended in 5ml high salt TES (2.5M NaCl). Lysostaphin is added to final concentration of approx 50ug/ml and the mixture is rotated slowly 1 hour at 37C to make protoplast cells. The solution is then placed in incubator (or place in a shaking water bath) and warmed to 55C. Five hundred micro liter of 20% sarcosyl in TES (final concentration 2%) is then added to lyse the cells. Next, guanidine HCl is added to a final concentration of 7M (3.69g in 5.5 ml). The mixture is swirled slowly at 55C for 60-90 min (solution should clear). A CsCl gradient is then set up in SW41 ultra clear tubes using 2.0ml 5.7M CsCl and overlaying with 2.85M CsCl. The gradient is carefully overlayed with the DNA-containing GuHCl solution. The gradient is spun at 30,000 rpm, 20C for 24 hr and the lower DNA band is collected. The volume is increased to 5 ml with TE buffer. The DNA is then treated with protease K (10 ug/ml) overnight at 37 C, and precipitated with ethanol. The precipitated DNA is resuspended in a desired buffer.

In the first method, a plasmid is directly isolated by screening a plasmid *S. aureus* genomic DNA library using a polynucleotide probe corresponding to a polynucleotide of the present invention. Particularly, a specific polynucleotide with 30-40 nucleotides is synthesized using an Applied Biosystems DNA synthesizer according to the sequence reported. The oligonucleotide is labeled, for instance, with ³²P-γ-ATP using T4 polynucleotide kinase and purified according to routine methods. (See, e.g., Maniatis et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Press, Cold Spring, NY (1982).) The library is transformed into a suitable host, as indicated above (such as XL-1 Blue (Stratagene)) using techniques known to those of skill in the art. See, e.g., Sambrook et al. MOLECULAR CLONING: A LABORATORY MANUAL (Cold Spring Harbor, N.Y. 2nd ed. 1989); Ausubel et al., CURRENT PROTOCOLS IN MOLECULAR BIOLOGY (John Wiley and Sons, N.Y. 1989). The transformants are plated on 1.5% agar plates (containing the appropriate selection agent, e.g., ampicillin) to a density of about 150 transformants (colonies) per plate. These plates are screened using Nylon membranes according to routine methods for bacterial colony screening. See, e.g., Sambrook et al. MOLECULAR CLONING: A LABORATORY MANUAL (Cold Spring Harbor, N.Y. 2nd ed. 1989); Ausubel et al., CURRENT PROTOCOLS IN MOLECULAR BIOLOGY (John

Wiley and Sons, N.Y. 1989) or other techniques known to those of skill in the art.

Alternatively, two primers of 15-25 nucleotides derived from the 5' and 3' ends of a polynucleotide of Table 1 are synthesized and used to amplify the desired DNA by PCR using a *S. aureus* genomic DNA prep (e.g., the deposited *S. aureus* ISP3) as a template. PCR is carried out under routine conditions, for instance, in 25 µl of reaction mixture with 0.5 µg of the above DNA template. A convenient reaction mixture is 1.5-5 mM MgCl₂, 0.01% (w/v) gelatin, 20 µM each of dATP, dCTP, dGTP, dTTP, 25 pmol of each primer and 0.25 Unit of Taq polymerase. Thirty five cycles of PCR (denaturation at 94°C for 1 min; annealing at 55°C for 1 min; elongation at 72°C for 1 min) are performed with a Perkin-Elmer Cetus automated thermal cycler. The amplified product is analyzed by agarose gel electrophoresis and the DNA band with expected molecular weight is excised and purified. The PCR product is verified to be the selected sequence by subcloning and sequencing the DNA product.

Finally, overlapping oligos of the DNA sequences of Table 1 can be synthesized and used to generate a nucleotide sequence of desired length using PCR methods known in the art.

Example 2(a): Expression and Purification staphylococcal polypeptides in *E. coli*

The bacterial expression vector pQE60 is used for bacterial expression in this example. (QIAGEN, Inc., 9259 Eton Avenue, Chatsworth, CA, 91311). pQE60 encodes ampicillin antibiotic resistance ("Ampr") and contains a bacterial origin of replication ("ori"), an IPTG inducible promoter, a ribosome binding site ("RBS"), six codons encoding histidine residues that allow affinity purification using nickel-nitrilo-tri-acetic acid ("Ni-NTA") affinity resin (QIAGEN, Inc., *supra*) and suitable single restriction enzyme cleavage sites. These elements are arranged such that an inserted DNA fragment encoding a polypeptide expresses that polypeptide with the six His residues (i.e., a "6 X His tag") covalently linked to the carboxyl terminus of that polypeptide.

The DNA sequence encoding the desired portion of a *S. aureus* protein of the present invention is amplified from *S. aureus* genomic DNA or from the deposited DNA clone using PCR oligonucleotide primers which anneal to the 5' and 3' sequences coding for the portion of the *S. aureus* polynucleotide. Additional nucleotides containing restriction sites to facilitate cloning in the pQE60 vector are added to the 5' and 3' sequences, respectively.

For cloning the mature protein, the 5' primer has a sequence containing an appropriate

restriction site followed by nucleotides of the amino terminal coding sequence of the desired *S. aureus* polynucleotide sequence in Table 1. One of ordinary skill in the art would appreciate that the point in the protein coding sequence where the 5' and 3' primers begin may be varied to amplify a DNA segment encoding any desired portion of the complete protein shorter or longer than the mature form. The 3' primer has a sequence containing an appropriate restriction site followed by nucleotides complementary to the 3' end of the desired coding sequence of Table 1, excluding a stop codon, with the coding sequence aligned with the restriction site so as to maintain its reading frame with that of the six His codons in the pQE60 vector.

The amplified *S. aureus* DNA fragment and the vector pQE60 are digested with restriction enzymes which recognize the sites in the primers and the digested DNAs are then ligated together. The *S. aureus* DNA is inserted into the restricted pQE60 vector in a manner which places the *S. aureus* protein coding region downstream from the IPTG-inducible promoter and in-frame with an initiating AUG and the six histidine codons.

The ligation mixture is transformed into competent *E. coli* cells using standard procedures such as those described by Sambrook et al., *supra*. *E. coli* strain M15/rep4, containing multiple copies of the plasmid pREP4, which expresses the lac repressor and confers kanamycin resistance ("Kanr"), is used in carrying out the illustrative example described herein. This strain, which is only one of many that are suitable for expressing a *S. aureus* polypeptide, is available commercially (QIAGEN, Inc., *supra*). Transformants are identified by their ability to grow on LB plates in the presence of ampicillin and kanamycin. Plasmid DNA is isolated from resistant colonies and the identity of the cloned DNA confirmed by restriction analysis, PCR and DNA sequencing.

Clones containing the desired constructs are grown overnight ("O/N") in liquid culture in LB media supplemented with both ampicillin (100 µg/ml) and kanamycin (25 µg/ml). The O/N culture is used to inoculate a large culture, at a dilution of approximately 1:25 to 1:250. The cells are grown to an optical density at 600 nm ("OD600") of between 0.4 and 0.6. Isopropyl-β-D-thiogalactopyranoside ("IPTG") is then added to a final concentration of 1 mM to induce transcription from the lac repressor sensitive promoter, by inactivating the lacI repressor. Cells subsequently are incubated further for 3 to 4 hours. Cells then are harvested by centrifugation.

The cells are then stirred for 3-4 hours at 4°C in 6M guanidine-HCl, pH 8. The cell

debris is removed by centrifugation, and the supernatant containing the *S. aureus* polypeptide is loaded onto a nickel-nitrilo-tri-acetic acid ("Ni-NTA") affinity resin column (QIAGEN, Inc., *supra*). Proteins with a 6 x His tag bind to the Ni-NTA resin with high affinity and can be purified in a simple one-step procedure (for details see: The
5 QIAexpressionist, 1995, QIAGEN, Inc., *supra*). Briefly the supernatant is loaded onto the column in 6 M guanidine-HCl, pH 8, the column is first washed with 10 volumes of 6 M guanidine-HCl, pH 8, then washed with 10 volumes of 6 M guanidine-HCl pH 6, and finally the *S. aureus* polypeptide is eluted with 6 M guanidine-HCl, pH 5.

The purified protein is then renatured by dialyzing it against phosphate-buffered
10 saline (PBS) or 50 mM Na-acetate, pH 6 buffer plus 200 mM NaCl. Alternatively, the protein can be successfully refolded while immobilized on the Ni-NTA column. The recommended conditions are as follows: renature using a linear 6M-1M urea gradient in 500 mM NaCl, 20% glycerol, 20 mM Tris/HCl pH 7.4, containing protease inhibitors. The renaturation should be performed over a period of 1.5 hours or more. After renaturation the
15 proteins can be eluted by the addition of 250 mM imidazole. Imidazole is removed by a final dialyzing step against PBS or 50 mM sodium acetate pH 6 buffer plus 200 mM NaCl. The purified protein is stored at 4° C or frozen at -80° C.

Alternatively, the polypeptides of the present invention can be produced by a non-denaturing method. In this method, after the cells are harvested by centrifugation, the cell
20 pellet from each liter of culture is resuspended in 25 ml of Lysis Buffer A at 4°C (Lysis Buffer A = 50 mM Na-phosphate, 300 mM NaCl, 10 mM 2-mercaptoethanol, 10% Glycerol, pH 7.5 with 1 tablet of Complete EDTA-free protease inhibitor cocktail (Boehringer Mannheim #1873580) per 50 ml of buffer). Absorbance at 550 nm is approximately 10-20 O.D./ml. The suspension is then put through three freeze/thaw cycles from -70°C (using a
25 ethanol-dry ice bath) up to room temperature. The cells are lysed via sonication in short 10 sec bursts over 3 minutes at approximately 80W while kept on ice. The sonicated sample is then centrifuged at 15,000 RPM for 30 minutes at 4°C. The supernatant is passed through a column containing 1.0 ml of CL-4B resin to pre-clear the sample of any proteins that may bind to agarose non-specifically, and the flow-through fraction is collected.

30 The pre-cleared flow-through is applied to a nickel-nitrilo-tri-acetic acid ("Ni-NTA") affinity resin column (Quiagen, Inc., *supra*). Proteins with a 6 X His tag bind to the Ni-NTA resin with high affinity and can be purified in a simple one-step procedure. Briefly, the

supernatant is loaded onto the column in Lysis Buffer A at 4°C, the column is first washed with 10 volumes of Lysis Buffer A until the A280 of the eluate returns to the baseline. Then, the column is washed with 5 volumes of 40 mM Imidazole (92% Lysis Buffer A / 8% Buffer B) (Buffer B = 50 mM Na-Phosphate, 300 mM NaCl, 10% Glycerol, 10 mM 2-mercaptoethanol, 500 mM Imidazole, pH of the final buffer should be 7.5). The protein is eluted off of the column with a series of increasing Imidazole solutions made by adjusting the ratios of Lysis Buffer A to Buffer B. Three different concentrations are used: 3 volumes of 75 mM Imidazole, 3 volumes of 150 mM Imidazole, 5 volumes of 500 mM Imidazole. The fractions containing the purified protein are analyzed using 8 %, 10 % or 14% SDS-PAGE depending on the protein size. The purified protein is then dialyzed 2X against phosphate-buffered saline (PBS) in order to place it into an easily workable buffer. The purified protein is stored at 4°C or frozen at -80°

The following is another alternative method may be used to purify *S. aureus* expressed in *E. coli* when it is present in the form of inclusion bodies. Unless otherwise specified, all of the following steps are conducted at 4-10°C.

Upon completion of the production phase of the *E. coli* fermentation, the cell culture is cooled to 4-10°C and the cells are harvested by continuous centrifugation at 15,000 rpm (Heraeus Sepatech). On the basis of the expected yield of protein per unit weight of cell paste and the amount of purified protein required, an appropriate amount of cell paste, by weight, is suspended in a buffer solution containing 100 mM Tris, 50 mM EDTA, pH 7.4. The cells are dispersed to a homogeneous suspension using a high shear mixer.

The cells are then lysed by passing the solution through a microfluidizer (Microfluidics, Corp. or APV Gaulin, Inc.) twice at 4000-6000 psi. The homogenate is then mixed with NaCl solution to a final concentration of 0.5 M NaCl, followed by centrifugation at 7000 x g for 15 min. The resultant pellet is washed again using 0.5M NaCl, 100 mM Tris, 50 mM EDTA, pH 7.4.

The resulting washed inclusion bodies are solubilized with 1.5 M guanidine hydrochloride (GuHCl) for 2-4 hours. After 7000 x g centrifugation for 15 min., the pellet is discarded and the *S. aureus* polypeptide-containing supernatant is incubated at 4°C overnight to allow further GuHCl extraction.

Following high speed centrifugation (30,000 x g) to remove insoluble particles, the GuHCl solubilized protein is refolded by quickly mixing the GuHCl extract with 20 volumes

of buffer containing 50 mM sodium, pH 4.5, 150 mM NaCl, 2 mM EDTA by vigorous stirring. The refolded diluted protein solution is kept at 4°C without mixing for 12 hours prior to further purification steps.

To clarify the refolded *S. aureus* polypeptide solution, a previously prepared tangential filtration unit equipped with 0.16 µm membrane filter with appropriate surface area (e.g., Filtron), equilibrated with 40 mM sodium acetate, pH 6.0 is employed. The filtered sample is loaded onto a cation exchange resin (e.g., Poros HS-50, Perseptive Biosystems). The column is washed with 40 mM sodium acetate, pH 6.0 and eluted with 250 mM, 500 mM, 1000 mM, and 1500 mM NaCl in the same buffer, in a stepwise manner. The absorbance at 280 nm of the effluent is continuously monitored. Fractions are collected and further analyzed by SDS-PAGE.

Fractions containing the *S. aureus* polypeptide are then pooled and mixed with 4 volumes of water. The diluted sample is then loaded onto a previously prepared set of tandem columns of strong anion (Poros HQ-50, Perseptive Biosystems) and weak anion (Poros CM-20, Perseptive Biosystems) exchange resins. The columns are equilibrated with 40 mM sodium acetate, pH 6.0. Both columns are washed with 40 mM sodium acetate, pH 6.0, 200 mM NaCl. The CM-20 column is then eluted using a 10 column volume linear gradient ranging from 0.2 M NaCl, 50 mM sodium acetate, pH 6.0 to 1.0 M NaCl, 50 mM sodium acetate, pH 6.5. Fractions are collected under constant A₂₈₀ monitoring of the effluent. Fractions containing the *S. aureus* polypeptide (determined, for instance, by 16% SDS-PAGE) are then pooled.

The resultant *S. aureus* polypeptide exhibits greater than 95% purity after the above refolding and purification steps. No major contaminant bands are observed from Commassie blue stained 16% SDS-PAGE gel when 5 µg of purified protein is loaded. The purified protein is also tested for endotoxin/LPS contamination, and typically the LPS content is less than 0.1 ng/ml according to LAL assays.

Example 2(b): Expression and Purification staphylococcal polypeptides in E. coli

Alternatively, the vector pQE10 can be used to clone and express polypeptides of the present invention. The difference being such that an inserted DNA fragment encoding a polypeptide expresses that polypeptide with the six His residues (i.e., a "6 X His tag") covalently linked to the amino terminus of that polypeptide. The bacterial expression vector

pQE10 (QIAGEN, Inc., 9259 Eton Avenue, Chatsworth, CA, 91311) is used in this example. The components of the pQE10 plasmid are arranged such that the inserted DNA sequence encoding a polypeptide of the present invention expresses the polypeptide with the six His residues (*i.e.*, a "6 X His tag")) covalently linked to the amino terminus.

5 The DNA sequences encoding the desired portions of a polypeptide of Table 1 are amplified using PCR oligonucleotide primers from either genomic *S. aureus* DNA or DNA from the plasmid clones listed in Table 1 clones of the present invention. The PCR primers anneal to the nucleotide sequences encoding the desired amino acid sequence of a polypeptide of the present invention. Additional nucleotides containing restriction sites to
10 facilitate cloning in the pQE10 vector are added to the 5' and 3' primer sequences, respectively.

For cloning a polypeptide of the present invention, the 5' and 3' primers are selected to amplify their respective nucleotide coding sequences. One of ordinary skill in the art would appreciate that the point in the protein coding sequence where the 5' and 3' primers begins
15 may be varied to amplify a DNA segment encoding any desired portion of a polypeptide of the present invention. The 5' primer is designed so the coding sequence of the 6 X His tag is aligned with the restriction site so as to maintain its reading frame with that of *S. aureus* polypeptide. The 3' is designed to include an stop codon. The amplified DNA fragment is then cloned, and the protein expressed, as described above for the pQE60 plasmid.

20 The DNA sequences encoding the amino acid sequences of Table 1 may also be cloned and expressed as fusion proteins by a protocol similar to that described directly above, wherein the pET-32b(+) vector (Novagen, 601 Science Drive, Madison, WI 53711) is preferentially used in place of pQE10.

25 **Example 2(c): Expression and Purification of *Staphylococcus* polypeptides in *E. coli***

The bacterial expression vector pQE60 is used for bacterial expression in this example (QIAGEN, Inc., 9259 Eton Avenue, Chatsworth, CA, 91311). However, in this example, the polypeptide coding sequence is inserted such that translation of the six His codons is prevented and, therefore, the polypeptide is produced with no 6 X His tag.

30 The DNA sequence encoding the desired portion of the *S. aureus* amino acid sequence is amplified from a *S. aureus* genomic DNA prep using PCR oligonucleotide primers which anneal to the 5' and 3' nucleotide sequences corresponding to the desired

portion of the *S. aureus* polypeptides. Additional nucleotides containing restriction sites to facilitate cloning in the pQE60 vector are added to the 5' and 3' primer sequences.

For cloning a *S. aureus* polypeptides of the present invention, 5' and 3' primers are selected to amplify their respective nucleotide coding sequences. One of ordinary skill in the art would appreciate that the point in the protein coding sequence where the 5' and 3' primers begin may be varied to amplify a DNA segment encoding any desired portion of a polypeptide of the present invention. The 3' and 5' primers contain appropriate restriction sites followed by nucleotides complementary to the 5' and 3' ends of the coding sequence respectively. The 3' primer is additionally designed to include an in-frame stop codon.

The amplified *S. aureus* DNA fragments and the vector pQE60 are digested with restriction enzymes recognizing the sites in the primers and the digested DNAs are then ligated together. Insertion of the *S. aureus* DNA into the restricted pQE60 vector places the *S. aureus* protein coding region including its associated stop codon downstream from the IPTG-inducible promoter and in-frame with an initiating AUG. The associated stop codon prevents translation of the six histidine codons downstream of the insertion point.

The ligation mixture is transformed into competent *E. coli* cells using standard procedures such as those described by Sambrook et al. *E. coli* strain M15/rep4, containing multiple copies of the plasmid pREP4, which expresses the lac repressor and confers kanamycin resistance ("Kanr"), is used in carrying out the illustrative example described herein. This strain, which is only one of many that are suitable for expressing *S. aureus* polypeptide, is available commercially (QIAGEN, Inc., *supra*). Transformants are identified by their ability to grow on LB plates in the presence of ampicillin and kanamycin. Plasmid DNA is isolated from resistant colonies and the identity of the cloned DNA confirmed by restriction analysis, PCR and DNA sequencing.

Clones containing the desired constructs are grown overnight ("O/N") in liquid culture in LB media supplemented with both ampicillin (100 µg/ml) and kanamycin (25 µg/ml). The O/N culture is used to inoculate a large culture, at a dilution of approximately 1:25 to 1:250. The cells are grown to an optical density at 600 nm ("OD600") of between 0.4 and 0.6. isopropyl-b-D-thiogalactopyranoside ("IPTG") is then added to a final concentration of 1 mM to induce transcription from the *lac* repressor sensitive promoter, by inactivating the *lacI* repressor. Cells subsequently are incubated further for 3 to 4 hours. Cells then are harvested by centrifugation.

To purify the *S. aureus* polypeptide, the cells are then stirred for 3-4 hours at 4°C in 6M guanidine-HCl, pH 8. The cell debris is removed by centrifugation, and the supernatant containing the *S. aureus* polypeptide is dialyzed against 50 mM Na-acetate buffer pH 6, supplemented with 200 mM NaCl. Alternatively, the protein can be successfully refolded by dialyzing it against 500 mM NaCl, 20% glycerol, 25 mM Tris/HCl pH 7.4, containing protease inhibitors. After renaturation the protein can be purified by ion exchange, hydrophobic interaction and size exclusion chromatography. Alternatively, an affinity chromatography step such as an antibody column can be used to obtain pure *S. aureus* polypeptide. The purified protein is stored at 4°C or frozen at -80°C.

The following alternative method may be used to purify *S. aureus* polypeptides expressed in *E. coli* when it is present in the form of inclusion bodies. Unless otherwise specified, all of the following steps are conducted at 4-10°C.

Upon completion of the production phase of the *E. coli* fermentation, the cell culture is cooled to 4-10°C and the cells are harvested by continuous centrifugation at 15,000 rpm (Heraeus Sepatech). On the basis of the expected yield of protein per unit weight of cell paste and the amount of purified protein required, an appropriate amount of cell paste, by weight, is suspended in a buffer solution containing 100 mM Tris, 50 mM EDTA, pH 7.4. The cells are dispersed to a homogeneous suspension using a high shear mixer.

The cells were then lysed by passing the solution through a microfluidizer (Microfluidics, Corp. or APV Gaulin, Inc.) twice at 4000-6000 psi. The homogenate is then mixed with NaCl solution to a final concentration of 0.5 M NaCl, followed by centrifugation at 7000 x g for 15 min. The resultant pellet is washed again using 0.5M NaCl, 100 mM Tris, 50 mM EDTA, pH 7.4.

The resulting washed inclusion bodies are solubilized with 1.5 M guanidine hydrochloride (GuHCl) for 2-4 hours. After 7000 x g centrifugation for 15 min., the pellet is discarded and the *S. aureus* polypeptide-containing supernatant is incubated at 4°C overnight to allow further GuHCl extraction.

Following high speed centrifugation (30,000 x g) to remove insoluble particles, the GuHCl solubilized protein is refolded by quickly mixing the GuHCl extract with 20 volumes of buffer containing 50 mM sodium, pH 4.5, 150 mM NaCl, 2 mM EDTA by vigorous stirring. The refolded diluted protein solution is kept at 4°C without mixing for 12 hours

prior to further purification steps.

To clarify the refolded *S. aureus* polypeptide solution, a previously prepared tangential filtration unit equipped with 0.16 μm membrane filter with appropriate surface area (e.g., Filtron), equilibrated with 40 mM sodium acetate, pH 6.0 is employed. The filtered sample is loaded onto a cation exchange resin (e.g., Poros HS-50, Perseptive Biosystems). The column is washed with 40 mM sodium acetate, pH 6.0 and eluted with 250 mM, 500 mM, 1000 mM, and 1500 mM NaCl in the same buffer, in a stepwise manner. The absorbance at 280 nm of the effluent is continuously monitored. Fractions are collected and further analyzed by SDS-PAGE.

Fractions containing the *S. aureus* polypeptide are then pooled and mixed with 4 volumes of water. The diluted sample is then loaded onto a previously prepared set of tandem columns of strong anion (Poros HQ-50, Perseptive Biosystems) and weak anion (Poros CM-20, Perseptive Biosystems) exchange resins. The columns are equilibrated with 40 mM sodium acetate, pH 6.0. Both columns are washed with 40 mM sodium acetate, pH 6.0, 200 mM NaCl. The CM-20 column is then eluted using a 10 column volume linear gradient ranging from 0.2 M NaCl, 50 mM sodium acetate, pH 6.0 to 1.0 M NaCl, 50 mM sodium acetate, pH 6.5. Fractions are collected under constant A_{280} monitoring of the effluent. Fractions containing the *S. aureus* polypeptide (determined, for instance, by 16% SDS-PAGE) are then pooled.

The resultant *S. aureus* polypeptide exhibits greater than 95% purity after the above refolding and purification steps. No major contaminant bands are observed from Coomassie blue stained 16% SDS-PAGE gel when 5 μg of purified protein is loaded. The purified protein is also tested for endotoxin/LPS contamination, and typically the LPS content is less than 0.1 ng/ml according to LAL assays.

Example 2(d): Cloning and Expression of *S. aureus* in Other Bacteria

S. aureus polypeptides can also be produced in: *S. aureus* using the methods of S. Skinner et al., (1988) Mol. Microbiol. 2:289-297 or J. I. Moreno (1996) Protein Expr. Purif. 8(3):332-340; *Lactobacillus* using the methods of C. Rush et al., 1997 Appl. Microbiol. Biotechnol. 47(5):537-542; or in *Bacillus subtilis* using the methods Chang et al., U.S. Patent No. 4,952,508.

Example 3: Cloning and Expression in COS Cells

A *S. aureus* expression plasmid is made by cloning a portion of the DNA encoding a *S. aureus* polypeptide into the expression vector pDNAI/Amp or pDNAIII (which can be obtained from Invitrogen, Inc.). The expression vector pDNAI/amp contains: (1) an *E. coli* origin of replication effective for propagation in *E. coli* and other prokaryotic cells; (2) an ampicillin resistance gene for selection of plasmid-containing prokaryotic cells; (3) an SV40 origin of replication for propagation in eukaryotic cells; (4) a CMV promoter, a polylinker, an SV40 intron; (5) several codons encoding a hemagglutinin fragment (i.e., an "HA" tag to facilitate purification) followed by a termination codon and polyadenylation signal arranged so that a DNA can be conveniently placed under expression control of the CMV promoter and operably linked to the SV40 intron and the polyadenylation signal by means of restriction sites in the polylinker. The HA tag corresponds to an epitope derived from the influenza hemagglutinin protein described by Wilson et al. 1984 Cell 37:767. The fusion of the HA tag to the target protein allows easy detection and recovery of the recombinant protein with an antibody that recognizes the HA epitope. pDNAIII contains, in addition, the selectable neomycin marker.

A DNA fragment encoding a *S. aureus* polypeptide is cloned into the polylinker region of the vector so that recombinant protein expression is directed by the CMV promoter. The plasmid construction strategy is as follows. The DNA from a *S. aureus* genomic DNA prep is amplified using primers that contain convenient restriction sites, much as described above for construction of vectors for expression of *S. aureus* in *E. coli*. The 5' primer contains a Kozak sequence, an AUG start codon, and nucleotides of the 5' coding region of the *S. aureus* polypeptide. The 3' primer, contains nucleotides complementary to the 3' coding sequence of the *S. aureus* DNA, a stop codon, and a convenient restriction site.

The PCR amplified DNA fragment and the vector, pDNAI/Amp, are digested with appropriate restriction enzymes and then ligated. The ligation mixture is transformed into an appropriate *E. coli* strain such as SURE™ (Stratagene Cloning Systems, La Jolla, CA 92037), and the transformed culture is plated on ampicillin media plates which then are incubated to allow growth of ampicillin resistant colonies. Plasmid DNA is isolated from resistant colonies and examined by restriction analysis or other means for the presence of the fragment encoding the *S. aureus* polypeptide.

For expression of a recombinant *S. aureus* polypeptide, COS cells are transfected with

an expression vector, as described above, using DEAE-dextran, as described, for instance, by Sambrook et al. (*supra*). Cells are incubated under conditions for expression of *S. aureus* by the vector.

Expression of the *S. aureus*-HA fusion protein is detected by radiolabeling and immunoprecipitation, using methods described in, for example Harlow et al., *supra*. To this end, two days after transfection, the cells are labeled by incubation in media containing ³⁵S-cysteine for 8 hours. The cells and the media are collected, and the cells are washed and the lysed with detergent-containing RIPA buffer: 150 mM NaCl, 1% NP-40, 0.1% SDS, 1% NP-40, 0.5% DOC, 50 mM TRIS, pH 7.5, as described by Wilson et al. (*supra*). Proteins are precipitated from the cell lysate and from the culture media using an HA-specific monoclonal antibody. The precipitated proteins then are analyzed by SDS-PAGE and autoradiography. An expression product of the expected size is seen in the cell lysate, which is not seen in negative controls.

15 *Example 4: Cloning and Expression in CHO Cells*

The vector pC4 is used for the expression of *S. aureus* polypeptide in this example. Plasmid pC4 is a derivative of the plasmid pSV2-dhfr (ATCC Accession No. 37146). The plasmid contains the mouse DHFR gene under control of the SV40 early promoter. Chinese hamster ovary cells or other cells lacking dihydrofolate activity that are transfected with these plasmids can be selected by growing the cells in a selective medium (alpha minus MEM, Life Technologies) supplemented with the chemotherapeutic agent methotrexate. The amplification of the DHFR genes in cells resistant to methotrexate (MTX) has been well documented. See, e.g., Alt et al., 1978, J. Biol. Chem. 253:1357-1370; Hamlin et al., 1990, Biochem. et Biophys. Acta, 1097:107-143; Page et al., 1991, Biotechnology 9:64-68. Cells grown in increasing concentrations of MTX develop resistance to the drug by overproducing the target enzyme, DHFR, as a result of amplification of the DHFR gene. If a second gene is linked to the DHFR gene, it is usually co-amplified and over-expressed. It is known in the art that this approach may be used to develop cell lines carrying more than 1,000 copies of the amplified gene(s). Subsequently, when the methotrexate is withdrawn, cell lines are obtained which contain the amplified gene integrated into one or more chromosome(s) of the host cell.

Plasmid pC4 contains the strong promoter of the long terminal repeat (LTR) of the Rouse Sarcoma Virus, for expressing a polypeptide of interest, Cullen, et al. (1985) Mol.

Cell. Biol. 5:438-447; plus a fragment isolated from the enhancer of the immediate-early gene of human cytomegalovirus (CMV), Boshart, et al., 1985, Cell 41:521-530. Downstream of the promoter are the following single restriction enzyme cleavage sites that allow the integration of the genes: *Bam* HI, *Xba* I, and *Asp* 718. Behind these cloning sites the plasmid contains the 3' intron and polyadenylation site of the rat preproinsulin gene. Other high efficiency promoters can also be used for the expression, e.g., the human β -actin promoter, the SV40 early or late promoters or the long terminal repeats from other retroviruses, e.g., HIV and HTLV1. Clontech's Tet-Off and Tet-On gene expression systems and similar systems can be used to express the *S. aureus* polypeptide in a regulated way in mammalian cells (Gossen et al., 1992, Proc. Natl. Acad. Sci. USA 89:5547-5551. For the polyadenylation of the mRNA other signals, e.g., from the human growth hormone or globin genes can be used as well. Stable cell lines carrying a gene of interest integrated into the chromosomes can also be selected upon co-transfection with a selectable marker such as gpt, G418 or hygromycin. It is advantageous to use more than one selectable marker in the beginning, e.g., G418 plus methotrexate.

The plasmid pC4 is digested with the restriction enzymes and then dephosphorylated using calf intestinal phosphates by procedures known in the art. The vector is then isolated from a 1% agarose gel. The DNA sequence encoding the *S. aureus* polypeptide is amplified using PCR oligonucleotide primers corresponding to the 5' and 3' sequences of the desired portion of the gene. A 5' primer containing a restriction site, a Kozak sequence, an AUG start codon, and nucleotides of the 5' coding region of the *S. aureus* polypeptide is synthesized and used. A 3' primer, containing a restriction site, stop codon, and nucleotides complementary to the 3' coding sequence of the *S. aureus* polypeptides is synthesized and used. The amplified fragment is digested with the restriction endonucleases and then purified again on a 1% agarose gel. The isolated fragment and the dephosphorylated vector are then ligated with T4 DNA ligase. *E. coli* HB101 or XL-1 Blue cells are then transformed and bacteria are identified that contain the fragment inserted into plasmid pC4 using, for instance, restriction enzyme analysis.

Chinese hamster ovary cells lacking an active DHFR gene are used for transfection. Five μ g of the expression plasmid pC4 is cotransfected with 0.5 μ g of the plasmid pSVneo using a lipid-mediated transfection agent such as Lipofectin™ or LipofectAMINE™ (LifeTechnologies Gaithersburg, MD). The plasmid pSV2-neo contains a dominant

selectable marker, the *neo* gene from Tn5 encoding an enzyme that confers resistance to a group of antibiotics including G418. The cells are seeded in alpha minus MEM supplemented with 1 mg/ml G418. After 2 days, the cells are trypsinized and seeded in hybridoma cloning plates (Greiner, Germany) in alpha minus MEM supplemented with 10, 25, or 50 ng/ml of methotrexate plus 1 mg/ml G418. After about 10-14 days single clones are trypsinized and then seeded in 6-well petri dishes or 10 ml flasks using different concentrations of methotrexate (50 nM, 100 nM, 200 nM, 400 nM, 800 nM). Clones growing at the highest concentrations of methotrexate are then transferred to new 6-well plates containing even higher concentrations of methotrexate (1 μ M, 2 μ M, 5 μ M, 10 mM, 20 mM). The same procedure is repeated until clones are obtained which grow at a concentration of 100-200 μ M. Expression of the desired gene product is analyzed, for instance, by SDS-PAGE and Western blot or by reversed phase HPLC analysis.

Example 5: Quantitative Murine Soft Tissue Infection Model for *S. aureus*

Compositions of the present invention, including polypeptides and peptides, are assayed for their ability to function as vaccines or to enhance/stimulate an immune response to a bacterial species (e.g., *S. aureus*) using the following quantitative murine soft tissue infection model. Mice (e.g., NIH Swiss female mice, approximately 7 weeks old) are first treated with a biologically protective effective amount, or immune enhancing/stimulating effective amount of a composition of the present invention using methods known in the art, such as those discussed above. See, e.g., Harlow et al., ANTIBODIES: A LABORATORY MANUAL, (Cold Spring Harbor Laboratory Press, 2nd ed. 1988). An example of an appropriate starting dose is 20ug per animal.

The desired bacterial species used to challenge the mice, such as *S. aureus*, is grown as an overnight culture. The culture is diluted to a concentration of 5×10^8 cfu/ml, in an appropriate media, mixed well, serially diluted, and titered. The desired doses are further diluted 1:2 with sterilized Cytodex 3 microcarrier beads preswollen in sterile PBS (3g/100ml). Mice are anesthetize briefly until docile, but still mobile and injected with 0.2 ml of the Cytodex 3 bead/bacterial mixture into each animal subcutaneously in the inguinal region. After four days, counting the day of injection as day one, mice are sacrificed and the contents of the abscess is excised and placed in a 15 ml conical tube containing 1.0ml of sterile PBS. The contents of the abscess is then enzymatically treated and plated as follows.

The abscess is first disrupted by vortexing with sterilized glass beads placed in the tubes. 3.0mls of prepared enzyme mixture (1.0ml Collagenase D (4.0 mg/ml), 1.0ml Trypsin (6.0 mg/ml) and 8.0 ml PBS) is then added to each tube followed by a 20 min. incubation at 37C. The solution is then centrifuged and the supernatant drawn off. 0.5 ml dH2O is then added and the tubes are vortexed and then incubated for 10 min. at room temperature. 0.5 ml media is then added and samples are serially diluted and plated onto agar plates, and grown overnight at 37C. Plates with distinct and separate colonies are then counted, compared to positive and negative control samples, and quantified. The method can be used to identify composition and determine appropriate and effective doses for humans and other animals by comparing the effective doses of compositions of the present invention with compositions known in the art to be effective in both mice and humans. Doses for the effective treatment of humans and other animals, using compositions of the present invention, are extrapolated using the data from the above experiments of mice. It is appreciated that further studies in humans and other animals may be needed to determine the most effective doses using methods of clinical practice known in the art.

Example 6: Murine Systemic Neutropenic Model for *S. aureus* Infection

Compositions of the present invention, including polypeptides and peptides, are assayed for their ability to function as vaccines or to enhance/stimulate an immune response to a bacterial species (e.g., *S. aureus*) using the following qualitative murine systemic neutropenic model. Mice (e.g., NIH Swiss female mice, approximately 7 weeks old) are first treated with a biologically protective effective amount, or immune enhancing/stimulating effective amount of a composition of the present invention using methods known in the art, such as those discussed above. See, e.g., Harlow et al., ANTIBODIES: A LABORATORY MANUAL, (Cold Spring Harbor Laboratory Press, 2nd ed. 1988). An example of an appropriate starting dose is 20ug per animal.

Mice are then injected with 250 - 300 mg/kg cyclophosphamide intraperitoneally. Counting the day of C.P. injection as day one, the mice are left untreated for 5 days to begin recovery of PMNL'S.

The desired bacterial species used to challenge the mice, such as *S. aureus*, is grown as an overnight culture. The culture is diluted to a concentration of 5×10^8 cfu/ml, in an

appropriate media, mixed well, serially diluted, and titered. The desired doses are further diluted 1:2 in 4% Brewer's yeast in media.

Mice are injected with the bacteria/brewer's yeast challenge intraperitoneally. The Brewer's yeast solution alone is used as a control. The mice are then monitored twice daily for the first week following challenge, and once a day for the next week to ascertain morbidity and mortality. Mice remaining at the end of the experiment are sacrificed. The method can be used to identify compositions and determine appropriate and effective doses for humans and other animals by comparing the effective doses of compositions of the present invention with compositions known in the art to be effective in both mice and humans. Doses for the effective treatment of humans and other animals, using compositions of the present invention, are extrapolated using the data from the above experiments of mice. It is appreciated that further studies in humans and other animals may be needed to determine the most effective doses using methods of clinical practice known in the art.

Example 7: Murine Lethal Sepsis Model

S. aureus polypeptides of the present invention can be evaluated for potential vaccine efficacy using the murine lethal sepsis model. In this model, mice are challenged with extremely low lethal doses (frequently between 1 and 10 colony forming units [cfu]) of virulent strains of *S. aureus*. Initial studies are conducted to determine a less virulent strain of *S. aureus*. Polypeptides of the present invention (e.g., such as the polypeptides described in Table 1, fragments thereof and fragments that comprise the epitopes shown in Table 4) produced as Example 2(a)-(d), and optionally conjugated with another immunogen are tested as vaccine candidates. Vaccine candidates immunized mice are then challenged with a lethal dose of *S. aureus* which protect against death when approximately 100 times the LD₅₀ of the strain employed are selected as protective antigens.

More specifically, female C2H/HeJ mice are immunized subcutaneously in groups of 10 with 15 ug of protein formulated in complete Freund's adjuvant (CFA). Twenty one days later, mice are boosted in the same way with protein formulated in incomplete Freund's adjuvant. Twenty-eight days following boost animals are bled and a prechallenge immune titer against *S. aureus* proteins is determined by ELISA.

35 days following the boost, a freshly prepared culture of *S. aureus* in BHI are diluted to approximately 35 to 100xLD₅₀ in sterile PBS and injected intraperitoneally into mice in a

volume of 100 ul. Mice are monitored for 14 days for mortality. Survival rate is compared with a sham group immunized with PBS and adjuvant alone.

Example 8: Identifying Vaccine Antigens Against Prevalant *S. aureus* Strains

5 It is further determined whether the majority of the most prevalent *S. aureus* strains express the vaccine antigen(s) and polypeptide(s) identified by the lethal model of Example 7 or the models of Example 5 or 6. Immunoblot analysis is performed with cell lysates prepared from Staphylococcus strains representative of the major capsular serotypes and probed with polyclonal antisera specific for the protective antigens. A preferred vaccine is
10 comprised of a serological epitope of the polypeptide of the present invention that is well conserved among the majority of infective Staphylococcus serotypes.

Example 9: Production of an Antibody

a) Hybridoma Technology

15 The antibodies of the present invention can be prepared by a variety of methods. (See, Current Protocols, Chapter 2.) As one example of such methods, cells expressing polypeptide(s) of the invention are administered to an animal to induce the production of sera containing polyclonal antibodies. In a preferred method, a preparation of polypeptide(s) of the invention is prepared and purified to render it substantially free of natural contaminants.
20 Such a preparation is then introduced into an animal in order to produce polyclonal antisera of greater specific activity.

Monoclonal antibodies specific for polypeptide(s) of the invention are prepared using hybridoma technology. (Kohler et al., Nature 256:495 (1975); Kohler et al., Eur. J. Immunol. 6:511 (1976); Kohler et al., Eur. J. Immunol. 6:292 (1976); Hammerling et al., in:
25 Monoclonal Antibodies and T-Cell Hybridomas, Elsevier, N.Y., pp. 563-681 (1981)). In general, an animal (preferably a mouse) is immunized with polypeptide(s) of the invention or, more preferably, with a secreted polypeptide-expressing cell. Such polypeptide-expressing cells are cultured in any suitable tissue culture medium, preferably in Earle's modified Eagle's medium supplemented with 10% fetal bovine serum (inactivated at about 56°C), and
30 supplemented with about 10 g/l of nonessential amino acids, about 1,000 U/ml of penicillin, and about 100 µg/ml of streptomycin.

The splenocytes of such mice are extracted and fused with a suitable myeloma cell line. Any suitable myeloma cell line may be employed in accordance with the present invention; however, it is preferable to employ the parent myeloma cell line (SP2O), available from the ATCC. After fusion, the resulting hybridoma cells are selectively maintained in HAT medium, and then cloned by limiting dilution as described by Wands et al. (Gastroenterology 80:225-232 (1981)). The hybridoma cells obtained through such a selection are then assayed to identify clones which secrete antibodies capable of binding the polypeptide(s) of the invention.

Alternatively, additional antibodies capable of binding to polypeptide(s) of the invention can be produced in a two-step procedure using anti-idiotypic antibodies. Such a method makes use of the fact that antibodies are themselves antigens, and therefore, it is possible to obtain an antibody which binds to a second antibody. In accordance with this method, protein specific antibodies are used to immunize an animal, preferably a mouse. The splenocytes of such an animal are then used to produce hybridoma cells, and the hybridoma cells are screened to identify clones which produce an antibody whose ability to bind to the protein-specific antibody can be blocked by polypeptide(s) of the invention. Such antibodies comprise anti-idiotypic antibodies to the protein-specific antibody and are used to immunize an animal to induce formation of further protein-specific antibodies.

For in vivo use of antibodies in humans, an antibody is "humanized". Such antibodies can be produced using genetic constructs derived from hybridoma cells producing the monoclonal antibodies described above. Methods for producing chimeric and humanized antibodies are known in the art and are discussed herein. (See, for review, Morrison, Science 229:1202 (1985); Oi et al., BioTechniques 4:214 (1986); Cabilly et al., U.S. Patent No. 4,816,567; Taniguchi et al., EP 171496; Morrison et al., EP 173494; Neuberger et al., WO 8601533; Robinson et al., WO 8702671; Boulianne et al., Nature 312:643 (1984); Neuberger et al., Nature 314:268 (1985).)

*b) Isolation Of Antibody Fragments Directed Against
Polypeptide(s) From A Library Of scFvs*

Naturally occurring V-genes isolated from human PBLs are constructed into a library of antibody fragments which contain reactivities against polypeptide(s) of the invention to

which the donor may or may not have been exposed (see e.g., U.S. Patent 5,885,793 incorporated herein by reference in its entirety).

Rescue of the Library

A library of scFvs is constructed from the RNA of human PBLs as described in PCT publication WO 92/01047. To rescue phage displaying antibody fragments, approximately 109 E. coli harboring the phagemid are used to inoculate 50 ml of 2xTY containing 1% glucose and 100 µg/ml of ampicillin (2xTY-AMP-GLU) and grown to an O.D. of 0.8 with shaking. Five ml of this culture is used to inoculate 50 ml of 2xTY-AMP-GLU, 2×10^8 TU of delta gene 3 helper (M13 delta gene III, see PCT publication WO 92/01047) are added and the culture incubated at 37°C for 45 minutes without shaking and then at 37°C for 45 minutes with shaking. The culture is centrifuged at 4000 r.p.m. for 10 min. and the pellet resuspended in 2 liters of 2xTY containing 100 µg/ml ampicillin and 50 µg/ml kanamycin and grown overnight. Phage are prepared as described in PCT publication WO 92/01047.

M13 delta gene III is prepared as follows: M13 delta gene III helper phage does not encode gene III protein, hence the phage(mid) displaying antibody fragments have a greater avidity of binding to antigen. Infectious M13 delta gene III particles are made by growing the helper phage in cells harboring a pUC19 derivative supplying the wild type gene III protein during phage morphogenesis. The culture is incubated for 1 hour at 37° C without shaking and then for a further hour at 37°C with shaking. Cells are spun down (IEC-Centra 8,400 r.p.m. for 10 min), resuspended in 300 ml 2xTY broth containing 100 µg ampicillin/ml and 25 µg kanamycin/ml (2xTY-AMP-KAN) and grown overnight, shaking at 37°C. Phage particles are purified and concentrated from the culture medium by two PEG-precipitations (Sambrook et al., 1990), resuspended in 2 ml PBS and passed through a 0.45 µm filter (Minisart NML; Sartorius) to give a final concentration of approximately 10¹³ transducing units/ml (ampicillin-resistant clones).

Panning of the Library

Immunotubes (Nunc) are coated overnight in PBS with 4 ml of either 100 µg/ml or 10 µg/ml of a polypeptide of the present invention. Tubes are blocked with 2% Marvel-PBS for 2 hours at 37°C and then washed 3 times in PBS. Approximately 10¹³ TU of phage is applied to the tube and incubated for 30 minutes at room temperature tumbling on an over and under turntable and then left to stand for another 1.5 hours. Tubes are washed 10 times with PBS 0.1% Tween-20 and 10 times with PBS. Phage are eluted by adding 1 ml of 100

mM triethylamine and rotating 15 minutes on an under and over turntable after which the solution is immediately neutralized with 0.5 ml of 1.0M Tris-HCl, pH 7.4. Phage are then used to infect 10 ml of mid-log E. coli TG1 by incubating eluted phage with bacteria for 30 minutes at 37°C. The E. coli are then plated on TYE plates containing 1% glucose and 100 µg/ml ampicillin. The resulting bacterial library is then rescued with delta gene 3 helper phage as described above to prepare phage for a subsequent round of selection. This process is then repeated for a total of 4 rounds of affinity purification with tube-washing increased to 20 times with PBS, 0.1% Tween-20 and 20 times with PBS for rounds 3 and 4.

Characterization of Binders

Eluted phage from the 3rd and 4th rounds of selection are used to infect E. coli HB 2151 and soluble scFv is produced (Marks, et al., 1991) from single colonies for assay. ELISAs are performed with microtitre plates coated with either 10 pg/ml of the polypeptide of the present invention in 50 mM bicarbonate pH 9.6. Clones positive in ELISA are further characterized by PCR fingerprinting (see, e.g., PCT publication WO 92/01047) and then by sequencing. These ELISA positive clones may also be further characterized by techniques known in the art, such as, for example, epitope mapping, binding affinity, receptor signal transduction, ability to block or competitively inhibit antibody/antigen binding, and competitive agonistic or antagonistic activity.

The disclosure of all publications (including patents, patent applications, journal articles, laboratory manuals, books, or other documents) cited herein and the sequence listings are hereby incorporated by reference in their entireties.

The present invention is not to be limited in scope by the specific embodiments described herein, which are intended as single illustrations of individual aspects of the invention. Functionally equivalent methods and components are within the scope of the invention, in addition to those shown and described herein and will become apparent to those skilled in the art from the foregoing description and accompanying drawings. Such modifications are intended to fall within the scope of the appended claims.

The entire disclosure of each document cited (including patents, patent applications, journal articles, abstracts, laboratory manuals, books, or other disclosures) in the Background of the Invention, Detailed Description, and Examples is hereby incorporated herein by reference. Further, the hard copy of the sequence listing submitted herewith and the

corresponding computer readable form are both incorporated herein by reference in their entireties. Moreover, the hard copy of and the corresponding computer readable form of the Sequence Listing of U.S. Patent Application Serial No. 60/151,933 is also incorporated herein by reference in its entirety.

Applicant's or agent's file referencenumber	PB515PCT	International application No.	UNASSIGNED
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INDICATIONS RELATING TO A DEPOSITED MICROORGANISM

(PCT Rule 13bis)

A. The indications made below relate to the microorganism referred to in the description on page <u>12</u> , line <u>29</u>	
B. IDENTIFICATION OF DEPOSIT Further deposits are identified on an additional sheet <input type="checkbox"/>	
Name of depositary institution American Type Culture Collection	
Address of depositary institution (including postal code and country) 10801 University Boulevard Manassas, Virginia 20110-2209 United States of America	
Date of deposit 07 April 1998	Accession Number 202108
C. ADDITIONAL INDICATIONS (leave blank if not applicable) This information is continued on an additional sheet <input type="checkbox"/>	
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States) Europe In respect to those designations in which a European Patent is sought a sample of the deposited microorganism will be made available until the publication of the mention of the grant of the European patent or until the date on which application has been refused or withdrawn or is deemed to be withdrawn, only by the issue of such a sample to an expert nominated by the person requesting the sample (Rule 28 (4) EPC). Continued on the Attached Pages 2 & 3	
E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable) The indications listed below will be submitted to the International Bureau later (specify the general nature of the indications e.g., "Accession Number of Deposit")	

<input checked="" type="checkbox"/> For receiving Office use only This sheet was received with the international application Sonya D. Barnes Authorized officer PCT Internat'l Appl Processing Div (703) 305-3665	<input type="checkbox"/> For International Bureau use only This sheet was received by the International Bureau on: Authorized officer
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ATCC Deposit No. 202108**Page No. 2****CANADA**

The applicant requests that, until either a Canadian patent has been issued on the basis of an application or the application has been refused, or is abandoned and no longer subject to reinstatement, or is withdrawn, the Commissioner of Patents only authorizes the furnishing of a sample of the deposited biological material referred to in the application to an independent expert nominated by the Commissioner, the applicant must, by a written statement, inform the International Bureau accordingly before completion of technical preparations for publication of the international application.

NORWAY

The applicant hereby requests that the application has been laid open to public inspection (by the Norwegian Patent Office), or has been finally decided upon by the Norwegian Patent Office without having been laid open inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the Norwegian Patent Office not later than at the time when the application is made available to the public under Sections 22 and 33(3) of the Norwegian Patents Act. If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on the list of recognized experts drawn up by the Norwegian Patent Office or any person approved by the applicant in the individual case.

AUSTRALIA

The applicant hereby gives notice that the furnishing of a sample of a microorganism shall only be effected prior to the grant of a patent, or prior to the lapsing, refusal or withdrawal of the application, to a person who is a skilled addressee without an interest in the invention (Regulation 3.25(3) of the Australian Patents Regulations).

FINLAND

The applicant hereby requests that, until the application has been laid open to public inspection (by the National Board of Patents and Regulations), or has been finally decided upon by the National Board of Patents and Registration without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art.

UNITED KINGDOM

The applicant hereby requests that the furnishing of a sample of a microorganism shall only be made available to an expert. The request to this effect must be filed by the applicant with the International Bureau before the completion of the technical preparations for the international publication of the application.

ATCC Deposit No.: 202108

Page No. 3

DENMARK

The applicant hereby requests that, until the application has been laid open to public inspection (by the Danish Patent Office), or has been finally decided upon by the Danish Patent office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the Danish Patent Office not later than at the time when the application is made available to the public under Sections 22 and 33(3) of the Danish Patents Act. If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the Danish Patent Office or any person by the applicant in the individual case.

SWEDEN

The applicant hereby requests that, until the application has been laid open to public inspection (by the Swedish Patent Office), or has been finally decided upon by the Swedish Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the International Bureau before the expiration of 16 months from the priority date (preferably on the Form PCT/RO/134 reproduced in annex Z of Volume I of the PCT Applicant's Guide). If such a request has been filed by the applicant any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the Swedish Patent Office or any person approved by a applicant in the individual case.

NETHERLANDS

The applicant hereby requests that until the date of a grant of a Netherlands patent or until the date on which the application is refused or withdrawn or lapsed, the microorganism shall be made available as provided in the 31F(1) of the Patent Rules only by the issue of a sample to an expert. The request to this effect must be furnished by the applicant with the Netherlands Industrial Property Office before the date on which the application is made available to the public under Section 22C or Section 25 of the Patents Act of the Kingdom of the Netherlands, whichever of the two dates occurs earlier.

What Is Claimed Is:

1. An isolated nucleic acid molecule comprising a polynucleotide having a nucleotide sequence selected from the group consisting of:

- 5 (a) a nucleotide sequence encoding any one of the amino acid sequences of the polypeptides shown in Table 1; or
- (b) a nucleotide sequence complementary to any one of the nucleotide sequences in (a);
- 10 (c) a nucleotide sequence at least 95% identical to any one of the nucleotide sequences shown in Table 1; or,
- (d) a nucleotide sequence at least 95% identical to a nucleotide sequence complementary to any one of the nucleotide sequences shown in Table 1.

15 2. An isolated nucleic acid molecule of claim 1 comprising a polynucleotide which hybridizes under stringent hybridization conditions to a polynucleotide having a nucleotide sequence identical to a nucleotide sequence in (a) or (b) of claim 1.

20 3. An isolated nucleic acid molecule of claim 1 comprising a polynucleotide which encodes an epitope-bearing portion of a polypeptide in (a) of claim 1.

4. The isolated nucleic acid molecule of claim 3, wherein said epitope-bearing portion of a polypeptide comprises an amino acid sequence listed in Table 4.

25 5. A method for making a recombinant vector comprising inserting an isolated nucleic acid molecule of claim 1 into a vector.

6. A recombinant vector produced by the method of claim 5.

30 7. A host cell comprising the vector of claim 6.

8. A method of producing a polypeptide comprising:

- (a) growing the host cell of claim 7 such that the protein is expressed by the cell; and

(b) recovering the expressed polypeptide.

9. An isolated polypeptide comprising an amino acid sequence selected from the group consisting of:

- (a) a complete amino acid sequences of Table 1;
- (b) a complete amino acid sequence of Table 1 except the N-terminal residue;
- (c) a fragment of a polypeptide of Table 1 having biological activity; and
- (d) a fragment of a polypeptide of Table 1 which binds to an antibody specific for a *S. aureus* polypeptide.

10. An isolated polypeptide comprising an amino acid sequence at least 95% identical to an amino acid sequence of Table 1.

11. An isolated epitope-bearing polypeptide comprising an amino acid sequence of Table 4.

12. An isolated antibody specific for the polypeptide of claim 9.

13. A host cell which produces an antibody of claim 12.

14. A vaccine, comprising:

- (1) one or more *S. aureus* polypeptides selected from the group consisting of a polypeptide of claim 9; and

- (2) a pharmaceutically acceptable diluent, carrier, or excipient;

wherein said polypeptide is present, in an amount effective to elicit protective antibodies in an animal to a member of the *Staphylococcus* genus.

15. A method of preventing or attenuating an infection caused by a member of the *Staphylococcus* genus in an animal, comprising administering to said animal a polypeptide of claim 9, wherein said polypeptide is administered in an amount effective to prevent or attenuate said infection.

16. A method of detecting *Staphylococcus* nucleic acids in a biological sample comprising:

- (a) contacting the sample with one or more nucleic acids of claim 1, under conditions such that hybridization occurs, and
- (b) detecting hybridization of said nucleic acids to the one or more *Staphylococcus* nucleic acid sequences present in the biological sample.

5

17. A method of detecting *Staphylococcus* nucleic acids in a biological sample obtained from an animal, comprising:

- (a) amplifying one or more *Staphylococcus* nucleic acid sequences in said sample using polymerase chain reaction, and
- (b) detecting said amplified *Staphylococcus* nucleic acid.

10

18. A kit for detecting *Staphylococcus* antibodies in a biological sample obtained from an animal, comprising

- (a) a polypeptide of claim 9 attached to a solid support; and
- (b) detecting means.

15

19. A method of detecting *Staphylococcus* antibodies in a biological sample obtained from an animal, comprising

- (a) contacting the sample with a polypeptide of claim 9; and
- (b) detecting antibody-antigen complexes.

20

20. A method of detecting a polypeptide of claim 9 comprising:

- (a) obtaining a biological sample suspected of containing said polypeptide; and
- (b) determining the presence or absence of said polypeptide in said biological sample.

25

21. The method of claim 20, wherein said method comprises a step of contacting the sample with an antibody.

30

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Arg Asp Glu Leu Arg Ala Gln	Arg Ile Leu Gln Asp Arg	Ala Phe Lys	
180	185	190	
Asn Asp Lys Ile Asp Phe Ile	Trp Ser His Thr Leu Lys	Ser Ile Asn	
195	200	205	
Glu Lys Asp Gly Lys Val Gly	Ser Val Thr Leu Thr Ser	Thr Lys Asp	
210	215	220	
Gly Ser Glu Glu Thr His	Glu Ala Asp Gly Val Phe	Ile Tyr Ile Gly	
225	230	235	240
Met Lys Pro Leu Thr Ala	Pro Phe Lys Asp Leu Gly	Ile Thr Asn Asp	
245	250	255	
Val Gly Tyr Ile Val Thr	Lys Asp Asp Met Thr Thr	Ser Val Pro Gly	
260	265	270	
Ile Phe Ala Ala Gly Asp	Val Arg Asp Lys Gly Leu	Arg Gln Ile Val	
275	280	285	
Thr Ala Thr Gly Asp Gly	Ser Ile Ala Ala Gln Ser	Ala Ala Glu Tyr	
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Ile Glu His Leu Asn Asp	Gln Ala		
305	310		

<210> 15
 <211> 1356
 <212> DNA
 <213> Homo sapiens

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 ttaccaagac cagttggcaa tgatattgta cattattcag attactttga aggggcacaa 480
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 gcagatactg aaacaattgg atgtagtcct gatggatata atatcaatga gaaatgtggc 660
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 ggtgaccaaa ttatgtttat tattgttcaa gaaatgcata aaaatcaaga attgaataat 840
 gacatgattg tttctactgt tatgagtaat ttaggttttt acaaagcgct tgaacaagaa 900
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 gaagtagaaa tgaatggaga aggtcgaatt ttagtaagac cttctggaac agaaccatta 1260
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<210> 16
 <211> 452
 <212> PRT
 <213> Homo sapiens

<400> 16
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 20 25 30
 Tyr Val Leu Ala His Asn Lys Gly Glu Lys His Pro Arg Val Leu Val
 35 40 45
 Gly Arg Asp Thr Arg Val Ser Gly Glu Met Leu Glu Ser Ala Leu Ile
 50 55 60
 Ala Gly Leu Ile Ser Ile Gly Ala Glu Val Met Arg Leu Gly Ile Ile
 65 70 75 80
 Ser Thr Pro Gly Val Ala Tyr Leu Thr Arg Asp Met Gly Ala Glu Leu
 85 90 95
 Gly Val Met Ile Ser Ala Ser His Asn Pro Val Ala Asp Asn Gly Ile
 100 105 110

Lys	Phe	Phe	Gly	Ser	Asp	Gly	Phe	Lys	Leu	Ser	Asp	Glu	Gln	Glu	Asn	
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Glu	Ile	Glu	Ala	Leu	Leu	Asp	Gln	Glu	Asn	Pro	Glu	Leu	Pro	Arg	Pro	
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Val	Gly	Asn	Asp	Ile	Val	His	Tyr	Ser	Asp	Tyr	Phe	Glu	Gly	Ala	Gln	
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Lys	Tyr	Leu	Ser	Tyr	Leu	Lys	Ser	Thr	Val	Asp	Val	Asn	Phe	Glu	Gly	
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Leu	Lys	Ile	Ala	Leu	Asp	Gly	Ala	Asn	Gly	Ser	Thr	Ser	Ser	Leu	Ala	
		180						185					190			
Pro	Phe	Leu	Phe	Gly	Asp	Leu	Glu	Ala	Asp	Thr	Glu	Thr	Ile	Gly	Cys	
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Ser	Pro	Asp	Gly	Tyr	Asn	Ile	Asn	Glu	Lys	Cys	Gly	Ser	Thr	His	Pro	
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Glu	Lys	Leu	Ala	Glu	Lys	Val	Val	Glu	Thr	Glu	Ser	Asp	Phe	Gly	Leu	
225				230				235							240	
Ala	Phe	Asp	Gly	Asp	Gly	Asp	Arg	Ile	Ile	Ala	Val	Asp	Glu	Asn	Gly	
			245					250						255		
Gln	Ile	Val	Asp	Gly	Asp	Gln	Ile	Met	Phe	Ile	Ile	Gly	Gln	Glu	Met	
		260					265						270			
His	Lys	Asn	Gln	Glu	Leu	Asn	Asn	Asp	Met	Ile	Val	Ser	Thr	Val	Met	
	275					280						285				
Ser	Asn	Leu	Gly	Phe	Tyr	Lys	Ala	Leu	Glu	Gln	Glu	Gly	Ile	Lys	Ser	
	290				295					300						
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			325					330						335		
Asp	Tyr	Asn	Thr	Thr	Gly	Asp	Gly	Leu	Leu	Thr	Gly	Ile	Gln	Leu	Ala	
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Ser	Val	Ile	Lys	Met	Thr	Gly	Lys	Ser	Leu	Ser	Glu	Leu	Ala	Gly	Gln	
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Met	Lys	Lys	Tyr	Pro	Gln	Ser	Leu	Ile	Asn	Val	Arg	Val	Thr	Asp	Lys	
	370				375						380					
Tyr	Arg	Val	Glu	Glu	Asn	Val	Asp	Val	Lys	Glu	Val	Met	Thr	Lys	Val	
385				390					395						400	
Glu	Val	Glu	Met	Asn	Gly	Glu	Gly	Arg	Ile	Leu	Val	Arg	Pro	Ser	Gly	
			405					410						415		

Thr Glu Pro Leu Val Arg Val Met Val Glu Ala Ala Thr Asp Glu Asp
 420 425 430

Ala Glu Arg Phe Ala Gln Gln Ile Ala Asp Val Val Gln Asp Lys Met
 435 440 445

Gly Leu Asp Lys
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<210> 17

<211> 1359

<212> DNA

<213> Homo sapiens

<400> 17

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gttgatgtgt gtgacacacc gctcatcaca aaggaaacat tagtaacatt gattgcgcat 360
cagcaggatg ctaatgtcga agcaactgta ttatctgcat cgattcaaca accatatgga 420
tacggaagaa tcgttcgaaa tgcgtcaggt cgtttagaac gcatagttga agagaaagat 480
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gcatgcaac gtcgtacgaa tcattatcac atgctaaatg gtgtgacaat catcgatcct 780
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<210> 18

<211> 453

<212> PRT

<213> Homo sapiens

<400> 18

Met Gly Phe Met Arg Arg His Ala Ile Ile Leu Ala Ala Gly Lys Gly
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Thr Arg Met Lys Ser Lys Lys Tyr Lys Val Leu His Glu Val Ala Gly
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Lys Pro Met Val Glu His Val Leu Glu Ser Val Lys Gly Ser Gly Val
 35 40 45

Asp Gln Val Val Thr Ile Val Gly His Gly Ala Glu Ser Val Lys Gly
 50 55 60

His Leu Gly Glu Arg Ser Leu Tyr Ser Phe Gln Glu Glu Gln Leu Gly
65 70 75 80

Thr Ala His Ala Val Gln Met Ala Lys Ser His Leu Glu Asp Lys Glu
85 90 95

Gly Thr Thr Ile Val Val Cys Gly Asp Thr Pro Leu Ile Thr Lys Glu
100 105 110

Thr Leu Val Thr Leu Ile Ala His His Glu Asp Ala Asn Ala Gln Ala
115 120 125

Thr Val Leu Ser Ala Ser Ile Gln Gln Pro Tyr Gly Tyr Gly Arg Ile
130 135 140

Val Arg Asn Ala Ser Gly Arg Leu Glu Arg Ile Val Glu Glu Lys Asp
145 150 155 160

Ala Thr Gln Ala Glu Lys Asp Ile Asn Glu Ile Ser Ser Gly Ile Phe
165 170 175

Ala Phe Asn Asn Lys Thr Leu Phe Glu Lys Leu Thr Gln Val Lys Asn
180 185 190

Asp Asn Ala Gln Gly Glu Tyr Tyr Leu Pro Asp Val Leu Ser Leu Ile
195 200 205

Leu Asn Asp Gly Gly Ile Val Glu Val Tyr Arg Thr Asn Asp Val Glu
210 215 220

Glu Ile Met Gly Val Asn Asp Arg Val Met Leu Ser Gln Ala Glu Lys
225 230 235 240

Ala Met Gln Arg Arg Thr Asn His Tyr His Met Leu Asn Gly Val Thr
245 250 255

Ile Ile Asp Pro Asp Ser Thr Tyr Ile Gly Pro Asp Val Thr Ile Gly
260 265 270

Ser Asp Thr Val Ile Glu Pro Gly Val Arg Ile Asn Gly Arg Thr Glu
275 280 285

Ile Gly Glu Asp Val Val Ile Gly Gln Tyr Ser Glu Ile Asn Asn Ser
290 295 300

Thr Ile Glu Asn Gly Ala Cys Ile Gln Gln Ser Val Val Asn Asp Ala
305 310 315 320

Ser Val Gly Ala Asn Thr Lys Val Gly Pro Phe Ala Gln Leu Arg Pro
325 330 335

Gly Ala Gln Leu Gly Ala Asp Val Lys Val Gly Asn Phe Val Glu Ile
340 345 350

Lys Lys Ala Asp Leu Lys Asp Gly Ala Lys Val Ser His Leu Ser Tyr
355 360 365

Ile Gly Asp Ala Val Ile Gly Glu Arg Thr Asn Ile Gly Cys Gly Thr

16

370 375 380

Ile Thr Val Asn Tyr Asp Gly Glu Asn Lys Phe Lys Thr Ile Val Gly
 385 390 395 400

Lys Asp Ser Phe Val Gly Cys Asn Val Asn Leu Val Ala Pro Val Thr
 405 410 415

Ile Gly Asp Asp Val Leu Val Ala Ala Gly Ser Thr Ile Thr Asp Asp
 420 425 430

Val Pro Asn Asp Ser Leu Ala Val Ala Arg Ala Arg Gln Thr Thr Lys
 435 440 445

Glu Gly Tyr Arg Lys
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<210> 19
 <211> 1317
 <212> DNA
 <213> Homo sapiens

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 tttagctata cacctgaaaa attttatgat agaaagcaaa ttacagtaaa aacttatcat 240
 gaagtatttg caatcaatga tgaaagacaa actgtatctg tattaataag aaagacaaac 300
 gaacaatttg aagaatctta cgataaactc attttaagcc ctggtgcaag tgcaaatagc 360
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 gatcaattca tcaaagcaaa tcaagttgat aaagtattgg ttgtaggtgc aggttatgtt 480
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<210> 20
 <211> 439
 <212> PRT
 <213> Homo sapiens

<400> 20

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Cys Ala Ser Gln Ile Arg Arg Leu Asp Lys Glu Ser Asp Ile Ile Ile
 20 25 30

Phe Glu Lys Asp Arg Asp Met Ser Phe Ala Asn Cys Ala Leu Pro Tyr
 35 40 45

Val Ile Gly Glu Val Val Glu Asp Arg Arg Tyr Ala Leu Ala Tyr Thr
 50 55 60

Pro Glu Lys Phe Tyr Asp Arg Lys Gln Ile Thr Val Lys Thr Tyr His
 65 70 75 80

Glu Val Ile Ala Ile Asn Asp Glu Arg Gln Thr Val Ser Val Leu Asn
 85 90 95

Arg Lys Thr Asn Glu Gln Phe Glu Glu Ser Tyr Asp Lys Leu Ile Leu
 100 105 110

Ser Pro Gly Ala Ser Ala Asn Ser Leu Gly Phe Glu Ser Asp Ile Thr
 115 120 125

Phe Thr Leu Arg Asn Leu Glu Asp Thr Asp Ala Ile Asp Gln Phe Ile
 130 135 140

Lys Ala Asn Gln Val Asp Lys Val Leu Val Val Gly Ala Gly Tyr Val
 145 150 155 160

Ser Leu Glu Val Leu Glu Asn Leu Tyr Glu Arg Gly Leu His Pro Thr
 165 170 175

Leu Ile His Arg Ser Asp Lys Ile Asn Lys Leu Met Asp Ala Asp Met
 180 185 190

Asn Gln Pro Ile Leu Asp Glu Leu Asp Lys Arg Glu Ile Pro Tyr Arg
 195 200 205

Leu Asn Glu Glu Ile Asn Ala Ile Asn Gly Asn Glu Ile Thr Phe Lys
 210 215 220

Ser Gly Lys Val Glu His Tyr Asp Met Ile Ile Glu Gly Val Gly Thr
 225 230 235 240

His Pro Asn Ser Lys Phe Ile Glu Ser Ser Asn Ile Lys Leu Asp Arg
 245 250 255

Lys Gly Phe Ile Pro Val Asn Asp Lys Phe Glu Thr Asn Val Pro Asn
 260 265 270

Ile Tyr Ala Ile Gly Asp Ile Ala Thr Ser His Tyr Arg His Val Asp
 275 280 285

Leu Pro Ala Ser Val Pro Leu Ala Trp Gly Ala His Arg Ala Ala Ser
 290 295 300

Ile Val Ala Glu Gln Ile Ala Gly Asn Asp Thr Ile Glu Phe Lys Gly
 305 310 315 320

Phe Leu Gly Asn Asn Ile Val Lys Phe Phe Asp Tyr Thr Phe Ala Ser
 325 330 335

Val Gly Val Lys Pro Asn Glu Leu Lys Gln Phe Asp Tyr Lys Met Val
 340 345 350

Glu Val Thr Gln Gly Ala His Ala Asn Tyr Tyr Pro Gly Asn Ser Pro
 355 360 365

Leu His Leu Arg Val Tyr Tyr Asp Thr Ser Asn Arg Gln Ile Leu Arg
 370 375 380

Ala Ala Ala Val Gly Lys Glu Gly Ala Asp Lys Arg Ile Asp Val Leu
 385 390 395 400

Ser Met Ala Met Met Asn Gln Leu Thr Val Asp Glu Leu Thr Glu Phe
 405 410 415

Glu Val Ala Tyr Ala Pro Pro Tyr Ser His Pro Lys Asp Leu Ile Asn
 420 425 430

Met Ile Gly Tyr Lys Ala Lys
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<210> 21

<211> 1353

<212> DNA

<213> Homo sapiens

<400> 21

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<210> 22

<211> 451

<212> PRT

<213> Homo sapiens

<400> 22

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 20 25 30

Val Ile Tyr Gly Ile Leu Asn Ile Tyr Phe Ile Gly Phe Leu Glu Asp
 35 40 45

Ser His Met Ile Ser Ala Ile Ser Leu Thr Leu Pro Val Phe Ala Ile
 50 55 60

Leu Met Gly Leu Gly Asn Leu Phe Gly Val Gly Ala Gly Thr Tyr Ile
 65 70 75 80

Ser Arg Leu Leu Gly Ala Lys Asp Tyr Ser Lys Ser Lys Phe Val Ser
 85 90 95

Ser Phe Ser Ile Tyr Gly Gly Ile Ala Leu Gly Leu Ile Val Ile Leu
 100 105 110

Val Thr Leu Pro Phe Ser Asp Gln Ile Ala Ala Ile Leu Gly Ala Arg
 115 120 125

Gly Glu Thr Leu Ala Leu Thr Ser Asn Tyr Leu Lys Val Met Phe Leu
 130 135 140

Ser Ala Pro Phe Val Ile Leu Phe Phe Ile Leu Glu Gln Phe Ala Arg
 145 150 155 160

Ala Ile Gly Ala Pro Met Val Ser Met Ile Gly Met Leu Ala Ser Val
 165 170 175

Gly Leu Asn Ile Ile Leu Asp Pro Ile Leu Ile Phe Gly Phe Asp Leu
 180 185 190

Asn Val Val Gly Ala Ala Leu Gly Thr Ala Ile Ser Asn Val Ala Ala
 195 200 205

Ala Leu Phe Phe Ile Ile Tyr Phe Met Lys Asn Ser Asp Val Val Ser
 210 215 220

Val Asn Ile Lys Leu Ala Lys Pro Asn Lys Glu Met Leu Ser Glu Ile
 225 230 235 240

Phe Lys Ile Gly Ile Pro Ala Phe Leu Met Ser Ile Leu Met Gly Phe
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Thr Gly Leu Val Leu Asn Leu Phe Leu Ala His Tyr Gly Asn Phe Ala
 260 265 270

Ile Ala Ser Tyr Gly Ile Ser Phe Arg Leu Val Gln Phe Pro Glu Leu
 275 280 285

Ile Ile Met Gly Leu Cys Glu Gly Val Val Pro Leu Ile Ala Tyr Asn
 290 295 300

Phe Met Ala Asn Lys Gly Arg Met Lys Asp Val Ile Lys Ala Val Ile

305 310 315 320
 Met Ser Ile Gly Val Ile Phe Val Val Cys Met Ser Ala Val Phe Thr
 325 330 335
 Ile Gly His His Met Val Gly Leu Phe Thr Thr Asp Gln Ala Ile Val
 340 345 350
 Glu Met Ala Thr Phe Ile Leu Lys Val Thr Met Ala Ser Leu Leu Leu
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 Asn Gly Ile Gly Phe Leu Phe Thr Gly Met Leu Gln Ala Thr Gly Gln
 370 375 380
 Gly Arg Gly Ala Thr Ile Met Ala Ile Leu Gln Gly Ala Ile Ile Ile
 385 390 395 400
 Pro Val Leu Phe Ile Met Asn Ala Leu Phe Gly Leu Thr Gly Val Ile
 405 410 415
 Trp Ser Leu Leu Ile Ala Glu Ser Leu Cys Ala Leu Ala Ala Met Leu
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<210> 23

<211> 1479

<212> DNA

<213> Homo sapiens

<400> 23

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 gtcattttca caccggataa tccggcaaat gatgaccga aaatgttaac ggcagaatta 1260
 gccaaaggtg caacacatca aaactatatt gaatttgatg atcgtgcaga agggataaaa 1320

catgcaattg acatagctga gcctggggat actgtcgttt tagcatcaaa aggaagagaa 1380
 ccataatcaaa tcatgccagg gcatattaag gtgccacatc gagatgattt aattggcctt 1440
 gaagcagctt acaaaaagtt cgggtggtggc cctgttgat 1479

<210> 24

<211> 493

<212> PRT

<213> Homo sapiens

<400> 24

Leu Asp Ala Ser Thr Leu Phe Lys Lys Val Lys Val Lys Arg Val Leu
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Gly Ser Leu Glu Gln Gln Ile Asp Asp Ile Thr Thr Asp Ser Arg Thr
 20 25 30

Ala Arg Glu Gly Ser Ile Phe Val Ala Ser Val Gly Tyr Thr Val Asp
 35 40 45

Ser His Lys Phe Cys Gln Asn Val Ala Asp Gln Gly Cys Lys Leu Val
 50 55 60

Val Val Asn Lys Glu Gln Ser Leu Pro Ala Asn Val Thr Gln Val Val
 65 70 75 80

Val Pro Asp Thr Leu Arg Val Ala Ser Ile Leu Ala His Thr Leu Tyr
 85 90 95

Asp Tyr Pro Ser His Gln Leu Val Thr Phe Gly Val Thr Gly Thr Asn
 100 105 110

Gly Lys Thr Ser Ile Ala Thr Met Ile His Leu Ile Gln Arg Lys Leu
 115 120 125

Gln Lys Asn Ser Ala Tyr Leu Gly Thr Asn Gly Phe Gln Ile Asn Glu
 130 135 140

Thr Lys Thr Lys Gly Ala Asn Thr Thr Pro Glu Thr Val Ser Leu Thr
 145 150 155 160

Lys Lys Ile Lys Glu Ala Val Asp Ala Gly Ala Glu Ser Met Thr Leu
 165 170 175

Glu Val Ser Ser His Gly Leu Val Leu Gly Arg Leu Arg Gly Val Glu
 180 185 190

Phe Asp Val Ala Ile Phe Ser Asn Leu Thr Gln Asp His Leu Asp Phe
 195 200 205

His Gly Thr Met Glu Ala Tyr Gly His Ala Lys Ser Leu Leu Phe Ser
 210 215 220

Gln Leu Gly Glu Asp Leu Ser Lys Glu Lys Tyr Val Val Leu Asn Asn
 225 230 235 240

Asp Asp Ser Phe Ser Glu Tyr Leu Arg Thr Val Thr Pro Tyr Glu Val
 245 250 255

Phe Ser Tyr Gly Ile Asp Glu Glu Ala Gln Phe Met Ala Lys Asn Ile
 260 265 270
 Gln Glu Ser Leu Gln Gly Val Ser Phe Asp Phe Val Thr Pro Phe Gly
 275 280 285
 Thr Tyr Pro Val Lys Ser Pro Tyr Val Gly Lys Phe Asn Ile Ser Asn
 290 295 300
 Ile Met Ala Ala Met Ile Ala Val Trp Ser Lys Gly Thr Ser Leu Glu
 305 310 315 320
 Thr Ile Ile Lys Ala Val Glu Asn Leu Glu Pro Val Glu Gly Arg Leu
 325 330 335
 Glu Val Leu Asp Pro Ser Leu Pro Ile Asp Leu Ile Ile Asp Tyr Ala
 340 345 350
 His Thr Ala Asp Gly Met Asn Lys Leu Ile Asp Ala Val Gln Pro Phe
 355 360 365
 Val Lys Gln Lys Leu Ile Phe Leu Val Gly Met Ala Gly Glu Arg Asp
 370 375 380
 Leu Thr Lys Thr Pro Glu Met Gly Arg Val Ala Cys Arg Ala Asp Tyr
 385 390 395 400
 Val Ile Phe Thr Pro Asp Asn Pro Ala Asn Asp Asp Pro Lys Met Leu
 405 410 415
 Thr Ala Glu Leu Ala Lys Gly Ala Thr His Gln Asn Tyr Ile Glu Phe
 420 425 430
 Asp Asp Arg Ala Glu Gly Ile Lys His Ala Ile Asp Ile Ala Glu Pro
 435 440 445
 Gly Asp Thr Val Val Leu Ala Ser Lys Gly Arg Glu Pro Tyr Gln Ile
 450 455 460
 Met Pro Gly His Ile Lys Val Pro His Arg Asp Asp Leu Ile Gly Leu
 465 470 475 480
 Glu Ala Ala Tyr Lys Lys Phe Gly Gly Gly Pro Val Asp
 485 490

<210> 25

<211> 1356

<212> DNA

<213> Homo sapiens

<400> 25

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 tttttaaatc aagagataaa tggagtcaca attgattcac gagcaatttc taaaaatatg 120
 ttatttatac catttaaaagg tgaaaatgtt gacggctatc gctttgtctc taaagcatta 180
 caagatgggtg ctggggctgc tttttatcaa agagggacac ctatagatga aaatgtaagc 240
 gggcctatta tatgggttga agacacatta acggcattac aacaattggc acaagcttac 300
 ttgagacatg taaaccctaa agtaattgcc gtcacagggt ctaatggtaa aacaacgact 360

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aaagatatga ttgaaagtgt attgcatacc gaatttaaag ttaagaaaac gcaaggtaat 420
tacaataatg aaattgggtt acctttaact attttggaat tagataatga tactgaaata 480
tcaatattgg agatggggat gtcaggtttc catgaaattg aatttctgtc aaacctcgct 540
caaccagata ttgcagttat aactaatatt ggtgagtcac atatgcaaga tttagggttcg 600
cgcgagggga ttgctaaagc taaatctgaa attacaatag gtctaaaaga taatggtacg 660
tttatatatg atggcgatga accattattg aaaccacatg ttaaagaagt tgaaaatgca 720
aaatgtatta gtattgggtg tgctactgat aatgcattag ttgttctgt tgatgataga 780
gatactacag gtatttcatt tacgattaat aataaagaac attacgatct gccaatatta 840
ggaaagcata atatgaaaaa tgcgacgatt gccattgcgg ttggtcatga attagggttg 900
acataataca caatctatca aaatttaaaa aatgtcagct taactggtat gcgtaggaa 960
caacatacat tagaaaatga tattactgtg ataaatgat cctataatgc aagtcctaca 1020
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ttagaagaaa agcatataga tgtgtttgat acgtttggta atgaagcgaa gtatatttat 1200
gattcgggcc agcaacatgt cgaaaaagca caacacttca attctaaaga cgatatgata 1260
gaagttttaa taaacgattt aaaagcgcat gaccgtgat tagttaaagg atcacgtggt 1320
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<210> 26

<211> 452

<212> PRT

<213> Homo sapiens

<400> 26

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Met Ile Asn Val Thr Leu Lys Gln Ile Gln Ser Trp Ile Pro Cys Glu
  1             5             10             15

```

```

Ile Glu Asp Gln Phe Leu Asn Gln Glu Ile Asn Gly Val Thr Ile Asp
      20             25             30

```

```

Ser Arg Ala Ile Ser Lys Asn Met Leu Phe Ile Pro Phe Lys Gly Glu
    35             40             45

```

```

Asn Val Asp Gly His Arg Phe Val Ser Lys Ala Leu Gln Asp Gly Ala
    50             55             60

```

```

Gly Ala Ala Phe Tyr Gln Arg Gly Thr Pro Ile Asp Glu Asn Val Ser
    65             70             75             80

```

```

Gly Pro Ile Ile Trp Val Glu Asp Thr Leu Thr Ala Leu Gln Gln Leu
      85             90             95

```

```

Ala Gln Ala Tyr Leu Arg His Val Asn Pro Lys Val Ile Ala Val Thr
    100            105            110

```

```

Gly Ser Asn Gly Lys Thr Thr Thr Lys Asp Met Ile Glu Ser Val Leu
    115            120            125

```

```

His Thr Glu Phe Lys Val Lys Lys Thr Gln Gly Asn Tyr Asn Asn Glu
    130            135            140

```

```

Ile Gly Leu Pro Leu Thr Ile Leu Glu Leu Asp Asn Asp Thr Glu Ile
    145            150            155            160

```

```

Ser Ile Leu Glu Met Gly Met Ser Gly Phe His Glu Ile Glu Phe Leu
      165            170            175

```

```

Ser Asn Leu Ala Gln Pro Asp Ile Ala Val Ile Thr Asn Ile Gly Glu

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180					185					190					
Ser	His	Met	Gln	Asp	Leu	Gly	Ser	Arg	Glu	Gly	Ile	Ala	Lys	Ala	Lys
		195					200					205			
Ser	Glu	Ile	Thr	Ile	Gly	Leu	Lys	Asp	Asn	Gly	Thr	Phe	Ile	Tyr	Asp
		210					215					220			
Gly	Asp	Glu	Pro	Leu	Leu	Lys	Pro	His	Val	Lys	Glu	Val	Glu	Asn	Ala
		225					230					235			240
Lys	Cys	Ile	Ser	Ile	Gly	Val	Ala	Thr	Asp	Asn	Ala	Leu	Val	Cys	Ser
				245					250						255
Val	Asp	Asp	Arg	Asp	Thr	Thr	Gly	Ile	Ser	Phe	Thr	Ile	Asn	Asn	Lys
			260					265					270		
Glu	His	Tyr	Asp	Leu	Pro	Ile	Leu	Gly	Lys	His	Asn	Met	Lys	Asn	Ala
		275					280					285			
Thr	Ile	Ala	Ile	Ala	Val	Gly	His	Glu	Leu	Gly	Leu	Thr	Tyr	Asn	Thr
		290					295					300			
Ile	Tyr	Gln	Asn	Leu	Lys	Asn	Val	Ser	Leu	Thr	Gly	Met	Arg	Met	Glu
		305					310					315			320
Gln	His	Thr	Leu	Glu	Asn	Asp	Ile	Thr	Val	Ile	Asn	Asp	Ala	Tyr	Asn
			325						330						335
Ala	Ser	Pro	Thr	Ser	Met	Arg	Ala	Ala	Ile	Asp	Thr	Leu	Ser	Thr	Leu
			340					345					350		
Thr	Gly	Arg	Arg	Ile	Leu	Ile	Leu	Gly	Asp	Val	Leu	Glu	Leu	Gly	Glu
		355					360					365			
Asn	Ser	Lys	Glu	Met	His	Ile	Gly	Val	Gly	Asn	Tyr	Leu	Glu	Glu	Lys
		370					375					380			
His	Ile	Asp	Val	Leu	Tyr	Thr	Phe	Gly	Asn	Glu	Ala	Lys	Tyr	Ile	Tyr
		385					390					395			400
Asp	Ser	Gly	Gln	Gln	His	Val	Glu	Lys	Ala	Gln	His	Phe	Asn	Ser	Lys
			405						410						415
Asp	Asp	Met	Ile	Glu	Val	Leu	Ile	Asn	Asp	Leu	Lys	Ala	His	Asp	Arg
		420						425					430		
Val	Leu	Val	Lys	Gly	Ser	Arg	Gly	Met	Lys	Leu	Glu	Glu	Val	Val	Asn
		435					440					445			
Ala	Leu	Ile	Ser												
		450													

<210> 27
 <211> 399
 <212> DNA
 <213> Homo sapiens

25

<400> 27

atgacaatga cagatccaat cgcagatatg cttactcgtg taagaaacgc aaacatgggtg 60
 cgtcacgaga agttagaatt acctgcatca aatattaaaa aagaaattgc tgaaatctta 120
 aagagtgaag gtttcattaa aaatgttgaa tacgtagaag atgataaaca aggtgtactt 180
 cgtttattct taaaatatgg tcaaaacgat gagcgtgtta tcacaggatt aaaacgtatt 240
 tcaaaaccag gtttacgtgt ttatgcaaaa gctagcgaaa tgcctaaagt attaatgggt 300
 ttaggtattg cattagtatc aacttctgaa ggtgtaatca ctgacaaaga agcaagaaaa 360
 cgtaatgttg gtggagaaat tatcgcatatc gtttggtta 399

<210> 28

<211> 132

<212> PRT

<213> Homo sapiens

<400> 28

Met Thr Met Thr Asp Pro Ile Ala Asp Met Leu Thr Arg Val Arg Asn
 1 5 10 15

Ala Asn Met Val Arg His Glu Lys Leu Glu Leu Pro Ala Ser Asn Ile
 20 25 30

Lys Lys Glu Ile Ala Glu Ile Leu Lys Ser Glu Gly Phe Ile Lys Asn
 35 40 45

Val Glu Tyr Val Glu Asp Asp Lys Gln Gly Val Leu Arg Leu Phe Leu
 50 55 60

Lys Tyr Gly Gln Asn Asp Glu Arg Val Ile Thr Gly Leu Lys Arg Ile
 65 70 75 80

Ser Lys Pro Gly Leu Arg Val Tyr Ala Lys Ala Ser Glu Met Pro Lys
 85 90 95

Val Leu Asn Gly Leu Gly Ile Ala Leu Val Ser Thr Ser Glu Gly Val
 100 105 110

Ile Thr Asp Lys Glu Ala Arg Lys Arg Asn Val Gly Gly Glu Ile Ile
 115 120 125

Ala Tyr Val Trp
 130

<210> 29

<211> 267

<212> DNA

<213> Homo sapiens

<400> 29

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 gatactgggt caccagaagt acaaatcgct gtacttactg cagaaatcaa cgcagtaaac 120
 gaacacttac gtacacacaa aaaagaccac cattcacgtc gtggattatt aaaaatggta 180
 ggtagcgtga gacatttatt aaactactta cgtagtaaag atattcaacg ttaccgtgaa 240
 ttaattaaat cacttggcat ccgtcgt 267

<210> 30

<211> 89

26

<212> PRT

<213> Homo sapiens

<400> 30

Met Ala Ile Ser Gln Glu Arg Lys Asn Glu Ile Ile Lys Glu Tyr Arg
 1 5 10 15

Val His Glu Thr Asp Thr Gly Ser Pro Glu Val Gln Ile Ala Val Leu
 20 25 30

Thr Ala Glu Ile Asn Ala Val Asn Glu His Leu Arg Thr His Lys Lys
 35 40 45

Asp His His Ser Arg Arg Gly Leu Leu Lys Met Val Gly Arg Arg Arg
 50 55 60

His Leu Leu Asn Tyr Leu Arg Ser Lys Asp Ile Gln Arg Tyr Arg Glu
 65 70 75 80

Leu Ile Lys Ser Leu Gly Ile Arg Arg
 85

<210> 31

<211> 666

<212> DNA

<213> Homo sapiens

<400> 31

taaggaggga atactgtggg tcaaaaaatt aatccaatcg gacttcgtgt tgggtattatc 60
 cgtgattggg aagctaaatg gtatgctgaa aaagacttcg cttcactttt acacgaagat 120
 ttaaaaaatcc gtaaatttat tgataatgaa ttaaaagaag catcagtttc tcaCgtagag 180
 attgaacgtg ctgcaaaccg tatcaacatt gCaattcata ctggtaaacc tgggtatggta 240
 attggtaaag gcggttcaga aatcgaaaaa ttacgcaaca aattaaatgc gtttaactgat 300
 aaaaaagtac acatcaacgt aattgaaatc aaaaaagtgt atcttgacgc tCGtttagta 360
 gctgaaaaca tcgcacgtca attagaaaac cgtgcttcat tccgtcgtgt acaaaaaaca 420
 gcaatcacta gagctatgaa acttggtgct aaaggatatca aaactcaagt atctgggtcgt 480
 ttaggcggag ctgacatcgc tcgtgctgaa caatattcag aagggaactgt tccacttcat 540
 acgttacgtg ctgacatcga ttatgcacac gctgaagctg acactactta cggtaaatta 600
 ggcgttaaag tatggattta tcgtggagaa gttcttcta ctaagaacac tagtggagga 660
 ggaaaa 666

<210> 32

<211> 217

<212> PRT

<213> Homo sapiens

<400> 32

Val Gly Gln Lys Ile Asn Pro Ile Gly Leu Arg Val Gly Ile Ile Arg
 1 5 10 15

Asp Trp Glu Ala Lys Trp Tyr Ala Glu Lys Asp Phe Ala Ser Leu Leu
 20 25 30

His Glu Asp Leu Lys Ile Arg Lys Phe Ile Asp Asn Glu Leu Lys Glu
 35 40 45

Ala Ser Val Ser His Val Glu Ile Glu Arg Ala Ala Asn Arg Ile Asn

27

50 55 60

Ile Ala Ile His Thr Gly Lys Pro Gly Met Val Ile Gly Lys Gly Gly
65 70 75 80

Ser Glu Ile Glu Lys Leu Arg Asn Lys Leu Asn Ala Leu Thr Asp Lys
85 90 95

Lys Val His Ile Asn Val Ile Glu Ile Lys Lys Val Asp Leu Asp Ala
100 105 110

Arg Leu Val Ala Glu Asn Ile Ala Arg Gln Leu Glu Asn Arg Ala Ser
115 120 125

Phe Arg Arg Val Gln Lys Gln Ala Ile Thr Arg Ala Met Lys Leu Gly
130 135 140

Ala Lys Gly Ile Lys Thr Gln Val Ser Gly Arg Leu Gly Gly Ala Asp
145 150 155 160

Ile Ala Arg Ala Glu Gln Tyr Ser Glu Gly Thr Val Pro Leu His Thr
165 170 175

Leu Arg Ala Asp Ile Asp Tyr Ala His Ala Glu Ala Asp Thr Thr Tyr
180 185 190

Gly Lys Leu Gly Val Lys Val Trp Ile Tyr Arg Gly Glu Val Leu Pro
195 200 205

Thr Lys Asn Thr Ser Gly Gly Gly Lys
210 215

<210> 33
<211> 498
<212> DNA
<213> Homo sapiens

<400> 33
atggctcgta gagaagaaga gacgaaagaa tttgaagaac gcgttggttac aatcaaccgt 60
gtagcaaaag ttgtaaaagg tggctcgctgt ttccgtttca ctgcattagt tgtagttgga 120
gacaaaaatg gtcgtgtagg ttccggtact ggtaaagctc aagaggtacc agaagcaatc 180
aaaaaagctg ttgaagcagc taaaaaagat ttagtagttg ttccacgtgt tgaaggtaca 240
actccacaca caattactgg ccgttacggg tcaggaagcg tatttatgaa accggctgca 300
cctgggtacag gagttatcgc tgggtggtcct gttcgtgccg tacttgaatt agcaggtatc 360
actgatatct taagtaaatac attaggatca aacacaccaa tcaacatggt tcgtgctaca 420
atcgatgggt tacaaaacct taaaaatgct gaagatggtg cgaaattacg tggcaaaaca 480
gtagaagaat tatacaat 498

<210> 34
<211> 166
<212> PRT
<213> Homo sapiens

<400> 34
Met Ala Arg Arg Glu Glu Glu Thr Lys Glu Phe Glu Glu Arg Val Val
1 5 10 15

28

Thr Ile Asn Arg Val Ala Lys Val Val Lys Gly Gly Arg Arg Phe Arg
20 25 30

Phe Thr Ala Leu Val Val Val Gly Asp Lys Asn Gly Arg Val Gly Phe
35 40 45

Gly Thr Gly Lys Ala Gln Glu Val Pro Glu Ala Ile Lys Lys Ala Val
50 55 60

Glu Ala Ala Lys Lys Asp Leu Val Val Val Pro Arg Val Glu Gly Thr
65 70 75 80

Thr Pro His Thr Ile Thr Gly Arg Tyr Gly Ser Gly Ser Val Phe Met
85 90 95

Lys Pro Ala Ala Pro Gly Thr Gly Val Ile Ala Gly Gly Pro Val Arg
100 105 110

Ala Val Leu Glu Leu Ala Gly Ile Thr Asp Ile Leu Ser Lys Ser Leu
115 120 125

Gly Ser Asn Thr Pro Ile Asn Met Val Arg Ala Thr Ile Asp Gly Leu
130 135 140

Gln Asn Leu Lys Asn Ala Glu Asp Val Ala Lys Leu Arg Gly Lys Thr
145 150 155 160

Val Glu Glu Leu Tyr Asn
165

<210> 35

<211> 390

<212> DNA

<213> Homo sapiens

<400> 35

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ttagtaccag gtgaaggtaa catcacagtt aataaccgtg acgtacgcga atacttacca 120
ttcgaatcat taattttaga cttaaaccac ccatttgatg taactgaaac taaaggtaac 180
tatgatgttt tagttaacgt tcatgggtggg gggtttcactg gacaagctca agctatccgt 240
cacggaatcg ctctgtcatt attagaagca gatcctgaat acagagggtc tttaaaacgc 300
gctggattac ttactcgtga cccacgtatg aaagaacata aaaaaccagg tcttaaagca 360
gctcgtcgtt cacctcaatt ctcaaaacgt 390

<210> 36

<211> 130

<212> PRT

<213> Homo sapiens

<400> 36

Met Ala Gln Val Glu Tyr Arg Gly Thr Gly Arg Arg Lys Asn Ser Val
1 5 10 15

Ala Arg Val Arg Leu Val Pro Gly Glu Gly Asn Ile Thr Val Asn Asn
20 25 30

Arg Asp Val Arg Glu Tyr Leu Pro Phe Glu Ser Leu Ile Leu Asp Leu

29

35 40 45
 Asn Gln Pro Phe Asp Val Thr Glu Thr Lys Gly Asn Tyr Asp Val Leu
 50 55 60
 Val Asn Val His Gly Gly Gly Phe Thr Gly Gln Ala Gln Ala Ile Arg
 65 70 75 80
 His Gly Ile Ala Arg Ala Leu Leu Glu Ala Asp Pro Glu Tyr Arg Gly
 85 90 95
 Ser Leu Lys Arg Ala Gly Leu Leu Thr Arg Asp Pro Arg Met Lys Glu
 100 105 110
 His Lys Lys Pro Gly Leu Lys Ala Ala Arg Arg Ser Pro Gln Phe Ser
 115 120 125
 Lys Arg
 130

<210> 37
 <211> 306
 <212> DNA
 <213> Homo sapiens

<400> 37
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 tcagcagaga agattgtaga aacagcgaaa cgttctgggtg cagatgtttc tggaccaatt 120
 ccgttaccaa ctgagaaatc agtttacaca atcatccgtg ccgtgcataa gtataaagat 180
 tcacgtgaac aattcgaaca acgtacacac aaacgtttaa tcgatattgt aaacccaaca 240
 ccaaaaacag ttgacgcttt aatgggctta aacttaccat ctggtgtaga catcgaaatc 300
 aaatta 306

<210> 38
 <211> 102
 <212> PRT
 <213> Homo sapiens

<400> 38
 Met Ala Lys Gln Lys Ile Arg Ile Arg Leu Lys Ala Tyr Asp His Arg
 1 5 10 15
 Val Ile Asp Gln Ser Ala Glu Lys Ile Val Glu Thr Ala Lys Arg Ser
 20 25 30
 Gly Ala Asp Val Ser Gly Pro Ile Pro Leu Pro Thr Glu Lys Ser Val
 35 40 45
 Tyr Thr Ile Ile Arg Ala Val His Lys Tyr Lys Asp Ser Arg Glu Gln
 50 55 60
 Phe Glu Gln Arg Thr His Lys Arg Leu Ile Asp Ile Val Asn Pro Thr
 65 70 75 80
 Pro Lys Thr Val Asp Ala Leu Met Gly Leu Asn Leu Pro Ser Gly Val
 85 90 95

Asp Ile Glu Ile Lys Leu
100

<210> 39
<211> 267
<212> DNA
<213> Homo sapiens

<400> 39
atggcctaaga aatctaaaaat agcaaaaagag agaaaaagag aagagttagt aaataaatat 60
tacgaattac gtaaagagtt aaaagcaaaa ggtgattacg aagcggttaag aaaattacca 120
agagattcat cacctacacg tttaactaga agatgtaaag taactggaag acctagaggt 180
gtattacgta aatttgaaat gtctcgtatt gcgttttagag aacatgcgca caaaggacaa 240
attccaggtg ttaaaaaatc aagttgg 267

<210> 40
<211> 89
<212> PRT
<213> Homo sapiens

<400> 40
Met Ala Lys Lys Ser Lys Ile Ala Lys Glu Arg Lys Arg Glu Glu Leu
1 5 10 15
Val Asn Lys Tyr Tyr Glu Leu Arg Lys Glu Leu Lys Ala Lys Gly Asp
20 25 30
Tyr Glu Ala Leu Arg Lys Leu Pro Arg Asp Ser Ser Pro Thr Arg Leu
35 40 45
Thr Arg Arg Cys Lys Val Thr Gly Arg Pro Arg Gly Val Leu Arg-Lys
50 55 60
Phe Glu Met Ser Arg Ile Ala Phe Arg Glu His Ala His Lys Gly Gln
65 70 75 80
Ile Pro Gly Val Lys Lys Ser Ser Trp
85

<210> 41
<211> 276
<212> DNA
<213> Homo sapiens

<400> 41
atggctcgta gtattaaaaa aggacctttc gtcgatgagc atttaaatgaa aaaagttgaa 60
gctcaagaag gaagcgaaaa gaaacaagta atcaaaacat ggtcacgctc ttctacaatt 120
ttccctaatt tcatcggaca tacttttgca gtatcgcacg gacgtaaaca cgtacctgta 180
tatgtaactg aagatatggt aggtcataaa ttaggtgagt ttgctcctac tcgtacattc 240
aaaggacacg ttgcagacga caagaaaaca agaaga 276

<210> 42
<211> 92
<212> PRT
<213> Homo sapiens

31

<400> 42

Met Ala Arg Ser Ile Lys Lys Gly Pro Phe Val Asp Glu His Leu Met
 1 5 10 15

Lys Lys Val Glu Ala Gln Glu Gly Ser Glu Lys Lys Gln Val Ile Lys
 20 25 30

Thr Trp Ser Arg Arg Ser Thr Ile Phe Pro Asn Phe Ile Gly His Thr
 35 40 45

Phe Ala Val Tyr Asp Gly Arg Lys His Val Pro Val Tyr Val Thr Glu
 50 55 60

Asp Met Val Gly His Lys Leu Gly Glu Phe Ala Pro Thr Arg Thr Phe
 65 70 75 80

Lys Gly His Val Ala Asp Asp Lys Lys Thr Arg Arg
 85 90

<210> 43

<211> 183

<212> DNA

<213> Homo sapiens

<400> 43

atggctaaaaa cttcaatggt tgctaagcaa caaaaaaac aaaaatatgc agttcgtgaa 60
 tacactcgtt gtgaacgttg tggccgtcca cattctgtat atcgtaaatt taaattatgc 120
 cgtatttgtt tccgtgaatt agcttaca aa ggccaatcc ctggcggtcg taaagctagc 180
 tgg 183

<210> 44

<211> 61

<212> PRT

<213> Homo sapiens

<400> 44

Met Ala Lys Thr Ser Met Val Ala Lys Gln Gln Lys Lys Gln Lys Tyr
 1 5 10 15

Ala Val Arg Glu Tyr Thr Arg Cys Glu Arg Cys Gly Arg Pro His Ser
 20 25 30

Val Tyr Arg Lys Phe Lys Leu Cys Arg Ile Cys Phe Arg Glu Leu Ala
 35 40 45

Tyr Lys Gly Gln Ile Pro Gly Val Arg Lys Ala Ser Trp
 50 55 60

<210> 45

<211> 699

<212> DNA

<213> Homo sapiens

<400> 45

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 aacttaaaaa aagaaggata cgatgtgtac tgtgcatacg atggtaatga tgcagtcgac 120

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ttaatttatg aagaagaacc agacatcgta ttactagata tcatgttacc tggtcgtgat 180
ggtagtgaag tatgtcgtga agtgcgcaaa aaatacgaaa tgccaataat aatgcttact 240
gctaaagatt cagaaattga taaagtgcct gggttagaac taggtgcaga tgactatgta 300
acgaaaccgt ttagtacgcg tgaattaatc gcacgtgtga aagcgaactt acgtcgtcat 360
tactcacaac cagcacaaga cactggaaat gtaacgaatg aaatcacaat taaagatatt 420
gtgatttatc cagacgcata ttctattaaa aaacgtggcg aagatattga attaacacat 480
cgtgaatttg aattgttcca ttatttatca aaacatatgg gacaagtaat gacacgtgaa 540
catttattac aaacagtatg gggctatgat tactttggcg atgtacgtac ggtcgtatgta 600
acgattcgtc gtttacgtga aaagattgaa gatgatccgt cacatcctga atatattgtg 660
acgcgtagag gcgttggata tttcctccaa caacatgag 699

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<210> 46

<211> 233

<212> PRT

<213> Homo sapiens

<400> 46

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Met Ala Arg Lys Val Val Val Val Asp Asp Glu Lys Pro Ile Ala Asp
  1             5             10             15

```

```

Ile Leu Glu Phe Asn Leu Lys Lys Glu Gly Tyr Asp Val Tyr Cys Ala
          20             25             30

```

```

Tyr Asp Gly Asn Asp Ala Val Asp Leu Ile Tyr Glu Glu Glu Pro Asp
          35             40             45

```

```

Ile Val Leu Leu Asp Ile Met Leu Pro Gly Arg Asp Gly Met Glu Val
          50             55             60

```

```

Cys Arg Glu Val Arg Lys Lys Tyr Glu Met Pro Ile Ile Met Leu Thr
          65             70             75             80

```

```

Ala Lys Asp Ser Glu Ile Asp Lys Val Leu Gly Leu Glu Leu Gly Ala
          85             90             95

```

```

Asp Asp Tyr Val Thr Lys Pro Phe Ser Thr Arg Glu Leu Ile Ala Arg
          100            105            110

```

```

Val Lys Ala Asn Leu Arg Arg His Tyr Ser Gln Pro Ala Gln Asp Thr
          115            120            125

```

```

Gly Asn Val Thr Asn Glu Ile Thr Ile Lys Asp Ile Val Ile Tyr Pro
          130            135            140

```

```

Asp Ala Tyr Ser Ile Lys Lys Arg Gly Glu Asp Ile Glu Leu Thr His
          145            150            155            160

```

```

Arg Glu Phe Glu Leu Phe His Tyr Leu Ser Lys His Met Gly Gln Val
          165            170            175

```

```

Met Thr Arg Glu His Leu Leu Gln Thr Val Trp Gly Tyr Asp Tyr Phe
          180            185            190

```

```

Gly Asp Val Arg Thr Val Asp Val Thr Ile Arg Arg Leu Arg Glu Lys
          195            200            205

```

```

Ile Glu Asp Asp Pro Ser His Pro Glu Tyr Ile Val Thr Arg Arg Gly
          210            215            220

```

Val Gly Tyr Phe Leu Gln Gln His Glu
225 230

<210> 47
<211> 937
<212> DNA
<213> Homo sapiens

<400> 47
atgccattat ttttacaacc aattttaaaa acaaaattat ggggcgggtca acgtctaagt 60
gagtttggat atcaattaga caatgataca actgggggaa tggtgggtgtg tgtcagcaca 120
tccaaatggt acgagcgaga ttattaatgg accatatcaa ggtcaaacat tagaccgtat 180
ttggtcagaa catcgtgaat tggttgggtga tttcccaagc aaagattttc cgcttctaac 240
taaaatagtg gatgcaagag aatcactttc tttcatgtg caccctgata attcttatgc 300
ttatgagcat gaaaacgggc aatatggcaa atctgaatgt tggatatatta tagatgcaga 360
agaagatgca gaaatagtta tagggacatt agcagagtct agagaagaag ttgcgaatca 420
tggttcaacac ggaacgatag agtcgatact tagatatatt aaagtaaac ctggagaatt 480
ctattttatt ccagcaggaa cagtwcatac tatttcttca ggaatattag catacgaac 540
gatgcaatcg tcagacatta catatagact ttatgatttc aatcgtcaag ataataata 600
taatgataga ccgttaaata ttgaaaagc ttttagcgtt attcagtaca atgcaccatt 660
acctaataatt ttgcctgaaa gcgaaattat tgaaaacat aagtgtacac acattgtatc 720
gaatgatttc tttacattgg ttaaatggga aatttctggc acgttaaat atatgaagcc 780
tagagagttc tgtttagtta cagtgttggga aggcgaaggg caaatgattg tctatggtga 840
aattttcaaa ctgactactg gtacaaactt tattttgact tctgaagatt tggatagtgt 900
ctttgaaggt gatttcacat tgatgattag ctatgtg 937

<210> 48
<211> 312
<212> PRT
<213> Homo sapiens

<400> 48
Met Pro Leu Phe Leu Gln Pro Ile Leu Lys Thr Lys Leu Trp Gly Gly
1 5 10 15
Gln Arg Leu Ser Glu Phe Gly Tyr Gln Leu Asp Asn Asp Thr Thr Gly
20 25 30
Glu Cys Trp Cys Val Ser Ala His Pro Asn Gly Thr Ser Glu Ile Ile
35 40 45
Asn Gly Pro Tyr Gln Gly Gln Thr Leu Asp Arg Ile Trp Ser Glu His
50 55 60
Arg Glu Leu Phe Gly Asp Phe Pro Ser Lys Asp Phe Pro Leu Leu Thr
65 70 75 80
Lys Ile Val Asp Ala Arg Glu Ser Leu Ser Ile His Val His Pro Asp
85 90 95
Asn Ser Tyr Ala Tyr Glu His Glu Asn Gly Gln Tyr Gly Lys Ser Glu
100 105 110
Cys Trp Tyr Ile Ile Asp Ala Glu Glu Asp Ala Glu Ile Val Ile Gly
115 120 125

Thr Leu Ala Glu Ser Arg Glu Glu Val Ala Asn His Val Gln His Gly
 130 135 140
 Thr Ile Glu Ser Ile Leu Arg Tyr Ile Lys Val Lys Pro Gly Glu Phe
 145 150 155 160
 Tyr Phe Ile Pro Ala Gly Thr Val His Thr Ile Ser Ser Gly Ile Leu
 165 170 175
 Ala Tyr Glu Thr Met Gln Ser Ser Asp Ile Thr Tyr Arg Leu Tyr Asp
 180 185 190
 Phe Asn Arg Gln Asp Asn Gln Tyr Asn Asp Arg Pro Leu Asn Ile Glu
 195 200 205
 Lys Ala Leu Asp Val Ile Gln Tyr Asn Ala Pro Leu Pro Asn Ile Leu
 210 215 220
 Pro Glu Ser Glu Ile Ile Glu Asn His Lys Cys Thr His Ile Val Ser
 225 230 235 240
 Asn Asp Phe Phe Thr Leu Val Lys Trp Glu Ile Ser Gly Thr Leu Asn
 245 250 255
 Tyr Met Lys Pro Arg Glu Phe Cys Leu Val Thr Val Leu Glu Gly Glu
 260 265 270
 Gly Gln Met Ile Val Asp Gly Glu Ile Phe Lys Leu Thr Thr Gly Thr
 275 280 285
 Asn Phe Ile Leu Thr Ser Glu Asp Leu Asp Ser Val Phe Glu Gly Asp
 290 295 300
 Phe Thr Leu Met Ile Ser Tyr Val
 305 310

<210> 49

<211> 837

<212> DNA

<213> Homo sapiens

<400> 49

atggctgtat tatatttagt gggcacacca attggttaatt tagcagatat tacttataga 60
 gcagttgatg tattgaaacg tgttgatatg attgcttggtg aagacactag agtaactagt 120
 aaactgtgta atcattatga tattccaact ccattaaagt catatcacga acataacaag 180
 gataagcaga ctgcttttat cattgaacag ttagaattag gtcttgacgt tgcgctcgta 240
 tctgatgctg gattgccctt aattagtgat cctggatacg aattagtagt ggcagccaga 300
 gaagctaata ttaaagtaga gactgtgcct ggacctaatg ctgggctgac ggctttgatg 360
 gctagtggat taccttcata tgtatataca tttttaggat ttttgccacg aaaagagaaa 420
 gaaaaaagtg ctgtattaga gcaacgtatg catgaaaata gcacattaat tatatacgaa 480
 tcaccgcacg gtgtgacaga tacattaaaa acaattgcaa agatagatgc aacacgacaa 540
 gtatcactag ggcgtgaatt aactaagaag ttccaacaaa ttgtaactga tgatgtaaca 600
 caattacaag cattgattca gcaaggcgat gtaccattga aaggcgaatt cgttatctta 660
 attgaagggtg ctaaagcgaa caatgagata tcgtgggttg atgatttata tatcaatgag 720
 catgttgatc atttatattca aacttcacag atgaaaccaa aacaagctat taaaaaagtt 780
 gctgaagaac gacaacttaa aacgaatgaa gtatataata tttatcatca aataagt 837

35

<210> 50

<211> 279

<212> PRT

<213> Homo sapiens

<400> 50

Met Ala Val Leu Tyr Leu Val Gly Thr Pro Ile Gly Asn Leu Ala Asp
 1 5 10 15

Ile Thr Tyr Arg Ala Val Asp Val Leu Lys Arg Val Asp Met Ile Ala
 20 25 30

Cys Glu Asp Thr Arg Val Thr Ser Lys Leu Cys Asn His Tyr Asp Ile
 35 40 45

Pro Thr Pro Leu Lys Ser Tyr His Glu His Asn Lys Asp Lys Gln Thr
 50 55 60

Ala Phe Ile Ile Glu Gln Leu Glu Leu Gly Leu Asp Val Ala Leu Val
 65 70 75 80

Ser Asp Ala Gly Leu Pro Leu Ile Ser Asp Pro Gly Tyr Glu Leu Val
 85 90 95

Val Ala Ala Arg Glu Ala Asn Ile Lys Val Glu Thr Val Pro Gly Pro
 100 105 110

Asn Ala Gly Leu Thr Ala Leu Met Ala Ser Gly Leu Pro Ser Tyr Val
 115 120 125

Tyr Thr Phe Leu Gly Phe Leu Pro Arg Lys Glu Lys Glu Lys Ser Ala
 130 135 140

Val Leu Glu Gln Arg Met His Glu Asn Ser Thr Leu Ile Ile Tyr Glu
 145 150 155 160

Ser Pro His Arg Val Thr Asp Thr Leu Lys Thr Ile Ala Lys Ile Asp
 165 170 175

Ala Thr Arg Gln Val Ser Leu Gly Arg Glu Leu Thr Lys Lys Phe Glu
 180 185 190

Gln Ile Val Thr Asp Asp Val Thr Gln Leu Gln Ala Leu Ile Gln Gln
 195 200 205

Gly Asp Val Pro Leu Lys Gly Glu Phe Val Ile Leu Ile Glu Gly Ala
 210 215 220

Lys Ala Asn Asn Glu Ile Ser Trp Phe Asp Asp Leu Ser Ile Asn Glu
 225 230 235 240

His Val Asp His Tyr Ile Gln Thr Ser Gln Met Lys Pro Lys Gln Ala
 245 250 255

Ile Lys Lys Val Ala Glu Glu Arg Gln Leu Lys Thr Asn Glu Val Tyr
 260 265 270

Asn Ile Tyr His Gln Ile Ser

275

<210> 51
 <211> 624
 <212> DNA
 <213> Homo sapiens

<400> 51
 atgaaatttg gaaaaacaat cgcagtagta ttagcatcta gtgtcttgct tgcaggatgt 60
 actacggata aaaaagaaat taaggcatat ttaaagcaag tggataaaat taaagatgat 120
 gaagaaccaa ttaaaactgt tggttaagaaa attgctgaat tagatgagaa aaagaaaaaa 180
 ttaactgaag atgtcaatag taaagataca gcagttcgcg gtaaagcagt aaaggattta 240
 attaaaaatg ccgatgatcg tctaaaggaa ttgaaaaag aagaagacgc aattaagaag 300
 tctgaacaag actttaagaa agcaaaaagt cacgttgata acattgataa tgatgttaaa 360
 cgtaaagaag taaaacaatt agatgatgta ttaaagaaaa aatataagtt acacagtgat 420
 tacgcgaaag catataaaaa ggctgtaaac tcagagaaaa cattatttaa atatttaaat 480
 caaaatgacg cgacacaaca aggtgttaac gaaaaatcaw aagcaataga acagaactat 540
 aaaaagttaa aagaagtatc agataagtat acaaaagtac taaataaggt tggtaaagaa 600
 aagcaagacg ttgatcaatt taaa 624

<210> 52
 <211> 208
 <212> PRT
 <213> Homo sapiens

<400> 52
 Met Lys Phe Gly Lys Thr Ile Ala Val Val Leu Ala Ser Ser Val Leu
 1 5 10 15
 Leu Ala Gly Cys Thr Thr Asp Lys Lys Glu Ile Lys Ala Tyr Leu Lys
 20 25 30
 Gln Val Asp Lys Ile Lys Asp Asp Glu Glu Pro Ile Lys Thr Val Gly
 35 40 45
 Lys Lys Ile Ala Glu Leu Asp Glu Lys Lys Lys Lys Leu Thr Glu Asp
 50 55 60
 Val Asn Ser Lys Asp Thr Ala Val Arg Gly Lys Ala Val Lys Asp Leu
 65 70 75 80
 Ile Lys Asn Ala Asp Asp Arg Leu Lys Glu Phe Glu Lys Glu Glu Asp
 85 90 95
 Ala Ile Lys Lys Ser Glu Gln Asp Phe Lys Lys Ala Lys Ser His Val
 100 105 110
 Asp Asn Ile Asp Asn Asp Val Lys Arg Lys Glu Val Lys Gln Leu Asp
 115 120 125
 Asp Val Leu Lys Glu Lys Tyr Lys Leu His Ser Asp Tyr Ala Lys Ala
 130 135 140
 Tyr Lys Lys Ala Val Asn Ser Glu Lys Thr Leu Phe Lys Tyr Leu Asn
 145 150 155 160
 Gln Asn Asp Ala Thr Gln Gln Gly Val Asn Glu Lys Ser Xaa Ala Ile

37

165

170

175

Glu Gln Asn Tyr Lys Lys Leu Lys Glu Val Ser Asp Lys Tyr Thr Lys
 180 185 190

Val Leu Asn Lys Val Gly Lys Glu Lys Gln Asp Val Asp Gln Phe Lys
 195 200 205

<210> 53

<211> 717

<212> DNA

<213> Homo sapiens

<400> 53

atcgaggaca gaatattggt aaagtatgaa catattgcta agcagcttaa tgcgtttata 60
 catcaatcta atttcaaacc cgggtataaa ttgccaagcg tgacgcaatt aaaagaacgt 120
 tatcaagtaa gtaagagtac tatcattaaa gcattaggct tattggaaca agatgggttg 180
 atctatcaag cacaaggcag tgggtatttat gtgagaaata ttgctgatgc caatcgtatc 240
 aacgtcttta agactaatgg tttctctaaa agtttaggtg aacaccgaat gacaagtaag 300
 gtacttggtt ttaaggagat tgcaacgccca cctaaatctg tacaagatga gctccaatta 360
 aatgcagatg ataccgtcta ctatttagag cgattaagat tcgtggacga tgatgtttta 420
 tgtatcgaat attcttatta tcataaagaa atcgtgaaat atttaaatga tgatattgct 480
 aagggctcta tcttcgacta tttagaatca aacatgaaac ttcgtattgg tttttcagat 540
 attttcttta atgtagatca actcacttca agtgaagctt cattactaca attgtctaca 600
 ggtgaaccat gtttacgtta ccaccagact ttttatacaa tgactggcaa accctttgat 660
 tcactctgaca tcgtatttca ttatcgtcat gcacagtttt atattcctag taaaaag 717

<210> 54

<211> 239

<212> PRT

<213> Homo sapiens

<400> 54

Ile Glu Asp Arg Ile Leu Leu Lys Tyr Glu His Ile Ala Lys Gln Leu
 1 5 10 15

Asn Ala Phe Ile His Gln Ser Asn Phe Lys Pro Gly Asp Lys Leu Pro
 20 25 30

Ser Val Thr Gln Leu Lys Glu Arg Tyr Gln Val Ser Lys Ser Thr Ile
 35 40 45

Ile Lys Ala Leu Gly Leu Leu Glu Gln Asp Gly Leu Ile Tyr Gln Ala
 50 55 60

Gln Gly Ser Gly Ile Tyr Val Arg Asn Ile Ala Asp Ala Asn Arg Ile
 65 70 75 80

Asn Val Phe Lys Thr Asn Gly Phe Ser Lys Ser Leu Gly Glu His Arg
 85 90 95

Met Thr Ser Lys Val Leu Val Phe Lys Glu Ile Ala Thr Pro Pro Lys
 100 105 110

38

Ser Val Gln Asp Glu Leu Gln Leu Asn Ala Asp Asp Thr Val Tyr Tyr
 115 120 125

Leu Glu Arg Leu Arg Phe Val Asp Asp Asp Val Leu Cys Ile Glu Tyr
 130 135 140

Ser Tyr Tyr His Lys Glu Ile Val Lys Tyr Leu Asn Asp Asp Ile Ala
 145 150 155 160

Lys Gly Ser Ile Phe Asp Tyr Leu Glu Ser Asn Met Lys Leu Arg Ile
 165 170 175

Gly Phe Ser Asp Ile Phe Phe Asn Val Asp Gln Leu Thr Ser Ser Glu
 180 185 190

Ala Ser Leu Leu Gln Leu Ser Thr Gly Glu Pro Cys Leu Arg Tyr His
 195 200 205

Gln Thr Phe Tyr Thr Met Thr Gly Lys Pro Phe Asp Ser Ser Asp Ile
 210 215 220

Val Phe His Tyr Arg His Ala Gln Phe Tyr Ile Pro Ser Lys Lys
 225 230 235

<210> 55
 <211> 716
 <212> DNA
 <213> Homo sapiens

<400> 55
 atgactgtag aatgggttagc agaacaatta aaagaacata atattcaatt aactgagact 60
 caaaaacaac agtttcaaac atattatcgt ttacttggtg aatggaatga aaagatgaat 120
 ttgacaagta ttacagatga acacgatgta tatttgaaac atttttatga ttccattgca 180
 cctagttttt attttgattt taatcagcct ataagtatat gtgatgtagg cgctggagct 240
 ggttttccaa gtattccgtt aaaaataatg tttccgcagt taaaagtgac gattgttgat 300
 tcattaaata agcgtattca atttttaaac catttagcgt cagaattaca attacaggat 360
 gtcagcttta tacacgatag agcagaaaca tttggtaagg gtgtctacag ggagtcttat 420
 gatgttggtta ctgcaagagc agtagctaga ttatccgtgt taagtgaatt gtgtttaccg 480
 ctagttaaaa aagggtggaca gtttggtgca ttaaaatctt caaaagggtga agaagaatta 540
 gaagaagcaa aatttgcaat tagtgtgtta ggtggtaatg ttacagaaac acataccttt 600
 gaattgccag aagatgctgg agagcgccag atgttcatta ttgataaaaa aagacagacg 660
 ccgaaaaagt atccaagaaa accagggacg ctaataagac tcctttactt gaaaaa 716

<210> 56
 <211> 239
 <212> PRT
 <213> Homo sapiens

<400> 56
 Met Thr Val Glu Trp Leu Ala Glu Gln Leu Lys Glu His Asn Ile Gln
 1 5 10 15

Leu Thr Glu Thr Gln Lys Gln Gln Phe Gln Thr Tyr Tyr Arg Leu Leu
 20 25 30

Val Glu Trp Asn Glu Lys Met Asn Leu Thr Ser Ile Thr Asp Glu His
 35 40 45

Asp Val Tyr Leu Lys His Phe Tyr Asp Ser Ile Ala Pro Ser Phe Tyr
50 55 60

Phe Asp Phe Asn Gln Pro Ile Ser Ile Cys Asp Val Gly Ala Gly Ala
65 70 75 80

Gly Phe Pro Ser Ile Pro Leu Lys Ile Met Phe Pro Gln Leu Lys Val
85 90 95

Thr Ile Val Asp Ser Leu Asn Lys Arg Ile Gln Phe Leu Asn His Leu
100 105 110

Ala Ser Glu Leu Gln Leu Gln Asp Val Ser Phe Ile His Asp Arg Ala
115 120 125

Glu Thr Phe Gly Lys Gly Val Tyr Arg Glu Ser Tyr Asp Val Val Thr
130 135 140

Ala Arg Ala Val Ala Arg Leu Ser Val Leu Ser Glu Leu Cys Leu Pro
145 150 155 160

Leu Val Lys Lys Gly Gly Gln Phe Val Ala Leu Lys Ser Ser Lys Gly
165 170 175

Glu Glu Glu Leu Glu Glu Ala Lys Phe Ala Ile Ser Val Leu Gly Gly
180 185 190

Asn Val Thr Glu Thr His Thr Phe Glu Leu Pro Glu Asp Ala Gly Glu
195 200 205

Arg Gln Met Phe Ile Ile Asp Lys Lys Arg Gln Thr Pro Lys Lys Tyr
210 215 220

Pro Arg Lys Pro Gly Thr Pro Asn Lys Thr Pro Leu Leu Glu Lys
225 230 235

<210> 57

<211> 1191

<212> DNA

<213> Homo sapiens

<400> 57

atggcacata ccattacgat tgttggctta ggaaactatg gcattgatga tttgccgcta 60
gggatatata aattttttaa gacacaagat aaagtattatg caagaacggt agatcatcca 120
gttatagaat cattgcaaga tgaattaaca tttcagagtt ttgaccatgt ttatgaagca 180
cataaccaat ttgaagatgt ctatattgat attgtggcgc aattggttga agctgcta 240
gaaaaagata ttgtctatgc ggttccgggt caccctagag ttgtcgagac aactacagt 300
aaattactgg cttagcaaaa ggacaatact gatatagatg tgaaagtttt aggtgggaaa 360
agctttattg atgatgtgtt tgaagcagtt aatgtagatc caaatgatgg cttcacactg 420
ttagatgcga catcattaca agaagtaaca cttaatgta gaacgcatac attgattacg 480
caagtttata gtgcaatgggt tgcgtctaatt ttgaaaatca cttaatgga acgatattcct 540
gatgattacc ctgttcaaat tgcactgggt gcacgaagcg atggtgcgga taacgttgtg 600
acatgcccat tatatgaatt ggatcatgat gaaaatgcat tcaataattt gacgagtgt 660
ttcgtaccaa aaatcataac atcgacatat ttgtatcatg actttgattt tgcaacggaa 720
gtgattgata ctttagttga tgaagataaa ggttgcccat gggataaagt gcaaacgc 780
gmaacgctaa agcgttattt acttgaagaa acatttgaat tggtcgaagc tattgacaat 840

gaagatgatt ggcatatgat tgaagaacta ggagatatatt tattacaagt gttattgcat 900
 actagtattg gtaaaaaaga agggatatatc gacattaaag aagtgattac aagtcttaat 960
 gctaaaatga ttcgtagaca cccacacata tttgggtgatg ccaatgctga aactatcgat 1020
 gacttaaaag aaatttggtc taaggcgaaa gatgctgaag gtaaacagcc aagagttaaa 1080
 tttgaaaaag tatttgcaga gcatttttta aatttatatg agaagacgaa ggataagtca 1140
 tttgatgagg ccgcgttaaa gcagtggcta gaaaaagggg agagtaatac a 1191

<210> 58

<211> 397

<212> PRT

<213> Homo sapiens

<400> 58

Met Ala His Thr Ile Thr Ile Val Gly Leu Gly Asn Tyr Gly Ile Asp
 1 5 10 15
 Asp Leu Pro Leu Gly Ile Tyr Lys Phe Leu Lys Thr Gln Asp Lys Val
 20 25 30
 Tyr Ala Arg Thr Leu Asp His Pro Val Ile Glu Ser Leu Gln Asp Glu
 35 40 45
 Leu Thr Phe Gln Ser Phe Asp His Val Tyr Glu Ala His Asn Gln Phe
 50 55 60
 Glu Asp Val Tyr Ile Asp Ile Val Ala Gln Leu Val Glu Ala Ala Asn
 65 70 75 80
 Glu Lys Asp Ile Val Tyr Ala Val Pro Gly His Pro Arg Val Ala Glu
 85 90 95
 Thr Thr Thr Val Lys Leu Leu Ala Leu Ala Lys Asp Asn Thr Asp Ile
 100 105 110
 Asp Val Lys Val Leu Gly Gly Lys Ser Phe Ile Asp Asp Val Phe Glu
 115 120 125
 Ala Val Asn Val Asp Pro Asn Asp Gly Phe Thr Leu Leu Asp Ala Thr
 130 135 140
 Ser Leu Gln Glu Val Thr Leu Asn Val Arg Thr His Thr Leu Ile Thr
 145 150 155 160
 Gln Val Tyr Ser Ala Met Val Ala Ala Asn Leu Lys Ile Thr Leu Met
 165 170 175
 Glu Arg Tyr Pro Asp Asp Tyr Pro Val Gln Ile Val Thr Gly Ala Arg
 180 185 190
 Ser Asp Gly Ala Asp Asn Val Val Thr Cys Pro Leu Tyr Glu Leu Asp
 195 200 205
 His Asp Glu Asn Ala Phe Asn Asn Leu Thr Ser Val Phe Val Pro Lys
 210 215 220
 Ile Ile Thr Ser Thr Tyr Leu Tyr His Asp Phe Asp Phe Ala Thr Glu
 225 230 235 240

Val Ile Asp Thr Leu Val Asp Glu Asp Lys Gly Cys Pro Trp Asp Lys
245 250 255

Val Gln Thr His Xaa Thr Leu Lys Arg Tyr Leu Leu Glu Glu Thr Phe
260 265 270

Glu Leu Phe Glu Ala Ile Asp Asn Glu Asp Asp Trp His Met Ile Glu
275 280 285

Glu Leu Gly Asp Ile Leu Leu Gln Val Leu Leu His Thr Ser Ile Gly
290 295 300

Lys Lys Glu Gly Tyr Ile Asp Ile Lys Glu Val Ile Thr Ser Leu Asn
305 310 315 320

Ala Lys Met Ile Arg Arg His Pro His Ile Phe Gly Asp Ala Asn Ala
325 330 335

Glu Thr Ile Asp Asp Leu Lys Glu Ile Trp Ser Lys Ala Lys Asp Ala
340 345 350

Glu Gly Lys Gln Pro Arg Val Lys Phe Glu Lys Val Phe Ala Glu His
355 360 365

Phe Leu Asn Leu Tyr Glu Lys Thr Lys Asp Lys Ser Phe Asp Glu Ala
370 375 380

Ala Leu Lys Gln Trp Leu Glu Lys Gly Glu Ser Asn Thr
385 390 395

<210> 59

<211> 804

<212> DNA

<213> Homo sapiens

<400> 59

aatgtaaatc attctaataa aacgacaact gtgtcttctt tacttggtata tgttacatat 60
attcacgata gagaggataa gaaaatggct caaatttcta aatataaacg tgtagttttg 120
aaactaagtg gtgaagcgtt agctggagaa aaaggatttg gcataaatcc agtaattatt 180
aaaagtgttg ctgagcaagt ggctgaagtt gctaaaatgg actgtgaaat cgcagtaatc 240
gttggtggcg gaaacatttg gagaggtaaa acaggtagtg acttaggtat ggaccgtgga 300
actgctgatt acatgggtat gcttgcaact gtaatgaatg ccttagcatt acaagatagt 360
ttagaacaat tggattgtga tacacgagta ttaacatcta ttgaaatgaa gcaagtggct 420
gaaccttata ttcgtcgtcg tgcaattaga cacttagaaa agaaacgcgt agttattttt 480
gctgcaggta ttggaaacct atacttctct acagatacta cagcggcatt acgtgctgca 540
gaagttgaag cagatgttat tttaatgggc aaaaataatg tagatggtgt atattctgca 600
gatcctaag taaacaaaga tgcggtaaaa tatgaacatt taacgcatat tcaaatgctt 660
caagaagggt tacaagtaat ggattcaaca gcacccatcat tctgtatgga taataacatt 720
ccgttaactg ttttctctat tatggaagaa ggaaatatta aacgtgctgt tatgggtgaa 780
aagataggta cgtaattac aaaa 804

<210> 60

<211> 268

<212> PRT

<213> Homo sapiens

<400> 60

42

Asn Val Asn His Ser Asn Lys Thr Thr Thr Val Ser Ser Leu Leu Val
 1 5 10 15
 Tyr Val Thr Tyr Ile His Asp Arg Glu Asp Lys Lys Met Ala Gln Ile
 20 25 30
 Ser Lys Tyr Lys Arg Val Val Leu Lys Leu Ser Gly Glu Ala Leu Ala
 35 40 45
 Gly Glu Lys Gly Phe Gly Ile Asn Pro Val Ile Ile Lys Ser Val Ala
 50 55 60
 Glu Gln Val Ala Glu Val Ala Lys Met Asp Cys Glu Ile Ala Val Ile
 65 70 75 80
 Val Gly Gly Gly Asn Ile Trp Arg Gly Lys Thr Gly Ser Asp Leu Gly
 85 90 95
 Met Asp Arg Gly Thr Ala Asp Tyr Met Gly Met Leu Ala Thr Val Met
 100 105 110
 Asn Ala Leu Ala Leu Gln Asp Ser Leu Glu Gln Leu Asp Cys Asp Thr
 115 120 125
 Arg Val Leu Thr Ser Ile Glu Met Lys Gln Val Ala Glu Pro Tyr Ile
 130 135 140
 Arg Arg Arg Ala Ile Arg His Leu Glu Lys Lys Arg Val Val Ile Phe
 145 150 155 160
 Ala Ala Gly Ile Gly Asn Pro Tyr Phe Ser Thr Asp Thr Thr Ala Ala
 165 170 175
 Leu Arg Ala Ala Glu Val Glu Ala Asp Val Ile Leu Met Gly Lys Asn
 180 185 190
 Asn Val Asp Gly Val Tyr Ser Ala Asp Pro Lys Val Asn Lys Asp Ala
 195 200 205
 Val Lys Tyr Glu His Leu Thr His Ile Gln Met Leu Gln Glu Gly Leu
 210 215 220
 Gln Val Met Asp Ser Thr Ala Ser Ser Phe Cys Met Asp Asn Asn Ile
 225 230 235 240
 Pro Leu Thr Val Phe Ser Ile Met Glu Glu Gly Asn Ile Lys Arg Ala
 245 250 255
 Val Met Gly Glu Lys Ile Gly Thr Leu Ile Thr Lys
 260 265

<210> 61

<211> 1068

<212> DNA

<213> Homo sapiens

<400> 61

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atgacaaaag aaaatatttg tatcgttttt ggagggaaaa gtgcagaaca cgaagtatcg 60
attctgacag cacaaaatgt attaaatgca atagataaag acaaatatca tggtgatatc 120
atztatatta ccaatgatgg tgattggaga aagcaaaata atattacagc tgaaattaaa 180
tctactgatg agcttcattt agaaaatgga gaggcgcttg agatttcaca gctattgaaa 240
gaaagtagtt caggacaacc atacgatgca gtattcccat tattacatgg tcctaattgg 300
gaagatggca cgattcaagg gctttttgaa gttttggatg taccatatgt aggaaatgg 360
gtattgtcag ctgcaagttc tatggacaaa cttgtaatga aacaattatt tgaacatcga 420
gggttaccac agttacctta tattagtttc ttacgttctg aatatgaaaa atatgaacat 480
aacattttaa aattagtaaa tgataaatta aattaccag tctttgttaa acctgctaac 540
ttaggggtcaa gtgtaggat cagtaaatgt aataatgaag cggaacttaa agaaggatt 600
aaagaagcat tccaatttga ccgtaagctt gttatagaac aaggcgtaa cgcacgtgaa 660
attgaagtag cagtttttagg aaatgactat cctgaagcga catggccagg tgaagtcgta 720
aaagatgtcg cgttttacga ttacaaatca aaatataaag atggtaagg tcaattacaa 780
attccagctg acttagacga agatgttcaa ttaacgctta gaaatatggc attagaggca 840
ttcaaagcga cagattgttc tggtttagtc cgtgctgatt tctttgtaac agaagacaac 900
caaatatata ttaatgaaac aaatgcaatg cctggattta cggttttcag tatgtatcca 960
aagttatggg aaaatatggg cttatcttat ccagaattga ttacaaaact tatcgagctt 1020
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```

<210> 62

<211> 356

<212> PRT

<213> Homo sapiens

<400> 62

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Met Thr Lys Glu Asn Ile Cys Ile Val Phe Gly Gly Lys Ser Ala Glu
  1             5             10             15

```

```

His Glu Val Ser Ile Leu Thr Ala Gln Asn Val Leu Asn Ala Ile Asp
      20             25             30

```

```

Lys Asp Lys Tyr His Val Asp Ile Ile Tyr Ile Thr Asn Asp Gly Asp
      35             40             45

```

```

Trp Arg Lys Gln Asn Asn Ile Thr Ala Glu Ile Lys Ser Thr Asp Glu
      50             55             60

```

```

Leu His Leu Glu Asn Gly Glu Ala Leu Glu Ile Ser Gln Leu Leu Lys
      65             70             75             80

```

```

Glu Ser Ser Ser Gly Gln Pro Tyr Asp Ala Val Phe Pro Leu Leu His
      85             90             95

```

```

Gly Pro Asn Gly Glu Asp Gly Thr Ile Gln Gly Leu Phe Glu Val Leu
      100            105            110

```

```

Asp Val Pro Tyr Val Gly Asn Gly Val Leu Ser Ala Ala Ser Ser Met
      115            120            125

```

```

Asp Lys Leu Val Met Lys Gln Leu Phe Glu His Arg Gly Leu Pro Gln
      130            135            140

```

```

Leu Pro Tyr Ile Ser Phe Leu Arg Ser Glu Tyr Glu Lys Tyr Glu His
      145            150            155            160

```

```

Asn Ile Leu Lys Leu Val Asn Asp Lys Leu Asn Tyr Pro Val Phe Val
      165            170            175

```


Lys Pro Ala Asn Leu Gly Ser Ser Val Gly Ile Ser Lys Cys Asn Asn
 180 185 190

Glu Ala Glu Leu Lys Glu Gly Ile Lys Glu Ala Phe Gln Phe Asp Arg
 195 200 205

Lys Leu Val Ile Glu Gln Gly Val Asn Ala Arg Glu Ile Glu Val Ala
 210 215 220

Val Leu Gly Asn Asp Tyr Pro Glu Ala Thr Trp Pro Gly Glu Val Val
 225 230 235 240

Lys Asp Val Ala Phe Tyr Asp Tyr Lys Ser Lys Tyr Lys Asp Gly Lys
 245 250 255

Val Gln Leu Gln Ile Pro Ala Asp Leu Asp Glu Asp Val Gln Leu Thr
 260 265 270

Leu Arg Asn Met Ala Leu Glu Ala Phe Lys Ala Thr Asp Cys Ser Gly
 275 280 285

Leu Val Arg Ala Asp Phe Phe Val Thr Glu Asp Asn Gln Ile Tyr Ile
 290 295 300

Asn Glu Thr Asn Ala Met Pro Gly Phe Thr Ala Phe Ser Met Tyr Pro
 305 310 315 320

Lys Leu Trp Glu Asn Met Gly Leu Ser Tyr Pro Glu Leu Ile Thr Lys
 325 330 335

Leu Ile Glu Leu Ala Lys Glu Arg His Gln Asp Lys Gln Lys Asn Lys
 340 345 350

Tyr Lys Ile Asp
 355

<210> 63

<211> 861

<212> DNA

<213> Homo sapiens

<400> 63

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ggaggtaaac gcatccgacc agttctgtta ttactcactt tagattcact aaataccgag 180
tatgagttag gtatgaagag cgcaattgca ctagaaatga ttcatacata ttcactttatt 240
catgatgacc taccagcgat ggataatgat gattatcgac gaggaataatt aacaaatcat 300
aaagtatatg gtgagtggac tgcgatatta gcagggtgatg ctttattaac taaagcattt 360
gaacttattt caagtgatga tagattaact gatgaagtaa aaataaaagt tctacaacgg 420
ctgtcaatag caagtggtea tgttggaaatg gtcggcggtc aaatgtaga tatgcaaagc 480
gaaggccaac caattgatct tgaaactttg gaaatgatac acaaaacaaa aacaggagca 540
ttattaactt ttgcggttat gagtgcagca gatatcgcta atgtcgatga tacaactaaa 600
gaacatttag aaagttatag ttatcattta ggtatgatgt tccagattaa agatgattta 660
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aataaaagta cgtacgtgag ttattagggt aaagatggcg cagaagataa attgacttat 780
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ttattagaaa tcgttgattt a                                     861

```

<210> 64

<211> 287

<212> PRT

<213> Homo sapiens

<400> 64

Met Thr Asn Leu Pro Met Asn Lys Leu Ile Asp Glu Val Asn Asn Glu
 1 5 10 15

Leu Ser Val Ala Ile Asn Lys Ser Val Met Asp Thr Gln Leu Glu Glu
 20 25 30

Ser Met Leu Tyr Ser Leu Asn Ala Gly Gly Lys Arg Ile Arg Pro Val
 35 40 45

Leu Leu Leu Leu Thr Leu Asp Ser Leu Asn Thr Glu Tyr Glu Leu Gly
 50 55 60

Met Lys Ser Ala Ile Ala Leu Glu Met Ile His Thr Tyr Ser Leu Ile
 65 70 75 80

His Asp Asp Leu Pro Ala Met Asp Asn Asp Asp Tyr Arg Arg Gly Lys
 85 90 95

Leu Thr Asn His Lys Val Tyr Gly Glu Trp Thr Ala Ile Leu Ala Gly
 100 105 110

Asp Ala Leu Leu Thr Lys Ala Phe Glu Leu Ile Ser Ser Asp Asp Arg
 115 120 125

Leu Thr Asp Glu Val Lys Ile Lys Val Leu Gln Arg Leu Ser Ile Ala
 130 135 140

Ser Gly His Val Gly Met Val Gly Gly Gln Met Leu Asp Met Gln Ser
 145 150 155 160

Glu Gly Gln Pro Ile Asp Leu Glu Thr Leu Glu Met Ile His Lys Thr
 165 170 175

Lys Thr Gly Ala Leu Leu Thr Phe Ala Val Met Ser Ala Ala Asp Ile
 180 185 190

Ala Asn Val Asp Asp Thr Thr Lys Glu His Leu Glu Ser Tyr Ser Tyr
 195 200 205

His Leu Gly Met Met Phe Gln Ile Lys Asp Asp Leu Leu Asp Cys Tyr
 210 215 220

Gly Asp Glu Ala Lys Leu Gly Lys Lys Val Gly Ser Asp Leu Glu Asn
 225 230 235 240

Asn Lys Ser Thr Tyr Val Ser Leu Leu Gly Lys Asp Gly Ala Glu Asp
 245 250 255

Lys Leu Thr Tyr His Arg Asp Ala Ala Val Asp Glu Leu Thr Gln Ile
 260 265 270

Asp Glu Gln Phe Asn Thr Lys His Leu Leu Glu Ile Val Asp Leu
 275 280 285

<210> 65
 <211> 819
 <212> DNA
 <213> Homo sapiens

<400> 65
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 aaaagcgaca agcgtcgagg caagttaacc atatcaaaga aatgggatca gacaactgct 180
 attttaactg ggaatttttt attggcatta ggacttgaac acttaattggc cggttaaagat 240
 aatcgtgtac atcaattgat atctgaatct atcgttgatg tttgtagagg ggaacttttc 300
 caatttcaag accaatttaa cagtcaacag acaattatta attatttacg acgtatcaat 360
 cgcaaaacag cactgttaat tcaaatatca actgaagttg gtgcaattac ttctcaatct 420
 gataaagaga ctgtacgaaa attgaaaatg attgggtcatt atataggat gagcttccaa 480
 atcattgatg atgtattaga cttcacaagt accgaaaaga aattaggtaa gccgggtcgga 540
 agtgatttgc ttaatgggtca tattacgtta ccgattttat tagaaatgcg taaaaatcca 600
 gacttcaaat tgaaaatcga acagttacgt cgtgatagtg aacgcaaaga atttgaagaa 660
 tgtatccaaa tcattagaaa atctgacagc atcgatgagg ctaaggcagt aagttcgaag 720
 tatttaagta aagcyttgaa tttgatttcy gagttaccag atggacatcc gagatcacta 780
 cytttaagtt tgacgaaaaa aatgggttca anaaacacg 819

<210> 66
 <211> 273
 <212> PRT
 <213> Homo sapiens

<400> 66
 Phe Val Ile Leu Ser Ser Gln Phe Gly Lys Asp Glu Gln Thr Ser Glu
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 Gln Thr Tyr Gln Val Ala Val Ala Leu Glu Leu Ile His Met Ala Thr
 20 25 30
 Leu Val His Asp Asp Val Ile Asp Lys Ser Asp Lys Arg Arg Gly Lys
 35 40 45
 Leu Thr Ile Ser Lys Lys Trp Asp Gln Thr Thr Ala Ile Leu Thr Gly
 50 55 60
 Asn Phe Leu Leu Ala Leu Gly Leu Glu His Leu Met Ala Val Lys Asp
 65 70 75 80
 Asn Arg Val His Gln Leu Ile Ser Glu Ser Ile Val Asp Val Cys Arg
 85 90 95
 Gly Glu Leu Phe Gln Phe Gln Asp Gln Phe Asn Ser Gln Gln Thr Ile
 100 105 110
 Ile Asn Tyr Leu Arg Arg Ile Asn Arg Lys Thr Ala Leu Leu Ile Gln
 115 120 125
 Ile Ser Thr Glu Val Gly Ala Ile Thr Ser Gln Ser Asp Lys Glu Thr
 130 135 140

47

Val Arg Lys Leu Lys Met Ile Gly His Tyr Ile Gly Met Ser Phe Gln
145 150 -155 160

Ile Ile Asp Asp Val Leu Asp Phe Thr Ser Thr Glu Lys Lys Leu Gly
165 170 175

Lys Pro Val Gly Ser Asp Leu Leu Asn Gly His Ile Thr Leu Pro Ile
180 185 190

Leu Leu Glu Met Arg Lys Asn Pro Asp Phe Lys Leu Lys Ile Glu Gln
195 200 205

Leu Arg Arg Asp Ser Glu Arg Lys Glu Phe Glu Glu Cys Ile Gln Ile
210 215 220

Ile Arg Lys Ser Asp Ser Ile Asp Glu Ala Lys Ala Val Ser Ser Lys
225 230 235 240

Tyr Leu Ser Lys Ala Leu Asn Leu Ile Ser Glu Leu Pro Asp Gly His
245 250 255

Pro Arg Ser Leu Xaa Leu Ser Leu Thr Lys Lys Met Gly Ser Xaa Asn
260 265 270

Thr

<210> 67

<211> 504

<212> DNA

<213> Homo sapiens

<400> 67

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gaagcaatta ataatgcyma agaaaagaca gctaataata ccggcttaaa attaataatt 180
gcaattaatt atgggtggcag agcagaactt gttcatagta ttaaaaaatat gtttgacgag 240
cttcatcaac aagggtttaaa tagtgataac atagatgaaa catatataaa caatcattta 300
atgacaaaag actatcctga tccagagttg ttaattcgta cttcaggaga acaaagaata 360
agtaatttct tgatttggca agtttcgtat agtgaattta tctttaatca aaaattatgg 420
cctgactttg acgaagatga attaatataa tgtataaaaa tttatcagtc acgtcaaaga 480
cgctttggcg gattgagtga ggag 504

<210> 68

<211> 168

<212> PRT

<213> Homo sapiens

<400> 68

Val Asn Tyr Ile Met Asn Leu Pro Val Asn Phe Leu Lys Thr Phe Leu
1 5 10 15

Pro Glu Leu Ile Glu Lys Asn Val Lys Val Glu Thr Ile Gly Phe Thr
20 25 30

Asp Lys Leu Pro Lys Ser Thr Ile Glu Ala Ile Asn Asn Ala Xaa Glu
35 40 45

Lys Thr Ala Asn Asn Thr Gly Leu Lys Leu-Ile Phe Ala Ile Asn Tyr
50 55 60

Gly Gly Arg Ala Glu Leu Val His Ser Ile Lys Asn Met Phe Asp Glu
65 70 75 80

Leu His Gln Gln Gly Leu Asn Ser Asp Ile Ile Asp Glu Thr Tyr Ile
85 90 95

Asn Asn His Leu Met Thr Lys Asp Tyr Pro Asp Pro Glu Leu Leu Ile
100 105 110

Arg Thr Ser Gly Glu Gln Arg Ile Ser Asn Phe Leu Ile Trp Gln Val
115 120 125

Ser Tyr Ser Glu Phe Ile Phe Asn Gln Lys Leu Trp Pro Asp Phe Asp
130 135 140

Glu Asp Glu Leu Ile Lys Cys Ile Lys Ile Tyr Gln Ser Arg Gln Arg
145 150 155 160

Arg Phe Gly Gly Leu Ser Glu Glu
165

<210> 69

<211> 1823

<212> DNA

<213> Homo sapiens

<400> 69

atgaagtggc taaaacaact acaatccctt cactactaaat ttgtaattgt ttatgtatta 60
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ctgcttgata attttaagaa gaattattacg cagtacgcga aacaattaga aattagtatt 180
gaaaaagtat atgacgaaaa gggctccgta aatgcacaaa aagatattca aaatttatta 240
agtgaagtat ccaaccgtca agaaattgga gaaattcgtt ttatagataa agaccaaatt 300
attattgcga cgacgaagca gtctaaccgt agtctaataca atcaaaaagc gaatgatagt 360
tctgtccaaa aagcactatc actaggacaa tcaaacgac atTTaatttt aaaagattat 420
ggcgggtggt aggaccgtgt ctgggtatat aatatcccag ttaaagtcga taaaaaggta 480
attggttaata tttatatcga atcaaaaatt aatgacgttt ataaccaatt aaataatata 540
aatcaaatat tcattgttgg tacagctatt tcattattaa tcacagtcac cctaggattc 600
tttatagcgc gaacgattac caaaccaatc accgatatgc gtaaccagac ggtcgaaatg 660
tccagaggta actatacgca acgtgtgaag atttatggta atgatgaaat tggcgaatta 720
gctttagcat ttaataactt gtctaaacgt gtacaagaag cgcaggctaa tactgaaagt 780
gagaaacgta gactggactc agttatcacc catatgagtg atgggtattat tgcaacagac 840
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gaagaaacag aacgaatgat tcgactggtc aatgacttgc tacagttatc taaaatggat 1320
aatgagtcgt atcaaatcaa caaagaaatt acgactttta catgttcatt aataaaaaa 1380
ttaatcgaca tgaaatgtct gcgaaagata caacatttat tcgagatatt ccgaaaaaga 1440
cgattttcac agaatttgat cctgataaaa tgacgcaagt atttgataat gtcattacaa 1500
atgcgatgaa atattctaga ggcgataaac gtgtcgagtt ccacgtgaaa caaaatccac 1560

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tttataatcg aatgacgatt cgtattaaag ataatggcat tggatttcct atcaataaag 1620
tcgataagat attcgaccga ttctatcgtg tagataaggc acgtacgcgt aaaatgggtg 1680
gtactggatt aggactagcc atttcgaaag agattgtgga agcgacacaaat ggtcgtattt 1740
gggcaaacag tgtagaaggt caaggtagat ctatctttat cacacttcca tgtgaagtca 1800
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<210> 70

<211> 608

<212> PRT

<213> Homo sapiens

<400> 70

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Met Lys Trp Leu Lys Gln Leu Gln Ser Leu His Thr Lys Phe Val Ile
  1           5           10           15
Val Tyr Val Leu Leu Ile Ile Ile Gly Met Gln Ile Ile Gly Leu Tyr
          20           25           30
Phe Thr Asn Asn Leu Glu Lys Glu Leu Leu Asp Asn Phe Lys Lys Asn
  35           40           45
Ile Thr Gln Tyr Ala Lys Gln Leu Glu Ile Ser Ile Glu Lys Val Tyr
  50           55           60
Asp Glu Lys Gly Ser Val Asn Ala Gln Lys Asp Ile Gln Asn Leu Leu
  65           70           75           80
Ser Glu Tyr Ala Asn Arg Gln Glu Ile Gly Glu Ile Arg Phe Ile Asp
          85           90           95
Lys Asp Gln Ile Ile Ile Ala Thr Thr Lys Gln Ser Asn Arg Ser Leu
          100          105          110
Ile Asn Gln Lys Ala Asn Asp Ser Ser Val Gln Lys Ala Leu Ser Leu
          115          120          125
Gly Gln Ser Asn Asp His Leu Ile Leu Lys Asp Tyr Gly Gly Gly Lys
          130          135          140
Asp Arg Val Trp Val Tyr Asn Ile Pro Val Lys Val Asp Lys Lys Val
          145          150          155          160
Ile Gly Asn Ile Tyr Ile Glu Ser Lys Ile Asn Asp Val Tyr Asn Gln
          165          170          175
Leu Asn Asn Ile Asn Gln Ile Phe Ile Val Gly Thr Ala Ile Ser Leu
          180          185          190
Leu Ile Thr Val Ile Leu Gly Phe Phe Ile Ala Arg Thr Ile Thr Lys
          195          200          205
Pro Ile Thr Asp Met Arg Asn Gln Thr Val Glu Met Ser Arg Gly Asn
          210          215          220
Tyr Thr Gln Arg Val Lys Ile Tyr Gly Asn Asp Glu Ile Gly Glu Leu
          225          230          235          240
Ala Leu Ala Phe Asn Asn Leu Ser Lys Arg Val Gln Glu Ala Gln Ala

```

50

245	250	255
Asn Thr Glu Ser Glu Lys Arg Arg Leu Asp Ser Val Ile Thr His Met		
260	265	270
Ser Asp Gly Ile Ile Ala Thr Asp Arg Arg Gly Arg Ile Arg Ile Val		
275	280	285
Asn Asp Met Ala Leu Lys Met Leu Gly Met Ala Lys Glu Asp Ile Ile		
290	295	300
Gly Tyr Tyr Met Leu Ser Val Leu Ser Leu Glu Asp Glu Phe Lys Leu		
305	310	315
Glu Glu Ile Gln Glu Asn Asn Asp Ser Phe Leu Leu Asp Leu Asn Glu		
	325	330
Glu Glu Gly Leu Ile Ala Arg Val Asn Phe Ser Thr Ile Val Gln Glu		
	340	345
Thr Gly Phe Val Thr Gly Tyr Ile Ala Val Leu His Asp Val Thr Glu		
	355	360
Gln Gln Gln Val Glu Arg Glu Arg Arg Glu Phe Val Ala Asn Val Ser		
	370	375
His Glu Leu Arg Thr Pro Leu Thr Ser Met Asn Ser Tyr Ile Glu Ala		
385	390	395
Leu Glu Glu Gly Ala Trp Lys Asp Glu Glu Leu Ala Pro Gln Phe Leu		
	405	410
Ser Val Thr Arg Glu Glu Thr Glu Arg Met Ile Arg Leu Val Asn Asp		
	420	425
Leu Leu Gln Leu Ser Lys Met Asp Asn Glu Ser Asp Gln Ile Asn Lys		
	435	440
Glu Ile Ile Asp Phe Asn Met Phe Ile Asn Lys Ile Ile Asn Arg His		
	450	455
Glu Met Ser Ala Lys Asp Thr Thr Phe Ile Arg Asp Ile Pro Lys Lys		
	465	470
Thr Ile Phe Thr Glu Phe Asp Pro Asp Lys Met Thr Gln Val Phe Asp		
	485	490
Asn Val Ile Thr Asn Ala Met Lys Tyr Ser Arg Gly Asp Lys Arg Val		
	500	505
Glu Phe His Val Lys Gln Asn Pro Leu Tyr Asn Arg Met Thr Ile Arg		
	515	520
Ile Lys Asp Asn Gly Ile Gly Ile Pro Ile Asn Lys Val Asp Lys Ile		
	530	535
Phe Asp Arg Phe Tyr Arg Val Asp Lys Ala Arg Thr Arg Lys Met Gly		
	545	550
		555
		560

Gly Thr Gly Leu Gly Leu Ala Ile Ser Lys Glu Ile Val Glu Ala His
565 570 575

Asn Gly Arg Ile Trp Ala Asn Ser Val Glu Gly Gln Gly Thr Ser Ile
580 585 590

Phe Ile Thr Leu Pro Cys Glu Val Ile Glu Asp Gly Asp Trp Asp Glu
595 600 605

<210> 71
<211> 2232
<212> DNA
<213> Homo sapiens

<400> 71

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cattctaatt gtcaagattt agtcatgaag gcaaatgaaa agtatttagt taagaatgca 180
caacaaccag aacgaggaaa gatatatgat cgtaatggta aagtgtctagc agaagatgta 240
gaaagatata aacttggtgc agtaatagat aaaaaggcga gtgccaattc taaaaaacct 300
aggcatgtag ttgataaaaa agagactgca aagaaattat ctacagtcac taatatgaag 360
ccagaggaaa ttgaaaagag acttagtcaa aagaaagctt tccaaattga atttggtgac 420
aaaggaacaa atttaacgta tcaggacaaa ttgaaaatag agaaaatgaa tttgcttgg 480
atttctttat tgcctgaaac agaacgcttt tatccaaatg gcaattttgc atcacactta 540
attggttagag ctcaaaaaaa tccggatact ggtgaactta aagggtgcat tggagttgaa 600
aagatttttg atagttattt aagtggatct aaaggatcat tgagatatat tcatgatatt 660
tggtgatata tcgcaccaaa tactaaaaaa gagaagcagc ctaaacgtgg tgatgatgtc 720
catttaacaa tcgattcaaa tattcaagta ttggttgaag aagctttaga tggcatggtt 780
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aaagtgccag atgttgaagg tcaagacaaa caaaaagcta ttgataatgt gagtgcaaaa 1860
tcattagaac cagttactat tggttctggc acacaaataa aagcacaatc tataaaagca 1920
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cctgacatgt caggatggac gaaagaagat gtcattgctt ttgaaaacct aacaaatatt 2040
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gataagtcgg ac 2232

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52

<210> 72

<211> 744

<212> PRT

<213> Homo sapiens

<400> 72

Met Ala Lys Gln Lys Ile Lys Ile Lys Lys Asn Lys Ile Gly Ala Val
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Leu Leu Val Gly Leu Phe Gly Leu Leu Phe Phe Ile Leu Val Leu Arg
 20 25 30

Ile Ser Tyr Ile Met Ile Thr Gly His Ser Asn Gly Gln Asp Leu Val
 35 40 45

Met Lys Ala Asn Glu Lys Tyr Leu Val Lys Asn Ala Gln Gln Pro Glu
 50 55 60

Arg Gly Lys Ile Tyr Asp Arg Asn Gly Lys Val Leu Ala Glu Asp Val
 65 70 75 80

Glu Arg Tyr Lys Leu Val Ala Val Ile Asp Lys Lys Ala Ser Ala Asn
 85 90 95

Ser Lys Lys Pro Arg His Val Val Asp Lys Lys Glu Thr Ala Lys Lys
 100 105 110

Leu Ser Thr Val Ile Asn Met Lys Pro Glu Glu Ile Glu Lys Arg Leu
 115 120 125

Ser Gln Lys Lys Ala Phe Gln Ile Glu Phe Gly Arg Lys Gly Thr Asn
 130 135 140

Leu Thr Tyr Gln Asp Lys Leu Lys Ile Glu Lys Met Asn Leu Pro Gly
 145 150 155 160

Ile Ser Leu Leu Pro Glu Thr Glu Arg Phe Tyr Pro Asn Gly Asn Phe
 165 170 175

Ala Ser His Leu Ile Gly Arg Ala Gln Lys Asn Pro Asp Thr Gly Glu
 180 185 190

Leu Lys Gly Ala Leu Gly Val Glu Lys Ile Phe Asp Ser Tyr Leu Ser
 195 200 205

Gly Ser Lys Gly Ser Leu Arg Tyr Ile His Asp Ile Trp Gly Tyr Ile
 210 215 220

Ala Pro Asn Thr Lys Lys Glu Lys Gln Pro Lys Arg Gly Asp Asp Val
 225 230 235 240

His Leu Thr Ile Asp Ser Asn Ile Gln Val Phe Val Glu Glu Ala Leu
 245 250 255

Asp Gly Met Val Glu Arg Tyr Gln Pro Lys Asp Leu Phe Ala Val Val
 260 265 270

Met Asp Ala Lys Thr Gly Glu Ile Leu Ala Tyr Ser Gln Arg Pro Thr
275 280 285

Phe Asn Pro Glu Thr Gly Lys Asp Phe Gly Lys Lys Trp Ala Asn Asp
290 295 300

Leu Tyr Gln Asn Thr Tyr Glu Pro Gly Ser Thr Phe Lys Ser Tyr Gly
305 310 315 320

Leu Ala Ala Ala Ile Gln Glu Gly Ala Phe Asp Pro Asp Lys Lys Tyr
325 330 335

Lys Ser Gly His Arg Asp Ile Met Gly Ser Arg Ile Ser Asp Trp Asn
340 345 350

Arg Val Gly Trp Gly Glu Ile Pro Met Ser Leu Gly Phe Thr Tyr Ser
355 360 365

Ser Asn Thr Leu Met Met His Leu Gln Asp Leu Val Gly Ala Asp Lys
370 375 380

Met Lys Ser Trp Tyr Glu Arg Phe Gly Phe Gly Lys Ser Thr Lys Gly
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Met Phe Asp Gly Glu Ala Pro Gly Gln Ile Gly Trp Ser Asn Glu Leu
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Gln Gln Lys Thr Ser Ser Phe Gly Gln Ser Thr Thr Val Thr Pro Val
420 425 430

Gln Met Leu Gln Ala Gln Ser Ala Phe Phe Asn Asp Gly Asn Met Leu
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Lys Pro Trp Phe Val Asn Ser Val Glu Asn Pro Val Ser Lys Arg Gln
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Phe Tyr Lys Gly Gln Lys Gln Ile Ala Gly Lys Pro Ile Thr Lys Asp
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Thr Ala Glu Lys Val Glu Lys Gln Leu Asp Leu Val Val Asn Ser Lys
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Lys Ser His Ala Ala Asn Tyr Arg Ile Asp Gly Tyr Glu Val Glu Gly
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Lys Thr Gly Thr Ala Gln Val Ala Ala Pro Asn Gly Gly Gly Tyr Val
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Lys Gly Pro Asn Pro Tyr Phe Val Ser Phe Met Gly Asp Ala Pro Lys
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Lys Asn Pro Lys Val Ile Val Tyr Ala Gly Met Ser Leu Ala Gln Lys
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Asn Asp Gln Glu Ala Tyr Glu Leu Gly Val Ser Lys Ala Phe Lys Pro
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Ile Met Glu Asn Thr Leu Lys Tyr Leu Asn Val Gly Lys Ser Lys Asp

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 Asp Thr Ser Asn Ala Glu Tyr Ser Lys Val Pro Asp Val Glu Gly Gln
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 Asp Lys Gln Lys Ala Ile Asp Asn Val Ser Ala Lys Ser Leu Glu Pro
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 Val Thr Ile Gly Ser Gly Thr Gln Ile Lys Ala Gln Ser Ile Lys Ala
 625 630 635 640
 Gly Asn Lys Val Leu Pro His Ser Lys Val Leu Leu Leu Thr Asp Gly
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 Asp Leu Thr Met Pro Asp Met Ser Gly Trp Thr Lys Glu Asp Val Ile
 660 665 670
 Ala Phe Glu Asn Leu Thr Asn Ile Lys Val Asn Leu Lys Gly Ser Gly
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 Phe Val Ser His Gln Ser Ile Ser Lys Gly Gln Lys Leu Thr Glu Lys
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 Asp Lys Ile Asp Val Glu Phe Ser Ser Glu Asn Val Asp Ser Asn Ser
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Asn Thr Val Gln Ser Leu Glu Ser Met Gly Phe Lys Glu Pro Thr Pro
 35 40 45

Ile Gln Lys Asp Ser Ile Pro Tyr Ala Leu Gln Gly Ile Asp Ile Leu
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Gly Gln Ala Gln Thr Gly Thr Gly Lys Thr Gly Ala Phe Gly Ile Pro
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Leu Ile Glu Lys Val Val Gly Lys Gln Gly Val Gln Ser Leu Ile Leu
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Ala Pro Thr Arg Glu Leu Ala Met Gln Val Ala Glu Gln Leu Arg Glu
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Phe Ser Arg Gly Gln Gly Val Gln Val Val Thr Val Phe Gly Gly Met
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Pro Ile Glu Arg Gln Ile Lys Ala Leu Lys Lys Gly Pro Gln Ile Val
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Val Gly Thr Pro Gly Arg Val Ile Asp His Leu Asn Arg Arg Thr Leu
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Lys Thr Asp Gly Ile His Thr Leu Ile Leu Asp Glu Ala Asp Glu Met
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Met Asn Met Gly Phe Ile Asp Asp Met Arg Phe Ile Met Asp Lys Ile
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Pro Ala Val Gln Arg Gln Thr Met Leu Phe Ser Ala Thr Met Pro Lys
 195 200 205

Ala Ile Gln Ala Leu Val Gln Gln Phe Met Lys Ser Pro Lys Ile Ile
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Lys Thr Met Asn Asn Glu Met Ser Asp Pro Gln Ile Glu Glu Phe Tyr
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 Arg Val Asp Glu Leu Thr Ser Ala Leu Ile Ser Lys Gly Tyr Lys Ala
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 Glu Gly Leu His Gly Asp Ile Thr Gln Ala Lys Arg Leu Glu Val Leu
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 Ala Ala Arg Gly Leu Asp Ile Ser Gly Val Ser His Val Tyr Asn Phe
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 Asp Ile Pro Gln Asp Thr Glu Ser Tyr Thr His Arg Ile Gly Arg Thr
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 Gly Arg Ala Gly Lys Glu Gly Ile Ala Val Thr Phe Val Asn Pro Ile
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 Glu Met Asp Tyr Ile Arg Gln Ile Glu Asp Ala Asn Gly Arg Lys Met
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 Val Asp Leu Val Ala Ala Leu Leu Gln Glu Leu Val Glu Ala Asn Asp
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 Glu Val Glu Val Gln Leu Thr Phe Glu Lys Pro Leu Ser Arg Lys Gly
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 Arg Asn Gly Lys Pro Ser Gly Ser Arg Asn Arg Asn Ser Lys Arg Gly
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 Asn Pro Lys Phe Asp Ser Lys Ser Lys Arg Ser Lys Gly Tyr Ser Ser
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 Lys Lys Lys Ser Thr Lys Lys Phe Asp Arg Lys Glu Lys Ser Ser Gly
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 Gly Ser Arg Pro Met Lys Gly Arg Thr Phe Ala Asp His Gln
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